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TAXONOMY AND DISTRIBUTION OF THE GENUS *SPARTINA*¹

David George Mobberley²

INTRODUCTION

The genus *Spartina* presents a number of taxonomic problems. Contemporary authors are not in substantial agreement in their treatment of its specific and infraspecific taxa and no one author is wholly in accord with another. Criteria heretofore proposed for separating various taxa are vague and often inconsistent, particularly for infraspecific taxa. Consequently, the accurate identification of a great number of specimens is difficult. The present investigation redefines the limits of a number of species, develops keys to the included taxa, determines geographic variability within the species and compiles evidence in support of a theory of hybrid origin for several of the species.

Recent trends in taxonomy emphasize the examination and critical evaluation of a large number of specimens of any given taxon collected throughout its geographic range. The genus *Spartina* is admirably suited to this type of investigation because most of the species are geographically distributed along a rather narrow belt of land bordering the Atlantic Ocean and adjacent waters in certain regions of both hemispheres. This feature, coupled with its abundance, often in large, dense, pure stands, makes it an attractive genus for study.

As first conceived, the scope of the present problem was limited to the species of *Spartina* occurring in North America, but several facets of the problem of nomenclature demanded an examination of specimens of South American and Old World species as well. As a consequence, all the known infrageneric taxa were included. Although only the species of eastern North America were observed under field conditions and collected in quantity, sufficient western and extra-North American material was made available by loans to allow a study of the entire genus.

HISTORICAL

The genus *Spartina* was placed in the tribe *Chlorideae* of the family *Gramineae* by Kunth (1829). Trinius (1840) considered the genus more appropriately placed in the *Agrostideae*, owing to the presence of one-flowered spikelets. Steudel (1855), following Kunth, included the genus with the *Chlorideae*, doubtless having considered the one-sided disposition of spikelets on the spikes of greater taxonomic significance than the one-flowered spikelets.

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Bentham (1881) transferred the genus to the Chamaeraphis group of the tribe Paniceae on the grounds that the spikelet contained but a single perfect floret and that the pedicel of the spikelet articulated below the glumes. Because these conditions were not found elsewhere in the Chlorideae, the placement of Spartina in this tribe was, according to Bentham, incorrect. Scribner (1883) held that Spartina was properly placed in the Chlorideae. He countered Bentham's proposal with the argument that articulation of the pedicel below the glumes seemed insufficient reason for the transfer, especially so in view of the fact that Bentham had overlooked the position of articulation in certain genera of tribes other than the Chlorideae. Most recent American and European authors have not subscribed to Bentham's disposition of the genus but are in agreement with Scribner. Hackel (1890), Parodi (1919), Bews (1929), Hitchcock (1935), Swallen (1939), Hutchinson (1944), and Chase (1950) all placed Spartina in the Chlorideae, usually placed in the subfamily Festucoideae.

Prat (1936), on the basis of epidermal cell structure and foliar anatomy, placed the Chlorideae and its included genus Spartina in the Chloridoideae subgroup of the subfamily Panicoideae.

In an abrupt departure from the more widely accepted procedure of including Spartina in the Chlorideae, Clapham, Tutin and Warburg (1952) placed the genus by itself in the tribe Spartineae which they held to be separable from the Chlorideae on the basis of its one-flowered spikelets.

Linnaeus (1753) included Dactylis cynosuroides and its variety β in the genus Dactylis. Dactylis cynosuroides is now placed in the genus Spartina as S. cynosuroides. During the latter part of the eighteenth century, several additional species were described under Dactylis by Loeffling (1758), Curtis (1787), Walter (1788), and Aiton (1789).

Schreber (1789) erected the genus Spartina but cited no species. Gmelin (1791) listed Spartina Schreb. and included a single species, S. schreberi, but did not accompany the epithet with a description. No reference was made to an earlier epithet nor was any locality given. Roth (1806) seems to have been the first to connect the spartinoid segment of Dactylis L. with Spartina Schreb.

In the early part of the nineteenth century a number of authors published short systematic lists of the species of Spartina then known. Among these were Roth (1806), Willdenow (1809), Beauvois (1812), Muhlenberg (1817), Link (1820), and Kunth (1829, 1833). Descriptions of new species appeared in several of these works. Muhlenberg's list was restricted to the species occurring in North America.

Other systematic accounts of the species were published about the same time under the genera Trachynotia (Michaux, 1803) and Limnetis (Richard in Persoon, 1807; Nuttall, 1818). The works of Michaux and Nuttall were limited to species occurring in North America.

What seems to have been the first comprehensive compilation of binomials appeared from the pen of Trinius (1840), who included six species with synonyms. Three new species were described. Subsequently, Steudel (1855) in his extensive treatment of the grasses listed twenty-three Spartina binomials, and gave descriptions for all but two of them. Many of the names had been included in the synonymy given by Trinius.

Merrill (1902) published the first monograph of the genus Spartina,

the treatment being restricted to the North American species. Nine species, one of which was new, were included. Subsequent rearrangement of species limits and nomenclatural changes by Hitchcock (1906, 1935) and Fernald (1916, 1947) have superseded many of Merrill's taxonomic conclusions.

Later systematic surveys of species of *Spartina* occurring in North America and the United States were prepared by Hitchcock (1935), Swallen (1939), and Chase (1950). These treatments list eight or nine species, provide keys to the species and append considerable synonymy. However, no evidence is offered to support the reduction of names to synonyms.

Species admitted to these systematic lists were *S. alterniflora*, *S. bakeri*, *S. cynosuroides*, *S. patens*, and *S. spartinae* of the Atlantic and Gulf coastal plains, *S. foliosa* of the coastal marshes of California, *S. pectinata* of the northern and western United States, and *S. gracilis* of the western plains and mountains. Some dispute had arisen over the disposition of *S. caespitosa* of the New England and Middle Atlantic seaboard. Hitchcock (1935) regarded it as a variety of *S. patens* while Swallen (1939) and Chase (1950) recognized it as a distinct species.

Publications dealing specifically with the South American species of *Spartina* include papers by Arechavaleta (1896) and Parodi (1919, 1937). The most comprehensive treatment was that of Parodi (1919) in which he listed five species occurring along the Atlantic seaboard in Argentina. These were *Spartina alterniflora*, *S. ciliata*, *S. densiflora*, *S. montevidensis*, and *S. patagonica*. A sixth species, *S. argentinensis*, from the inland region of Santa Fe province, Argentina, was described as new. Parodi included descriptions of the species, notes on habitats and a key.

A more recent monographic study of the genus is that of Saint-Yves (1932). The treatment is world-wide in scope. He stated that the genus was divisible into two distinct groups (*Polychlorophyllae* and *Oligochlorophyllae*) on the basis of the amount of chlorophyllous tissue within the prominent ribs on the adaxial surface of the leaf blades. The blades of the *Polychlorophyllae* were said to be flat and those of the *Oligochlorophyllae* generally involute. This division of the genus by Saint-Yves parallels the first pair of choices in the taxonomic keys of Merrill (1902 Hitchcock (1935), Swallen (1939), and Chase (1950), wherein "blades flat" or "blades involute" are used as contrasting criteria. Saint-Yves further subdivided the two groups into four species and a hybrid each, based upon features of the inflorescences. Four of the eight species he recognized, *S. maritima*, *S. cynosuroides*, *S. juncea*, and *S. densiflora*, represent combinations of a number of species of other authors, most of which were assigned by him to infraspecific ranks. The other four species, *S. arundinacea*, *S. ciliata*, *S. gracilis*, and *S. foliosa*, were not subdivided. Two hybrids, *S. cynosuroides* X *S. gracilis* and X *S. longispica*, were described. The hybrid origin of X *S. longispica* was briefly discussed.

In modern taxonomy, increasing attention is being paid to cytotaxonomic investigations. Important contributions to an understanding of cytology of the genus *Spartina* have been made by Huskins (1931) and Church (1936, 1940). Huskins confirmed the hypothesis of the hybrid origin of *S. townsendii* with cytological data. This work is reviewed more fully in connection with the discussion of *S. townsendii*.

Church (1940) investigated chromosome numbers in the North Ameri-

can species and reported the existence of a high degree of polyploidy with chromosomes present in multiples of seven. This, he concluded was evidence that seven is the basic number within the genus. Five of the nine species, S. bakeri, S. caespitosa, S. cynosuroides, S. gracilis, and S. spartinae, were found to have the hexaploid number of forty-two. These determinations were based upon a single specimen of each species. A single specimen of S. leiantha (S. foliosa) was found to have the octoploid number of fifty-six. The remaining three species, S. alterniflora, S. patens, and S. pectinata, were each shown to possess an intraspecific polyploid series. Within S. alterniflora, an octoploid and a decaploid were discovered which were observed to differ in habitat preference and morphology. Spartina patens was reported to consist of three different polyploids. Specimens from Massachusetts, Virginia, and Florida were shown to possess the tetraploid, hexaploid, and octoploid number of chromosomes, respectively. Spartina pectinata was observed to have both the hexaploid and dodecaploid number of chromosomes. New England specimens were hexaploid, while germinating seeds from Nebraska material were dodecaploid. No morphological comparisons were reported because the dodecaploids did not mature.

MATERIALS AND METHODS

The materials used in this study were largely dried and pressed specimens from a number of institutional herbaria in the western hemisphere together with the author's collections from stations within the United States. Because of the large number of specimens examined during the course of this investigation, citation of specimens has been omitted except in the cases of type specimens and certain rare instances. Where citation is made, each is followed by an appended key letter or letters which identify the institution. These key letters are the standardized abbreviations given by Lanjouw and Stafleu (1952) in *Index Herbariorum*. A complete list of herbarium specimens studied is on file in the herbarium of the Iowa State College.

Collections were made by the author during the months of July, August, and September, 1952, at selected coastwise stations ranging from Galveston, Texas eastward to Florida and northward to New Hampshire. Additional collections were made at localities in Michigan, Indiana, Wisconsin and Iowa.

Herbarium specimens were prepared from field material. The plants were dried overnight in presses suspended over kerosene lanterns. Except under the most inclement weather conditions, most of the specimens were dry enough for storage within 48 hours. However, specimens of Spartina alterniflora with their thick, fleshy, succulent culms required some additional care, often having to be left in the presses for a week or more.

Preliminary investigation had shown that some features of the inflorescences of species of Spartina possessed much measurable variability. In order to survey more adequately the ranges in variability, numerous panicles were collected and stored in small paper sacks. This procedure was followed for two reasons; first, to accumulate quantities of material

from large, pure stands in order that an adequate sampling of variability be secured, and second, to utilize a rapid and easy method of collecting small amounts of material from as many stations as possible throughout the area visited. Where the plants were found in large, dense stands, a reasonable attempt was made to get as representative a sample as possible. The plants often tend to grow in small clumps with several culms arising from the same rhizome. In no case was more than one panicle per clump selected.

In order to obtain material for cytological study of chromosome numbers, fresh root tips were killed and fixed in the field in freshly mixed Carnoy's fluid consisting of one part glacial acetic acid and two parts absolute alcohol. Only specimens to which the root tips were physically connected became a part of that particular collection. The voucher specimens from which the root tips were taken were deposited in the herbarium of Iowa State College.

Study of specimens consisted largely of measurement of various parts of the plants and critical examination of certain features not amenable to measurement. The general procedure used in dealing with all species is presented below. Occasionally, measurement or examination of a specific character was found to have some value only for a problem concerned with a single species. Methods involved in cases of this sort are described in the text. Measurement of anther lengths and morphological parts requiring accuracy to 0.1 mm were made with the aid of a Spencer low-power binocular microscope equipped with 1X, 2X, and 3X objectives and 10X field oculars. The right ocular was furnished with an ocular micrometer that had been calibrated with a stage micrometer. All other measurements were made with an ordinary 15 cm rule graduated in millimeters. Measurement data are reported in metric units.

Culm height was measured from the base of the culm to the apex of the panicle. Non-flowering culms were omitted from the study.

Measurements of leaf blade lengths and widths were made on the second blade from the summit. Certain exceptions to this procedure are noted in the text. Blade width measurements were made at a point approximately 1 cm from the base of the blade because the tissue at the base was often flanged out, cracked or broken. In the case of tightly involute blades, the tissue was softened with a detergent-methyl alcohol solution (Pohl, 1954), and flattened for more accurate determination of width.

Measurement of the ligule was made on the second leaf from the summit. In the event that lengths of the hairs varied, as they frequently did, an estimated average was struck.

Panicle length was measured from the point of insertion of the lowest spike to the tip of the terminal spike or to the tip of the panicle axis if it exceeded the spike. Only panicles with normally tightly appressed spikes appear to have a measurable width. The width of those panicles with somewhat spreading spikes seems to be influenced by degree of maturity and certainly by the pressing and mounting operations. Where width of the panicle was ascertainable, measurement was made at a point midway between base and apex.

Measurement of spikes was made from the point of insertion on the panicle axis to the tip of the terminal spikelet or to the tip of the spike

rachis if it exceeded the spikelet. Spike length was measured on a median or near median spike. Exceptions to this procedure are noted in the text. In the event that two-spiked panicles were encountered, the basal spike was measured. Occasionally the length of the peduncle proved useful. Its length was measured along the spike rachis from the point of insertion on the panicle axis to the base of the lowest spikelet.

Length of the spikelet was found to be coincident with the length of the second glume except rarely, where the floret exceeded the second glume. In such cases, spikelet length was measured to the tip of the floret. Spikelets are usually sessile but occasionally some are pedicellate. The spikelet length value in these instances included the pedicel.

The degree of crowding of spikelets along the rachis of a spike has been shown to have some taxonomic significance. A fairly satisfactory numerical expression of the crowding was derived by dividing spikelet number into spike length. This numerical equivalent seems to achieve greater precision than do the words imbricate, crowded, remote, and similar terms. However, the numerical values are not without some minor inaccuracies, for no attention was given to spikelet or peduncle lengths or variations in the lengths of distances between spikelets.

Attempts were made to determine chromosome numbers for some of the species. A smear technique was used throughout the investigation. The hand and camera lucida drawings of the chromosomes have been filed with the voucher specimens.

The data supporting taxonomic conclusions are presented in the form of descriptions, tables, histograms, maps, illustrations, and scatter diagrams. In compiling distributional data for presentation on maps, only specimens which had actually been seen and verified by the author were included. The drawings of spikelets were made with the aid of a camera lucida attached to the Spencer binocular microscope described above. Unless otherwise indicated, the drawings were originally 10X or 20X and were reduced to 3.75X or 7.5X for presentation in this paper.

Liberal use has been made of the scatter diagram allowing for the simultaneous presentation of two variables. The methods of Anderson (1949) have been followed in the construction of these diagrams. In general, the purpose in using the scatter diagram was to show whether a population sample was homogeneous or actually divisible in regard to the two variable characters utilized. In some few cases, ratios rather than absolute measurements were introduced into the diagrams.

In the systematic treatment which follows, the taxa recognized in this paper are discussed in considerable detail. Complete synonymies are given and, where nomenclatural problems warrant, evidence is presented showing why the names are regarded as synonyms. Descriptions are offered and are partially supported by illustrations of spikelets.

A discussion of the important taxonomic distinctions between the taxa is deferred to the section of this paper dealing with keys to the species.

TAXONOMIC CONSIDERATIONS

The Genus Spartina

Spartina Schreb. Gen. Pl. ed. 8. 43 (1789)

Trachynotia Michx. Fl. Bor. Amer. 1:63. (1803)

Limnethis Rich. in Pers. Syn. Pl. 1:72. (1807)

Poncelletia Thouars, Fl. Trist. Acugn. 36. (1811)

Tristania Poir. in Lam. Encycl. Suppl. IV. 526. in obs. (1816)

Solenachne Steud. Syn. Pl. Glum. 12. (1855)

Chauvinia Steud. Syn. Pl. Glum. 362. (1855)

Description

Plants perennial; culms 0.1--3.5 meters tall, erect, terete, caespitose from knotty bases or solitary from strong, scaly rhizomes; cauline leaves 3--15; sheaths firm, smooth to somewhat striate; ligule a ring of hairs; blades flat or involute; panicle 3--70 cm long; panicle axis usually 3-angled; spikes 1--75; appressed or spreading, alternately to irregularly arranged along a common axis; rachis of the spikes 3-angled; spikelets 1-flowered, laterally compressed, usually sessile in two rows on two sides of the 3-angled rachis, articulating below the glumes; first glume shorter than the floret, 1-nerved, the keel glabrous, pilose or hispid; second glume usually longer than the floret, 1--3-nerved, the keel glabrous, pilose or hispid; lemma shorter than the palea, usually firm in texture, 1--3-nerved, the lateral nerves obscure, the keel glabrous, pilose or hispid; palea thin and papery, 2-nerved, obscurely keeled, the keel glabrous or rarely hispid near the tip; stamens 3; ovary 1, stigmas 2, plumose; lodicules 0; seeds not commonly produced, plants apparently reproducing mostly by vegetative means; basic chromosome number 7.

Discussion

Spartina schreberi Gmel. has been regarded as the type species of the genus but unfortunately, the name has never enjoyed nomenclatural standing. Gmelin (1791) supplied the specific epithet but failed to provide a description, a plate or a reference to an earlier description, the name was not validly published. If a specimen upon which the name is based ever existed, its whereabouts at the present time is unknown. The invalid binomial, Spartina schreberi, is the only one, to the author's knowledge, that has ever been cited as the name of the type species. If the genus Spartina is to be properly typified, it appears necessary that another species be chosen as the type. Accordingly, the well-known and widely distributed Spartina cynosuroides (L.) Roth is designated the type species. The holotype is conserved in the herbarium of the British Museum.

Spartina, a genus of grasses, has been placed in the tribe Chlorideae, a heterogeneous assemblage of genera held together solely by the presence of a one-sided arrangement of spikelets in spikes. In this genus, the spikes appear to be one-sided although the spikelets are actually arranged in two rows on two sides of a somewhat flattened, triangular

rachis. In no other particulars of the inflorescence, except in the character linking all members of the subfamily Festucoideae does Spartina agree with the other genera referred to the Chlorideae. In the absence of more precise data concerning phylogenetic relationships, it seems advisable for the present to consider the genus a member of the Chlorideae.

Within the genus, three complexes of species may be delineated. Each of the complexes lends itself quite well to treatment as a subgenus, but owing to the small number of species, naming of subgeneric taxa does not seem advisable. The three complexes with their included species are presented in Table 1.

Table 1. Three Complexes of the Genus Spartina with Included Species.

Complex 1	Complex 2	Complex 3
<u>Spartina arundinacea</u>	<u>Spartina alterniflora</u>	<u>Spartina bakeri</u>
<u>S. ciliata</u>	<u>S. foliosa</u>	<u>S. X caespitosa</u>
<u>S. spartinae</u>	<u>S. longispica</u>	<u>S. cynosuroides</u>
	<u>S. maritima</u>	<u>S. densiflora</u>
	<u>S. neyrautii</u>	<u>S. gracilis</u>
	<u>S. townsendii</u>	<u>S. patens</u>
		<u>S. pectinata</u>

The first of the three complexes consists of those species with numerous short, closely imbricate spikes, so arranged that the entire panicle has a smooth, cylindrical contour. The culms are hard and slender. The spikelets are always pubescent, at least on the keels of the glumes, and the pubescence is strongly hispid or villous. While rhizomes may be produced, they are seldom longer than a few centimeters at most.

The second complex contains those species characterized by more or less thick, fleshy, succulent culms; smooth, glabrous leaves; and spikelets that are usually less closely imbricate than those of the species within the other complexes. The plants rarely show a tendency to be streaked or tinged with purple, though they may often become somewhat brown in color. A distinctly disagreeable odor emanating from the fresh culms may persist in dried specimens for several months or more.

Plants of the third complex are characterized by hard culms; scabrous leaves (at least on the margins); more or less spreading spikes; and usually closely imbricate spikelets. The inflorescence rarely assumes a smooth, cylindrical contour. Pubescence on the keels of glumes and lemmas is hispid. Very often plants of this complex are streaked or tinged with purple, especially in the inflorescences and occasionally on the sheaths and culms.

SYNOPSIS OF THE SPECIES

The sixteen species of the genus Spartina are treated below in synopsis form. In addition to this synopsis, keys to the species are provided in a succeeding section.

1. Panicle spikelike, smoothly cylindric; spikes numerous, short, appressed, often twisted; culms slender, hard, not fleshy nor succulent; rhizomes short or wanting; keels of the glumes hispid or villous. 2
1. Panicle not commonly spikelike (except in S. foliosa); spikes appressed or spreading; culms thick, fleshy, succulent, having a disagreeably sulphurous odor when fresh; rhizomes usually flaccid, scales inflated; pubescence on the spikelets usually pilose; plants rarely streaked or tinged with purple (observed only in a single specimen of S. foliosa). 3
1. Panicle not spikelike; spikes usually spreading; culms hard, not fleshy nor succulent; rhizomes when present firm, scales not inflated; pubescence on the spikelets hispid to pectinate; plants often streaked or tinged with purple. 6
2. Spikelets lanceolate, exceeding 8 mm in length; plants of the islands of Tristan de Cunha, Diego Alvarez, and St. Paul and Amsterdam. 1. Spartina arundinacea
2. Spikelets ovate, densely pilose, exceeding 8 mm in length; plants of the sandy beaches and dunes of southern Brazil, Uruguay and northern Argentina. 2. Spartina ciliata
2. Spikelets lanceolate, not usually exceeding 8 mm in length; plants distributed from Florida to Costa Rica along the Gulf of Mexico and Caribbean Sea and in south-central Paraguay and northern Argentina. 3. Spartina spartinae
3. Spikes usually short, closely appressed, crowded near the apex of the panicle, often twisted; spikelets glabrous to pilose; glumes usually curved; plants of California and Baja California. 5. Spartina foliosa
3. Spikes elongate, not usually closely appressed; not noticeably crowded near the apex of the panicle, never twisted; spikelets glabrous to pilose; glumes not curved; plants distributed elsewhere. 4
4. Spike rachis exceeding 1 mm in width; spikelets usually appressed, moderately imbricate, not copiously pilose; plants of the River Plate region of Argentina and Uruguay. 6. Spartina longispica

4. Spike rachis not usually exceeding 1 mm in width; spikelets not usually appressed, remotely to closely imbricate, not copiously pilose; plants of coastal marshes of North America, South America and Europe. 4. Spartina alterniflora
4. Spike rachis more or less than 1 mm in width; spikelets usually copiously pilose, appressed or spreading, moderately imbricate; plants of Europe and Africa. 5
5. Blades exceeding 17 cm in length, not readily disarticulating from the sheath; spike rachis exceeding 1 mm in width; spikelets appressed; plants of southwestern France. 8. Spartina neyrautii
5. Blades not exceeding 12 cm in length, readily disarticulating from the sheath; spike rachis not usually exceeding 1 mm in width; spikelets appressed or spreading, not exceeding 15 mm in length; plants of the coastal marshes of western Europe and Africa. 7. Spartina maritima
5. Blades exceeding 12 cm in length, not usually disarticulating from the sheath; spike rachis usually exceeding 1 mm in width; spikelets usually spreading, exceeding 15 mm in length, plants of the coastal marshes of western Europe. 9. Spartina townsendii
6. Rhizomes wanting or short and at once upturned; culms caespitose; extra-vaginal shoots closely appressed to the culm; spikes usually fewer than 15. 7
6. Rhizomes present; culms usually solitary; spikes few to many. 8
7. Second glume 1-nerved, tip usually acute, culms not exceeding 1.5 meters in height; plants of Brazil, Uruguay, Argentina, and Chile. 13. Spartina densiflora
7. Second glume 2--3-nerved, tip obtuse to acuminate, culms usually exceeding 1.5 meters in height; plants of salt or fresh water habitats in Florida and Georgia. 14. Spartina bakeri
7. Second glume 3-nerved, tip acuminate to aristate; culms not exceeding 1.3 meters in height; plants of disturbed ground in and around coastal marshes from Maine to Maryland. 11. Spartina X caespitosa
8. Rhizomes thick and hard, light brown to brownish-purple, scales closely imbricate; blades usually flat; spikes few to many. 9
8. Rhizomes thin and wiry, whitish, scales not closely imbricate; blades usually involute; spikes fewer than 15. 10
9. Second glume not awned; first glume usually shorter than one-half the length of the second; lemma not deeply bilobed at the

tip, tip acute to obtuse; plants of the eastern seaboard from Massachusetts to Florida and along the coast of the Gulf of Mexico from Florida to Texas. . . . 12. Spartina cynosuroides

9. Second glume aristate to awned; first glume usually longer than one-half the length of the second; lemma deeply bilobed, tip acuminate. 10

10. Culms usually exceeding 1.2 meters in height; spikes commonly more than 10; second glume usually long-awned; plants of Nova Scotia and New Brunswick west to Alberta, south to Virginia, Kentucky, Texas, and Washington. 16. Spartina pectinata

10. Culms not usually exceeding 1.2 meters in height; spikes fewer than 10; second glume acuminate to aristate; plants of disturbed ground in and around coastal marshes from Maine to Maryland. 11. Spartina X caespitosa

11. Spikes slender, not usually closely appressed to the panicle axis; pubescence on keels of glumes and lemmas hispid; plants of eastern North America along the Atlantic seaboard from Nova Scotia to Florida, inland in Michigan and New York, Gulf and Caribbean coasts from Florida to Honduras, throughout the West Indies, and in Europe in the western Mediterranean basin. 15. Spartina patens

11. Spikes short, broad, closely appressed to the culms; pubescence on keels of glumes and lemmas copiously long-hispid; plants of plains and mountain regions of North America from Mexico to the Mackenzie Territory. 14. Spartina gracilis

1. SPARTINA ARUNDINACEA

Spartina arundinacea (Thouars) Carmich. Trans. Linn. Soc. 12:504. (1819)

Ponceletia arundinacea Thouars, Fl. Trist. Acugn. 36. (1811)
(Type in P from Tristan de Cunha)

Tristania arundinacea (Thouars) Poir. in Lam. Encycl. Suppl. IV:526. (1816) in obs.

Description (Plate 1, a-c)

Culms to 1.5 meters tall, erect, terete, indurate, striate, glabrous, from knotty bases; rhizomes short, thick, upturned; sheaths indurate, often shining, smooth to somewhat striate, margin of the orifice pilose; ligule a ring of hairs about 1 mm in length; blades involute, glabrous to scabrous on the adaxial surface, glabrous on the abaxial, scabrous on the margins, tip acuminate, the terminal blade not exceeding the panicle; panicle up to 16 cm long, 1--2 cm wide; panicle axis 3-angled, glabrous

to sparingly hispid on the angles, spikes up to 50, 2--4 cm long, 2--5 mm wide, flattened dorsiventrally, irregularly arranged, tightly appressed, closely imbricate, twisted, giving the panicle a smooth contour; rachis of the spikes glabrous to sparingly hispid on the angles not prolonged beyond the terminal spikelet; spikelets 20--40, 6--12 mm long, 1--3 mm wide, alternate, tightly appressed, closely imbricate, often pedicellate, the pedicel about 1 mm long; first glume linear, 4--10 mm long, glabrous, keel hispid, tip acuminate; second glume 6--12 mm long, glabrous to minutely hispidulous, keel hispid above, margins glabrous to hispidulous, tip acuminate; lemma glabrous to minutely hispidulous, keel hispid above, often glabrous below, margins glabrous, tip obtuse, shallowly lobed; palea thin and papery, margins notched at the apex of each nerve, tip obtuse or rounded, slightly exceeding the lemma; anthers rarely exceeding 3 mm in length; stigmas usually subequal.

Observations

As only four specimens of this species were available for examination, it is impossible to show ranges in the variability of morphological features. The descriptive material above was taken from the specimens examined. Admittedly, these constitute a rather unsatisfactory sample, all the more so because of the scattered distribution of the species. Chromosome numbers are unknown for this species.

Nomenclature

The earliest description of the plant is that of Du Petit-Thouars (1811) under the genus Ponceletia. Although the type specimen was not seen, study of the detailed description shows that Spartina arundinacea is properly typified. The combination S. arundinacea Carmich. has been used by other authors, but inasmuch as the name-bringing synonym was cited in synonymy by Carmichael (1819) it appears that the author citation should include the original authority for the epithet.

Distribution

Of the four specimens examined, two were from the island of Tristan de Cunha (37°06' S., 12°17' W.) and another from the islands of St. Paul and Amsterdam (38°43' S., 77°32' E.). The fourth is merely labeled St. Paul Island near Tristan de Cunha. This specimen, from the Torrey Herbarium (NY), was in all probability collected on St. Paul Island. No apparent morphological differences exist between specimens from the two widely separated island groups.

Hemsley (1844) reports the plant to be common on all three of the islands which make up the Tristan de Cunha group as well as on the two islands, St. Paul and Amsterdam. Christopherson (1934) found the plant occurring abundantly on Gough Island (Diego Alvarez) 220 miles south-east of Tristan de Cunha.

Little precise data are available concerning the habitat of the species. Carmichael (1819) states that the plants grow abundantly in thick clumps at the base of the volcanic mountain on Tristan de Cunha. The species has been observed to flower in February.

Specimens examined. St. Paul and Amsterdam, Nauman 112 US; St. Paul near Tristan de Cunha, NY; Tristan de Cunha, MacGillivray 345 GH; Stableford ISC.

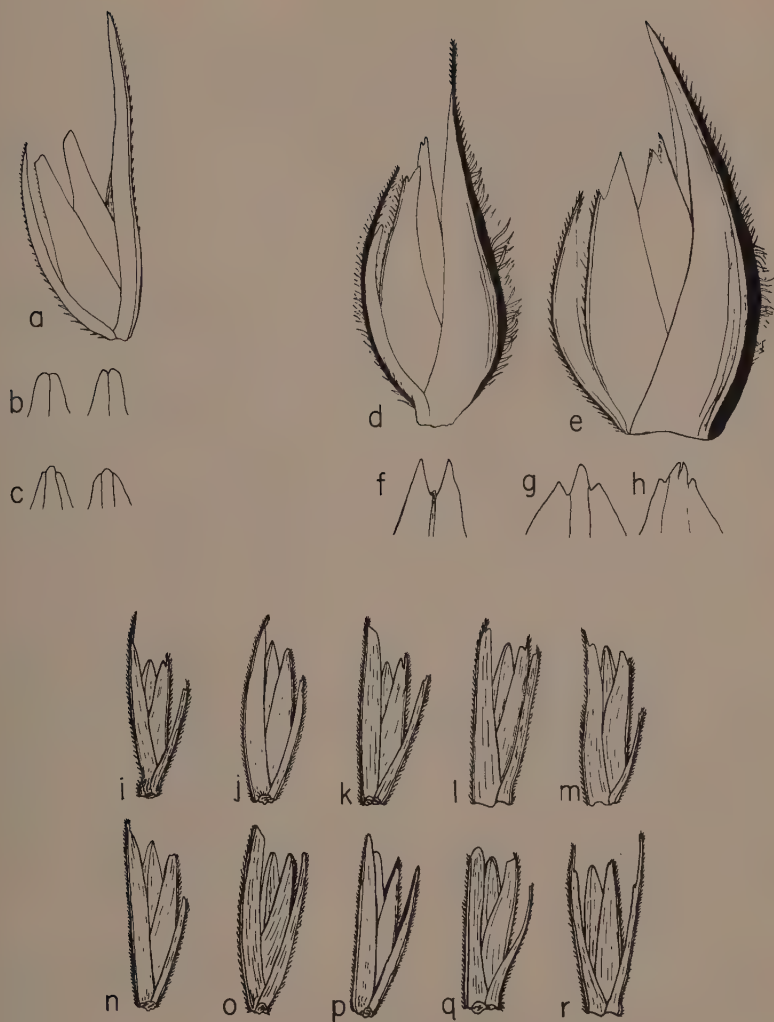


PLATE I

a-c. *Spartina arundinacea*.

a. spikelet 3.75X, b. lemma tips 7.5X, c. palea tips 7.5X.

d-h. *Spartina ciliata*.

d-e. spikelets 3.75X, f. lemma tip 7.5X. g-h. palea tips 7.5X.

i-r. *Spartina spartinae*.

i-r. variations in spikelets 3.75X.

2. SPARTINA CILIATA

Spartina ciliata Brong. Du Perr. Voy. Coq. Bot. 15. (1829)

(Type in P from Isle of St. Catherine, Brazil)

Spartina ciliata Kunth. Rev. Gram. 1:94. (1829) nomen nudum.

Spartina coarctata Trin. Mem. Acad. St. Petrsbg. Sci. Nat. VI.

6:110. (1836) (Type in LE from Uruguay)

Solenachne phalaroides Steud. Syn. Pl. Glum. 12. (1855)

(Type in P from Isle of St. Catherine, Brazil)

Description (Plate 1, d-h)

Culms to 2 meters tall, erect, indurate, terete, glabrous, caespitose from hard, knotty bases, occasionally with short rhizomes; sterile leaf-bearing shoots often arising from the lower nodes, these tightly appressed to the culm; sheaths smooth, glaucous or shining, the lower much broken by the emergent shoots, margin glabrous, hyaline, margin of the orifice densely pilose; ligule a ring of hairs 2--3 mm long; blades narrowly involute, scabrous on the adaxial surface, glabrous on the abaxial, glabrous to scabrous on the margins, tip acuminate; panicle 15--40 cm long, 1--2 cm wide; panicle axis 3--4-angled, twisted, sparingly pilose; spikes 10--50, 2--5 cm long, flattened dorsiventrally, irregularly arranged, tightly appressed, densely imbricate, twisted giving the panicle a smooth contour; rachis of the spikes pilose on the angles, not prolonged beyond the terminal spikelet; spikelets 15--25, 10--20 mm long, 4--8 mm wide, alternate, tightly appressed, closely imbricate, often twisted; first glume linear, 6--10 mm long, thin and papery, glabrous to sparingly pilose, keel pilose below, becoming strongly hispid above, margins often hispidulous; second glume 12--20 mm long, thin and chaffy, glabrous to sparingly pilose, keel broad and thick, densely pilose below, becoming strongly hispid above, tapering into a short, sharp-pointed awn, margins hispidulous; lemma glabrous, the upper two-thirds of the keel pilose, margins hispidulous, produced beyond the keel forming a sharply bifid apex; palea thin and membranaceous, margins notched at the apex of each nerve, tip obtuse or rounded, slightly exceeding the lemma; anthers 4--7 mm long, often purple.

Observations

Specimens of Spartina ciliata are essentially uniform throughout the geographic range, although some variation does occur in height of the culm and length of the panicle. These differences are, in all probability, due to environmental circumstances. None of the variations could be correlated with geographic distribution. Chromosome numbers in this species are unknown.

Nomenclature

The binomial Spartina ciliata has been attributed to both Brongniart (1829) and Kunth (1829). The valid name is Brongniart's, as Kunth's was not accompanied by a description until a later publication (Kunth, 1833). The type specimen was not seen but a fragment deposited in the U.S. National Herbarium was available for study.

Distribution

The species is limited to the east coast of South America, ranging from south-central Brazil to Argentina.

It grows in dense clumps on sandy beaches and dunes well above the intertidal zone. Parodi (1919) included a photograph showing the typical habitat on the sandy coastal areas. The species flowers from November to May.

3. SPARTINA SPARTINAE

Spartina spartinae (Trin.) Hitchc. Contr. U.S. Nat. Herb. 17:329. (1913)

Vilfa spartinae Trin. Mem. Acad. St. Petrsbg. Sci. Nat. VI. 6:82. (1840) (Type in LE from Texas)

Spartina junciformis Eng. and Gray. Bost. Jour. Nat. Hist. 5:238. (1847) (Type in GH from Texas)

Spartina gracilis Hook. ex Chap. Fl. So. U.S. ed. 1. 556. (1860) non Trin. (1840)

Spartina gouni Fourn. Mex. Pl. 2:135 (1886) (Type in P from Mexico)

Spartina multiflora Vasey ex Beal, Grasses No. Amer. 2:400 (1896) pro syn. (Type in US from Florida)

Spartina pittieri Hack. Oesterr. Bot. Ztschr. 52:237. (1902) (Type in W from Costa Rica)

Spartina densiflora var. obtusa Hack. Fedde Rep. Nov. Spec. 6:345. (1909) (Type in W from Paraguay)

Spartina argentinensis Par. Rev. Fac. Agron. Vet. Buenos Aires 2:258. (1919) (Type from Argentina)

Spartina densiflora Brong. var. junciformis (Eng. and Gray) St. Y. Cand. 5:77. (1932) (Based on S. junciformis Eng. and Gray)

Spartina densiflora Brong. var. junciformis (Eng. and Gray) St. Y. subv. argentinensis (Par.) St. Y. Cand. 5:79. (1932) (Based on S. argentinensis Par.)

Spartina densiflora Brong. var. junciformis (Eng. and Gray) St. Y. subv. genuina St. Y. Cand. 5:78. (1932) (Based on S. junciformis Eng. and Gray)

Spartina densiflora Brong. auct. non Beal, W. J. Grasses No. Amer. 2:396. (1896)

Description (Plate 1, i-r)

Culms to 2 meters tall, erect, terete, indurate, commonly glabrous, occasionally glaucous, from hard, knotty bases, rhizomes wanting; nodes often exposed; sheaths short, glabrous, occasionally glaucous, sparingly scabrous on the margins, margin of the orifice glabrous to hispid, never pilose; ligule a ring of hairs 1--2 mm long; blades involute when fresh, up to 5 mm wide, scabrous on the adaxial surface, glabrous on the abaxial, scabrous on the margins, tip acuminate; panicle 6--70 cm long, 4--20 mm wide; panicle axis 3--4-angled, often scabrous on the margins, twisted; spikes 6--75, 0.5--7 cm long, the lower often the longer, 2--4 mm wide, alternately to irregularly arranged, tightly appressed, closely imbricate, flattened dorsiventrally; rachis of the spikes glabrous to scabrous on the angles; spikelets 10--60, 4--8(-10) mm long, the lower

often the longer; first glume linear, 2--6(-8) mm long, glabrous to hispidulous, keel hispid, tip acuminate; second glume 4--8(-10) mm long, glabrous to hispidulous, keel hispid, tip acuminate to obtuse; lemma glabrous to hispidulous, upper two-thirds of the keel hispid, tip usually acuminate, rarely obtuse; palea thin and papery, often minutely hispid near the tip, tip acuminate to obtuse; anthers 3--5 mm long, occasionally purple; somatic chromosome numbers 28, 42.

Observations

Certain vegetative characters of *Spartina spartinae* show measurable variability. Data were obtained from specimens from across the entire range of geographic distribution of the species (Table 2). Mean and extreme values for height of the culm, length of the blade of the second leaf from the apex and width of the blade were recorded by geographic areas. Plants of the Yucatan Peninsula are included with those from Central America and Mississippi specimens are included with those of Texas and Louisiana.

Table 2. Morphological Characters of *S. spartinae*
Based on Observations of 119 Specimens.

		Height of culm (cm)	Blade length (cm)	Blade width (mm)	Panicle length (cm)	Spike length (cm)
Florida	extreme	38.0	26.0	2.0	7.0	1.0
	mean	89.5	33.3	2.6	17.0	2.3
	extreme	184.0	54.0	3.5	45.0	3.4
Texas and Louisiana	extreme	49.0	21.0	1.5	8.0	1.1
	mean	93.8	37.1	2.8	17.3	2.3
	extreme	200.0	58.0	4.1	38.0	3.5
Mexico	extreme	46.0	12.0	1.5	8.0	1.5
	mean	104.3	28.2	3.0	20.1	2.4
	extreme	140.0	44.0	4.5	35.0	3.6
Central America	extreme	120.0	42.0	2.9	20.0	1.4
	mean	161.1	82.7	3.4	42.1	2.8
	extreme	195.0	125.0	4.5	61.0	4.5
South America	extreme	78.0	22.0	1.5	14.5	2.0
	mean	119.4	32.6	3.0	25.6	2.9
	extreme	166.0	51.0	4.0	39.0	4.2
Total	extreme	38.0	12.0	1.5	7.0	1.0
	mean	105.7	39.7	2.9	21.6	2.4
	extreme	200.0	125.0	4.0	61.0	4.5

The mean value for culm height in the Central American collections is considerably higher than the mean value for the total population. Because the measurement data from these Central American collections were taken largely from specimens collected near Belize, British Honduras, it may be that the results do not accurately show the variability within the Central American segment of the population. Corroborative evidence supporting this assumption is provided by Swallen (1932) who states that specimens of *S. spartinae* from the Mexican portion of the Yucatan Peninsula do not exceed 150 cm in height. Although taller plants are apparently more common in parts of Central America, there is, in the author's collections from Cameron Parish, La., a specimen which is just 200 cm tall (Mobberley 59, ISC).

As in the case of the culm, the mean value of blade length in the Central American segment is much higher than that in other segments. However, the mean blade width value from this segment does not vary a great deal from the mean value for the total population sample.

Beal (1896) used the presence of exposed nodes as a vegetative character of taxonomic significance. Although this feature was observed in many of the specimens examined, its occurrence was not consistent enough in any one segment to warrant attaching any diagnostic value to it.

The general aspect of the panicle seems to show some uniformity throughout the population. The spikes are usually densely imbricate and flattened dorsiventrally. The entire panicle is given a smooth contour by the twisting of these flattened spikes. Occasionally the panicles become dense and heavy enough to nod. Some variation was observed in the lengths of panicles and spikes (Table 2). Higher mean values for panicle and spike length appear in the Central American material, but for the reason discussed above in connection with culm height these higher values are not thought to be significant.

Considerable variation was observed in the morphology of the spikelet, notably at the tip of the second glume. Plate 1, i--r, illustrates the range of variation which was encountered. The illustrations are not intended to portray all the existing variations, but are drawn from selected spikelets which are more or less representative of the entire population. An effort was made to learn whether the variability in the character of the tip is correlated with geographic distribution, but the various types were not limited to one particular geographic area, clone or panicle. However, some uniformity was found within individual spikes.

Relative lengths of first and second glumes have been found to be of diagnostic value in certain of the Gramineae. Appraisal of this feature in *S. spartinae* revealed that the second glume is about 2 mm longer than the first. In rare cases, spikelets were found in which the length of the first glume barely equalled one-half the length of the second; in one instance (Plate 1, r) the first glume exceeded the second in length.

The anthers show no particular features of diagnostic value. Occasionally purple anthers are found, but the majority are yellowish.

Among other morphological features which possess some taxonomic usefulness are the absence of rhizomes, the caespitose habit, numerous and closely imbricate spikes, length of the spikelets and pubescence on the spikelets. These features will be discussed in greater detail in another section of this paper dealing with the distinctions between species.

Chromosome numbers for S. spartinae have been reported from only one previous source (Church, 1940). His determination of the hexaploid number of 42 is not in agreement with this investigators findings in which the tetraploid number of 28 was discovered (Table 3). Church's report is based upon Florida material (Church 510, Miami, Fla.), while the author's determinations are based on Texas and Louisiana specimens.

Table 3. Chromosome Numbers in S. spartinae

Specimen		Chromosome number
Mobberley 26	Port Bolivar, Galveston Co., Texas	2n = 28
Mobberley 32	Sabine Pass, Jefferson Co., Texas	2n = 28
Mobberley 33	Sabine Pass, Jefferson Co., Texas	2n = 28
Mobberley 40	Sabine, Jefferson Co., Texas	2n = 28
Mobberley 41	Sabine, Jefferson Co., Texas	2n = 28
Mobberley 52	Hackberry, Cameron Parish, La.	2n = 28

Conditions found elsewhere in the genus indicate that polyploidy is more or less to be expected. Accordingly, Florida specimens were critically compared with specimens from Texas and Louisiana in an effort to learn whether the apparent cytologically differing segments of the population differed in any gross morphological features. No significant differences were uncovered in any of the traits compared.

Nomenclature

The name Spartina spartinae (Trin.) Merr. has been widely used despite its erroneous author citation. Merrill (1902) introduced the combination by transfer of Vilfa spartinae Trin., to Spartina but rejected it as meaningless. Hitchcock (1913) adopted the invalidly published combination and cited Merrill as the authority. There being no obstacle to the use of the combination, it may continue to be employed. However, under the provisions of Article 81 of the International Code of Botanical Nomenclature, the epithet is to be regarded as new under the authority of Hitchcock. The binomial with the proper author citation becomes Spartina spartinae (Trin.) Hitchc.¹

A photograph of the type specimen of Vilfa spartinae was examined. This examination, coupled with a study of a fragment of the type (deposited in US), leaves no doubt as to the accuracy of the typification.

Spartina argentinensis was described by Parodi (1919). Although the type specimen was not available for examination, nothing appears in the description that would indicate that it is different from S. spartinae. Spikelet lengths are given as 5--7 mm, spikes 20--40 in number and

¹The author is indebted to Dr. H.W. Rickett, bibliographer to the New York Botanical Garden, for assistance in the solution of this nomenclatural problem.

2--3 cm in length, but these values are met throughout the geographic range of S. spartinae. As may be seen from Parodi's published photograph of the type specimen (Parodi, 1919), the spikes seem somewhat less tightly appressed. However, several specimens with spikes tightly appressed, identified by Parodi as S. argentinensis, are deposited in the U.S. National Herbarium. It is apparent that Parodi did not regard this more or less lax attitude of the spikes as of fundamental taxonomic significance. In addition, this feature was observed occasionally among specimens from Texas and Mexico. Because nothing except geographic discontinuity allows a separation of the South American from the North American plants, it seems advisable to regard S. argentinensis as a synonym of S. spartinae.

Saint-Yves (1932) reduced S. spartinae to S. densiflora var. junciformis and erected within that variety two subvarieties, genuina and argentinensis, differentiated on the basis of geographic distribution, panicle length and length and appression of the spikes. The former subvariety is supposed to occur in North and Central America and have panicles 20--30 cm long and tightly appressed spikes 2--2.5 cm long. The latter subvariety is supposedly characterized by its occurrence in South America, panicles 30--35 cm long, and somewhat less tightly appressed spikes 3--4 cm long. This investigation has shown that S. spartinae (S. junciformis) is a species quite distinct from S. densiflora and that their combination by Saint-Yves is unwarranted. Effective separation of the two is discussed in another section of this paper dealing with distinctions between South American species of the genus. All available specimens of S. spartinae were evaluated in accordance with the criteria proposed by Saint-Yves in order to learn whether or not these subvarietal taxa may be distinguished from each other. Some of the South American specimens do have the spikes less tightly appressed but, as noted above in the discussion of S. argentinensis, the same condition was observed in North and Central American material. That the criteria concerning panicle and spike lengths are not answered is evident from the data presented in Table 2. Panicle and spike lengths of many North American specimens are in excess of the stipulated upper limits for the South American subvariety. Panicle lengths of many South American specimens are less than the stipulated lower limits for the North American subvariety. The least spike length value found among South American specimens is just equal to the stipulated lower limit for the North American subvariety. The foregoing evidence indicates that these subvarietal taxa have been erected without justification and, accordingly, they are treated as synonyms of S. spartinae.

Distribution

The distribution of Spartina spartinae is limited to the western hemisphere and consists of two disjunct geographic segments. In North and Central America the species ranges from Florida around the gulf coast to Texas, Mexico, and Costa Rica. A single specimen (Bailey 1011, US) is known from the Bahamas, B.W.I. Fig. 1 shows the distribution of the North and Central American segment of the population.

In South America, the species appears to have a more restricted distribution limited to Northern Argentina and Paraguay.



Fig.1. Distribution of *Spartina spartinae* in North America

A condition which might bear further investigation is the inland distribution of the South American representatives. In South America, the species is apparently not known from the coastal areas. This is somewhat in contrast to the North and Central American situation where plants are found on the sandy beaches as well as some distance inland. Some Texas collections indicate that the plant may be found inland as much as 60 miles in the region of Gonzales County. Mexican collections show that the species occurs both coastwise and inland, often at considerable altitude. Comparison of the specimens from both areas shows no discrete morphological differences. The upland specimens tend to be a little less robust and often possess fewer spikes than do the coastal plants, but otherwise they are similar.

At collection stations in southeastern Texas and southern Louisiana, the author found the species to be very common. The plants grow in a variety of habitats from the sandy beaches, dunes, roadsides, roadside ditches and wet meadows to arid fields and pastures. In fields and pastures near the coast, the species is easily the most prominent if not the most abundant component of the vegetation. It has a rather striking appearance by virtue of the large, tall clumps (up to 70 cm in diameter) which seem to overtop the remainder of the vegetation. The clumps may have as many as forty or fifty fertile culms. Clumps on the beaches and dunes are usually somewhat smaller in diameter and less deeply rooted than those encountered in other habitats. Occasionally, clumps with only three or four fertile culms were found on the beaches.

The species does not seem to provide palatable forage; cattle were observed to turn it down in favor of the more succulent *S. patens* and species of *Paspalum*.

S. spartinae flowers from March to October in North America and from December to May in South America.

4. SPARTINA ALTERNIFLORA

- Spartina alterniflora Lois Fl. Gall. 719. (1907)
(Type from Bayonne, France)
- Dactylis cynosuroides var. β L. Sp. Pl. ed. 1. 1:71. (1753)
(Type in BM from Virginia)
- Dactylis maritima Walt. Fl. Car. 77. (1788) non Curtis (1787)
- Trachynotia alterniflora (Lois.) DC. Fl. Fr. 6:279. (1815)
- Spartina glabra Muhl. Descr. Gram. 55. (1817)
- Limnetis glabra Nutt. Gen. 1:38. (1818) nomen nudum
- Spartina laevigata Link, Jahr. Gewächsk. I. 3:92. (1820)
(Type in B from North America)
- Spartina brasiliensis Raddi, Agrost. Bras. Atti Del. Reale Accad. 2:21. (1823) (Type in PI from Rio de Janeiro, Brazil)
- Trachynotia alternifolia Steud. Nom. Bot. ed. 2. 695. (1841)
(Error for T. alterniflora)
- Spartina dissitiflora Steud. Syn. Pl. Glum. 215. (1885)
(Type in P from Bahia, Brazil)
- Spartina stricta (Ait.) Roth var. alterniflora (Lois.) Gray,
Man. Bot. ed. 2. 552. (1856) (Based on S. alterniflora Lois.)
- Spartina stricta (Ait.) Roth var. glabra (Muhl.) Gray,
Man. Bot. ed. 2. 552. (1886) (Based on S. glabra Muhl.)
- Spartina stricta (Ait.) Roth var. maritima (Walt.) B.S.P. Prelim.
Cat. N.Y. Pl. 66. (1888) (Based on D. maritima Walt.)
- Spartina glabra Muhl. var. alterniflora (Lois.) Merr. Bull. U.S.
Dept. Agr. Pl. Ind. 9:9. (1902) (Based on S. alterniflora Lois.)
- Spartina glabra Muhl. var. pilosa Merr. Bull. U.S. Dept. Agr.
Pl. Ind. 9:9. (1902) (Type in US from Atlantic City, New Jersey)
- Spartina alterniflora Lois. var. glabra (Muhl.) Fern. Rhod. 18:178.
(1916) (Based on S. glabra Muhl.)
- Spartina alterniflora Lois. var. pilosa (Merr.) Fern. Rhod. 18:179.
(1916) (Based on S. glabra pilosa Merr.)
- Spartina intermedia Bose ex St. Y. Cand. 5:50. (1932) in obs.
- Spartina maritima (Curt.) Fern. subsp. glabra (Muhl.) St. Y.
Cand. 5:48. (1932)
- Spartina maritima (Curt.) Fern. subsp. glabra (Muhl.) St. Y.
var. alterniflora (Merr.) St. Y. Cand. 5:53. (1932)
(Based on S. alterniflora Lois.)
- Spartina maritima (Curt.) Fern. subsp. glabra (Muhl.) St. Y.
var. alterniflora (Merr.) St. Y. forma gracilis St. Y. Cand. 5:54.
(1932) (Type not designated)
- Spartina maritima (Curt.) Fern. subsp. glabra (Muhl.) St. Y.
var. brasiliensis (Raddi) St. Y. Cand. 5:56. (1932)
- Spartina maritima (Curt.) Fern. subsp. glabra (Muhl.) St. Y.
var. brasiliensis (Raddi) St. Y. subv. fallax St. Y. Cand. 5:57.
(1932) (Type not designated)
- Spartina maritima (Curt.) Fern. subsp. glabra (Muhl.) St. Y.
var. brasiliensis (Raddi) St. Y. subv. raddii St. Y. Cand. 5:56.
(1932) (Based on S. brasiliensis Raddi)
- Spartina maritima (Curt.) Fern. subsp. glabra (Muhl.) St. Y.
var. glabra (Gray) St. Y. Cand. 5:49. (1932)

- Spartina maritima (Curt.) Fern. subsp. glabra (Muhl.) St. Y.
 var. glabra (Gray) St. Y. subv. typica St. Y. Cand. 5:51. (1932)
 (Based on D. maritima Walt.)
- Spartina maritima (Curt.) Fern. subsp. glabra (Muhl.) St. Y. var.
glabra (Gray) St. Y. subv. pilosa (Merr.) St. Y. Cand. 5:51. (1932)
- X Spartina merrillii Chev. Bull. Soc. Bot. Fr. 80:787. (1933)
 (Type in NY from Long Island, New York)

Description (Plate II, a-f)

Culms to 2.5 meters tall, erect, terete, fleshy, glabrous, usually solitary or in small clumps from widely spreading, elongate, flaccid, whitish rhizomes, scales inflated, not closely imbricate; sheaths smooth, the lower often wrinkled, glabrous, margin of the orifice glabrous to minutely pilose; ligule a ring of hairs 1--2 mm long; blades 3--25 mm wide, flat, occasionally becoming involute, glabrous to sparingly pilose on the adaxial surface, glabrous on the abaxial, margins usually glabrous, tip acuminate; panicle 10--40 cm long, often partially enclosed within the uppermost sheath; panicle axis 3-angled, glabrous, rarely sparingly scabrous on the angles; spikes 3--25, 5--15 cm long, alternate, loosely appressed, subremote to moderately imbricate, sessile or pedunculate; rachis of the spikes glabrous to sparingly scabrous on the angles, often prolonged beyond the terminal spikelet; spikelets 10--30, 8--14 mm long, alternate, spreading, occasionally appressed, subremote to closely imbricate; first glume linear, 4--10 mm long, usually longer than one-half the second, glabrous to sparingly pilose, keel glabrous to sparingly pilose, keel glabrous to sparingly pilose, margins glabrous, tip acute; second glume 8--14 mm long, glabrous to pilose, keel glabrous to pilose, tip acuminate to obtuse, occasionally apiculate; lemma glabrous to sparingly pilose, keel glabrous to very sparingly pilose, tip usually acuminate; palea thin and papery, slightly exceeding the lemma, tip obtuse or rounded; anthers 3--6 mm long; somatic chromosome numbers 56, 70.

Observations

Spartina alterniflora exhibits a high degree of variability in certain vegetative characters as well as in parts of the inflorescence. Among the characters analyzed were culm height, blade length and width, panicle length, spike number and length, spikelet number and length, ratio of spike length to spike number and pubescence on the spikelets. Of these characters, none could be correlated with another to show any population differences within the species.

The variability in culm height and spike number is less pronounced among specimens from the maritime provinces of Canada. These plants are smaller, less robust in appearance and have fewer spikes than some of the specimens from the remainder of the North American range of distribution. However, a number of specimens from the remainder of the range are quite similar to the more northerly specimens. Attempts to find suitable taxonomic distinctions between the more robust and less robust specimens failed.

Of all the characters examined, only two, the ratio of spike length to spikelet number and pubescence on the spikelets, exhibit variability that is definitely correlated with geographic distribution. Visual inspection

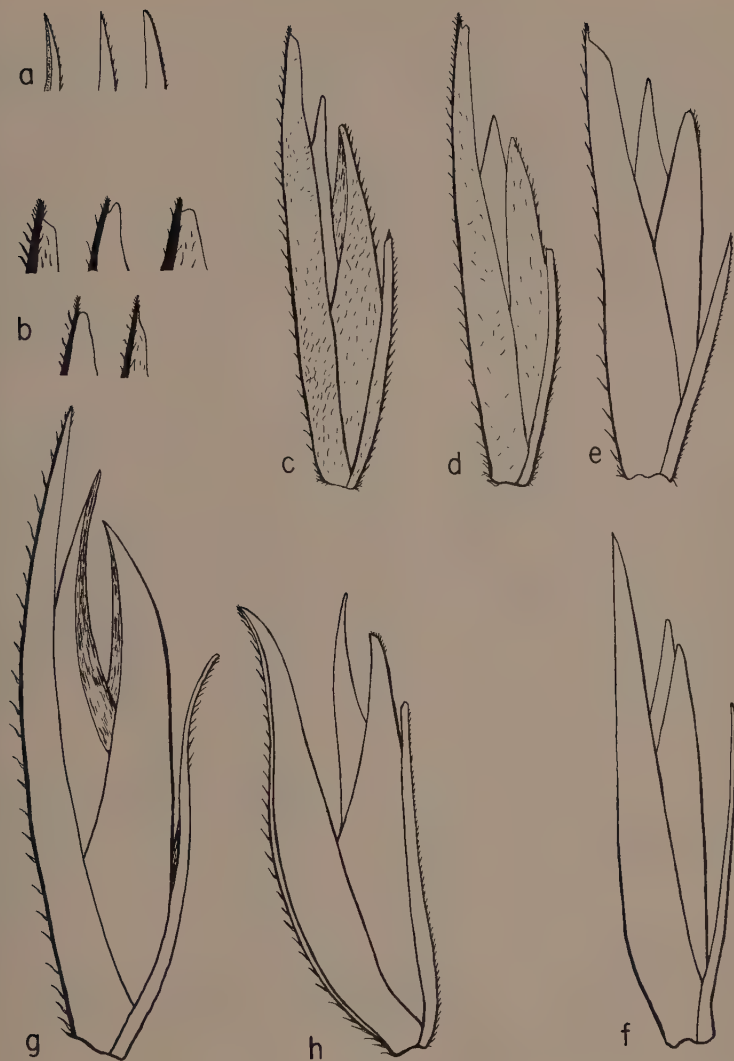


PLATE II

- a-f. *Spartina alterniflora*. a. first glume tips 7.5X,
 b. second glume tips 7.5X,
 c-f. variations in spikelets and
 pubescence 3.75X.
- g-h. *Spartina foliosa*. g-h. spikelets 3.75X.

of specimens collected from Maine northward to the maritime provinces of Canada showed that spikelets are not as closely imbricate along the spike as they are in plants from the remainder of the North American range. Further, plants from the southern half of the range (South Carolina to Texas) have spikelets that are noticeably less pubescent than those of plants from the northern half of the range. Plants of the intervening region (North Carolina to New Hampshire) seem to have spikelets more closely imbricate than do the northern specimens and considerably more pubescent than the southern specimens. In an effort to learn the extent of the variability exhibited by these two characters, the range of distribution of the species was divided into three segments on the basis of the observed variability. Section A included that part of the range from Maine northward, B from New Hampshire to North Carolina and C from South Carolina to Texas.

In order to effect a means by which numerous specimens could be compared, the two variables were reduced to numerical terms. A numerical equivalent for the degree of imbrication of spikelets is the quotient of spike length divided by spikelet number. Eight general classes of pubescence were recognized based upon the presence or absence of pubescence on laminae and keels of the second glume and lemma. In the interest of greater accuracy, the laminae were given independent consideration. While the amount of pubescence on any one part seems to vary somewhat (grading from glabrous to copiously pilose) a part was considered pubescent if it possessed at least five pilose hairs. Of all the combinations possible, only eight were found among the specimens examined. These varying degrees of pubescence were provided with numerical values according to the following table.

Table 4. Pubescence Values for *Spartina alterniflora*.

Second glume		Lemma		Numerical value
Lamina	Keel	Lamina	Keel	
Glabrous	Glabrous	Glabrous	Glabrous	1
Glabrous	Pilose	Glabrous	Glabrous	2
Glabrous	Pilose	Glabrous	Pilose	3
Glabrous	Pilose	Pilose	Glabrous	4
Pilose	Pilose	Glabrous	Glabrous	5
Pilose	Pilose	Glabrous	Pilose	6
Pilose	Pilose	Pilose	Glabrous	7
Pilose	Pilose	Pilose	Pilose	8

Three scatter diagrams (Fig. 2 A, B, C) are provided, one for each section of the range of distribution. Each specimen is introduced into the diagram by plotting its pubescence value against its numerical equivalent of imbrication of the spikelets. It may be seen from the diagrams that plants from the geographic extremes tend to group themselves into two somewhat segregated clusters. While the evidence presented shows population differences, the differences are not considered sufficient to

warrant the application of specific or infraspecific names. Some authors have sought to distinguish infraspecific taxa based upon the amount of pubescence on the lemma but it should be noted in this connection that pubescence on spikelets of some individual specimens varied from spikelet to spikelet. However, in this work, the pubescence value for any single specimen was always based upon the median spikelet of the median spike.

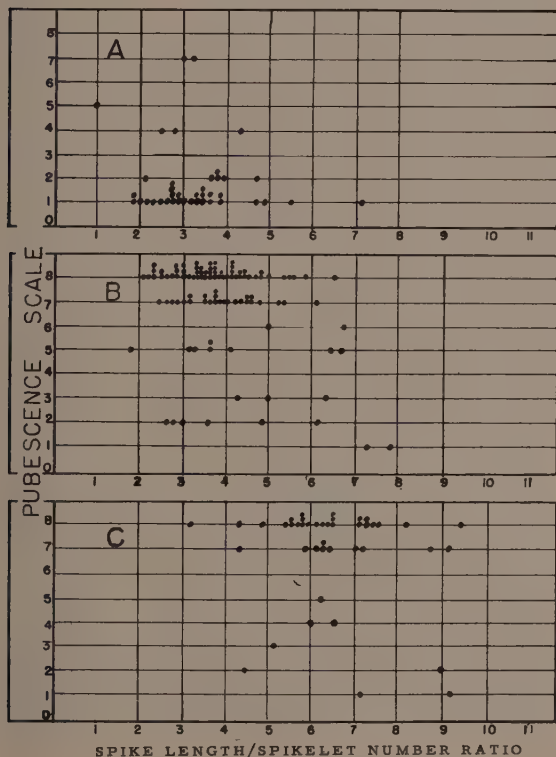


Fig. 2. Comparison of specimens of *Spartina alterniflora* of differing geographic localities by pubescence values and spikelet imbrication values.

A. Maine and the Maritime Provinces of Canada

B. New Hampshire to North Carolina

C. South Carolina to Texas

Church (1940) has shown the existence of two levels of polyploidy within this species; an octoploid ($2n = 56$) and a decaploid ($2n = 70$) which are supposedly morphologically and ecologically distinct. The octoploid form, found in the intertidal zone of the strand areas, was characterized by a height of 5--8 dm, leaf length of 2--4 dm, six to ten spikes and spikelets 10--12 mm long. The decaploid form, found along the muddy banks of tidal lagoons, was characterized by a height of 10--25 cm, leaf

length of 6--8 dm, fifteen to twenty spikes and spikelets 11--14 mm long. Nomenclaturally, the octoploid was referred to S. alterniflora var. glabra Fern. and the decaploid to S. alterniflora var. pilosa Fern. Some of the specimens from Section A of the range seem to agree with most of the criteria proposed and the less robust plants of this region may be the octoploid form. However, a majority of the specimens from Sections B and C of the range do not answer to the criteria. Many specimens with six to ten spikes exceed ten decimeters in height. In addition, many specimens with fifteen or more spikes have spikelets less than 11 mm in length. Two levels of polyploidy may exist throughout the range but, owing to the high degree of morphological variability, which is doubtless influenced by environmental conditions, they defy taxonomic separation.

S. alterniflora appears to be indigenous to the Americas but it has been introduced into other parts of the world, particularly England and France. The first record of a collection of this species in Europe is that of Loiseleur (1807), from the Bas-Pyrenees region of southwestern France. All specimens examined which had been collected from southwestern France agreed for the most part with the less robust North American form with its loosely imbricated spikelets. Although chromosome numbers are unknown for plants from France, it seems quite likely that this segment of the population may be the octoploid form. Specimens from collection stations in England and northern France exhibit about as much variability as those from the northern half of the North American range, although none of the European specimens are completely glabrous. Cytological studies have shown the decaploid to be present in England (Huskins, 1931). The role of this decaploid in the formation of the allopolyploid S. townsendii is discussed more fully in connection with that species.

The range of the species also includes South America and the West Indies. The specimens from collection stations in South America do not exhibit characters by which they may be distinguished from North American material. They exhibit about the same range of variability with respect to the imbrication value (Figs 2, 6A). The pubescence on the lemma keel seems to be absent except for a few soft trichomes near the tip. This is a fairly constant feature of the South American material. A single character which may set apart many (but not all) of the South American specimens is the pedunculate spike. However, the presence of peduncles is not consistent; it is also occasional in North American material.

Nomenclature

Spartina alterniflora seems to be properly typified. Although the type specimen was not seen, study of the description verifies the accuracy of application of the name.

Numerous infraspecific taxa have been proposed but little credence can be given to the arguments supporting them. Fernald (1916, 1950) resolved a good share of the nomenclatural problems. His treatments correct many of the earlier names erroneously applied to the subdivisions of the species in eastern North America. According to Fernald, three "pronounced varieties" are to be found (Fernald, 1950), S. alterniflora Lois. "typical," S. alterniflora var. glabra (Muhl.) Fern. and S.

alterniflora var. pilosa (Merr.) Fern. The "typical" is supposedly distinguished from the two named varieties on the basis of the imbrication of the spikelets which are said to be subremote, barely overlapping. It is thought to be distinct from var. glabra in that lemmas are minutely pilose, and that the rachis of the spike is prolonged beyond the terminal spikelets. The "typical" is supposedly separable from var. pilosa solely on the basis of spikelet imbrication. Fig. 2, A, B, C, shows that there is no sharp line of demarcation separating two groups of the population upon this basis. Further, glabrous lemmas were observed in a number of the specimens with more remotely disposed spikelets. Exsertion of the rachis of a spike beyond the terminal spikelet is found quite consistently among specimens from all parts of the range of distribution. The fact that the rachis does not exceed the terminal spikelet in some specimens does not appear to be correlated with other morphological differences. No other criteria were found which would aid in the satisfactory delimitation of these taxa. In view of these obstacles to accurate identification, it seems advisable to regard these infraspecific names as synonyms of S. alterniflora in accordance with Hitchcock (1935), Swallen (1939), and Chase (1950).

In the recent monographic study of the genus by Saint-Yves (1932), S. alterniflora is considered as S. maritime subsp. glabra St. Y. Under the subspecies, three varieties, four subvarieties, and one forma are recognized. Variety brasiliensis is considered discrete from the remainder on the basis of its pedunculate spikes and distribution in South America. As noted above, this feature is somewhat inconsistent in South American material and does not appear to be a satisfactory means of separation. Two subvarieties are proposed for var. brasiliensis, based upon the variable number of spikes. Three to eight spikes characterize subv. fallax and fifteen to twenty are typical of subv. radii. Examination of a number of South American specimens shows variation in spike number from two to eighteen without distribution into two distinct groups.

Variety alterniflora of Saint-Yves, distributed in North America and Europe, is supposedly separable from var. glabra (also distributed in North America and Europe) on the basis of spikelet length, said to be 12--14 mm in var. alterniflora and 9--11 mm in var. glabra. A second criterion is the relative lengths of the glumes. First glumes are supposedly shorter than or equal to one-half the length of the second in var. glabra and longer than one-half the length of the second in var. alterniflora. Lengths of the spikelets and glumes have been found to be quite variable throughout the species, but they do not show the correlation implied by the criteria of Saint-Yves.

Saint-Yves proposed two subvarieties for var. glabra. Subvariety typica is said to be separable from subv. pilosa by virtue of the glabrous spikelets and six to ten spikes. Pilose spikelets and three to five spikes are said to characterize var. pilosa. Study of the specimens from Europe and North America shows that spike number is not correlated with the variable pubescence of the spikelets. A forma gracilis is proposed for var. alterniflora. It is described as merely a slender form of the typical alterniflora. Saint-Yves states that specimens of var. alterniflora grade imperceptibly between the slender form and the typical.

Results of a critical analysis of the taxa proposed by Saint-Yves show

that the criteria for separating them are inconsistent; it therefore seems advisable to regard these names as synonyms of S. alterniflora Lois.

A hybrid, X S. merrillii, between S. alterniflora and S. cynosuroides was suggested by Chevalier (1933). The evidence for hybrid affinities exhibited by the "hybrid" is a greater number of spikes than ordinarily encountered in S. alterniflora. However, the type specimen (Bicknell 11300, Long Island, N.Y., NY) possesses two panicles, one with twenty-one spikes, the other with seven spikes. Duplicates cited by Chavalier were also examined. (Kearney, Long Island, N.Y., US, F; MacKenzie 7339, Leonardo N.J., 7501, Ocean City, N.J., NY; Ferguson 724, 725, 726, 759, Long Island, N.Y., NY). So far as can be determined, none of the specimens reveal any marked morphological dissimilarities between them and the remainder of the highly variable population. Spike number varies from three to twenty-five throughout the species. The "hybrids" may be nothing more than specimens with a greater number of spikes.

Distribution

As noted in the preceding discussion, this species ranges over the east coast of North and South America, England, and France. It has also been introduced, through oyster culture, into northwestern United States. Maps showing the distribution of the species are shown in Figs. 3, 4, 5. A single specimen is known from Guadeloupe, West Indies (Perottet, Antilles, Guadeloupe, 1842P; fragment in US).

The plant is normally one of the intertidal zone, inhabiting the muddy banks of lagoons which are subject to tidal activity. It is usually found in salt water habitats and often occurs in large, dense, pure stands that range from muddy permanent shore lines well out onto the strand. However, Fernald (1941) found the plant in fresh to brackish water sites along the James River, Virginia. In pure stands which were observed to stretch for over one hundred meters or more, some variation was observed in the height of the plant and in the vigor of the panicle. In some cases, very slender plants were found intermingled with the larger ones, but visual inspection showed that a continuous range of variation was present.

Perhaps the most striking feature of this species is its ability to bind soil along the coast. Rhizomes are thick, fleshy and are embedded quite deeply in the silt. The large air spaces in the rhizomes and the highly inflated scales doubtless aid this species in persisting in areas where little atmospheric oxygen is likely to be found in the soil. To the unhappy plant collector, the most striking thing about the plant in its native habitat is the strong, unpleasant odor that accompanies the dislodged culms. This noxious, sulphurous odor has persisted in some of the author's collections for nearly a year.

Flowering dates seem to be quite variable for this species. In North America, it flowers from June to October; in South America from December to June; and in Europe from July to November. Flowering seems to be somewhat erratic, for among Florida specimens examined in the field in early August, some inflorescences had emerged some time before and had been completely eaten away by insects, while others were still well enclosed within the leaf sheaths. Most New England plants examined in late August were at or beyond the peak of anthesis.



Fig. 3. Distribution of Spartina alterniflora in North America.



Fig. 4. Distribution of Spartina alterniflora in South America.

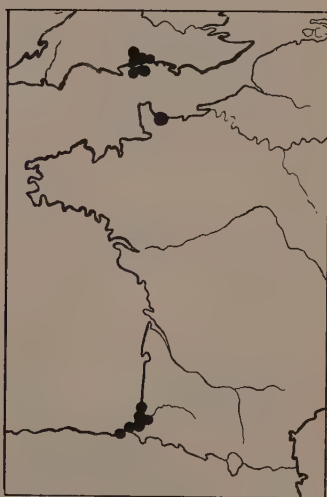


Fig. 5. Distribution of Spartina alterniflora in Europe.

5. SPARTINA FOLIOSA

Spartina foliosa Trin. Mem. Acad. St. Petrsbg. Sci. Nat. VI.
6:114. (1840) (Type in LE from California)

Spartina leiantha Benth. Bot. Voy. Sulph. 56. (1844)
(Type in K from Baja, California)

Spartina densiflora Brong. var. typica St. Y. subv. brongniertii St. Y.
forma acuta St. Y. Cand. 5:76. (1932) (Based on a specimen
from Humboldt County, California deposited in the Delessert
Herbarium, G)

Description (Plate II, g-h)

Culms to 1.5 meters tall, erect, terete, fleshy, glabrous, commonly with adventitious roots produced from the lower nodes, usually solitary or in small clumps, from widely spreading, more or less slender, flaccid, whitish rhizomes, the scales not closely imbricate; sheaths smooth, the lower somewhat wrinkled, margin of the orifice pilose; ligule a ring of hairs 1--2 mm long; blades flat to loosely involute, glabrous on both surfaces, margins glabrous to very sparingly scabrous, tip acuminate; panicle 12--25 cm long, usually cylindrical, often partially enclosed within the uppermost sheath; panicle axis 3--4-angled, flattened, twisted, glabrous; spikes 3--25, 2--8 cm long, the lower usually the longer, alternately to irregularly arranged, subremote below, becoming crowded toward the apex of the panicle, sessile, usually tightly appressed and often twisted, giving the panicle a cylindrical contour; rachis of the spikes glabrous, rarely prolonged beyond the terminal spikelet; spikelets 8--30, 8--25 mm long, alternate, appressed or occasionally spreading, moderately imbricate, often subremote on the lower spikes, not readily disarticulating; first glume linear, 6--12 mm long, often curved, usually more than one-half as long as the second, glabrous to sparingly pilose, keel glabrous to sparingly pilose above, glabrous to long-pilose at the base, tip acuminate to obtuse or rounded; second glume 8--25 mm long, often curved, glabrous to sparingly pilose, keel glabrous to sparingly pilose, glabrous to long-pilose at the base, margins glabrous to sparingly pilose, tip acuminate to blunt or rounded; lemma glabrous to sparingly pilose, keel glabrous, margins glabrous to sparingly pilose, tip obtuse, rounded or lobed; palea thin and papery, glabrous, tip acuminate, rarely rounded, slightly exceeding the lemma; anthers 3--6 mm long; somatic chromosome number 56.

Observations

Spartina foliosa is a rather uniform species. The vegetative features do not exhibit any peculiarities of taxonomic usefulness. Plants of this species seem to agree very closely in vegetative parts with other members of the same complex. The panicle, on the other hand, is strikingly different in several aspects. The spikes on a single panicle show definite variability in length. Basal spikes are usually much longer than the median. In addition, the lower spikes are much less closely imbricated along the panicle axis than are the upper ones. Numerous, tightly appressed spikes give the panicle a cylindrical contour. In this respect the species approaches a characteristic of Complex 1. The species

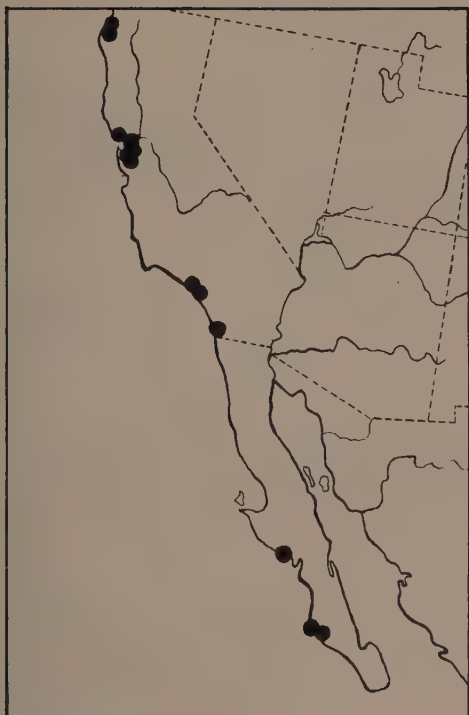


Fig. 6. Distribution of *Spartina foliosa* along the west coast of North America.

does exhibit some variability in panicle length, spike number and length, spikelet length and pubescence on the spikelets. However, none of the variations encountered could be correlated with geographic distribution or other morphological features in any way that would suggest population differences or differing levels of polyploidy. The octoploid chromosome number ($2n = 56$) has been reported for the species (Church, 1940).

Nomenclature

A fragment of the type specimen deposited in the United States National Herbarium was available for examination. There is no doubt of the accuracy of the typification.

The specimen upon which the name *S. densiflora* ... forma *acuta* St. Y. was based was not available for examination. Saint-Yves (1932) notes that Hitchcock had identified the specimen as *S. foliosa* with which it agreed in all particulars save spikelet length. This specimen possessed spikelets 7--8 mm long while the spikelet lengths in *S. foliosa* were, according to Saint-Yves, 12--16 mm. Spikelet lengths found during the present investigation range from 8--25 mm. The fact that the forma of Saint-Yves is based upon a single character which exhibits so much variability seems reason enough to reject the name and treat it as a synonym of *S. foliosa*. While habitat has not been shown to influence directly the size of spikelets, it may be that it does. In this case, it may be significant to note that the label of the specimen in question bears the information that it was collected in the drier part of the salt marsh at Eureka, California.

Distribution

Spartina foliosa appears to be limited to the Pacific coast of North America (Fig. 6). It occurs in the intertidal zone along the edges of the strand, in the salt marshes and in tidal lagoons. The plant flowers from July to November. The species does not seem to produce viable seed regularly. Labels accompanying collections from California indicate that ergotization of the ovary is of common occurrence.

6. SPARTINA LONGISPICA

Spartina longispica Haum. and Par. ex St. Y. Cand. 5:92. (1932)
(pro hybr.) (Type in G from Pipinas, Argentina)

Description (Plate III, a-b)

Culms to 2 meters tall, erect, terete, often fleshy, glabrous, usually solitary from short, tough or elongate, flaccid rhizomes, the scales moderately imbricate; sheaths smooth to somewhat striate, the lower often wrinkled, glabrous, margin of the orifice glabrous, rarely pilose; ligule a ring of hairs 1--2 mm long; blades flattened below, becoming involute toward the apex, glabrous to scabrous on the adaxial surface, glabrous on the abaxial, margins glabrous to scabrous, tip acuminate; panicle 14--45 cm long, often partially enclosed within the uppermost sheath; panicle axis 3-angled, glabrous to sparingly scabrous on the edges; spikes 4--16, 4--18 cm long, 2--5 mm wide, alternate, loosely to closely appressed, usually pedunculate; rachis of the spikes glabrous to scabrous, rarely prolonged beyond the terminal spikelet; spikelets 10--35, 12--24 mm long, 1.5--2.5 mm wide, alternate, often tightly appressed, occasionally spreading, moderately imbricate, often pedicellate, the pedicel pilose; first glume linear, 6--12 mm long, about one-half the length of the second, glabrous to sparingly pilose, keel pilose to hispid, margins glabrous or very sparingly hispidulous, tip acuminate to obtuse; second glume 12--24 mm long, glabrous to sparingly pilose, keel pilose to hispid, margins glabrous or sparingly hispidulous, tip acuminate to obtuse; lemma glabrous to sparingly pilose, keel glabrous below, sparingly hispid or pilose above, tip acuminate, rounded or obtuse; palea thin and papery, glabrous, tip obtuse to acuminate; anthers 3--7 mm long.

Observations

Spartina longispica was described by Saint-Yves (1932). He considered it a hybrid between the intertidal S. brasiliensis and the dry-land, S. montevidensis. Evidence with which Saint-Yves purported to show hybrid affinities was found in the nature of the rhizomes, the leaf blades and the inflorescence. The rhizomes of S. brasiliensis are long and flaccid. Rhizomes in S. montevidensis are wanting or very short. The intermediate condition, supposedly reflected by the hybrid is suggested by short, stolon-like rhizomes. Examination of the cross-section of leaf blades of the species concerned showed important evidence of hybridization. Differences were found in the amount and position of chlorenchyma within the leaf blades. Inasmuch as living material was not available, this portion of Saint-Yves work was not verified during the present investigation. Through a study of the inflorescences of the putative parents

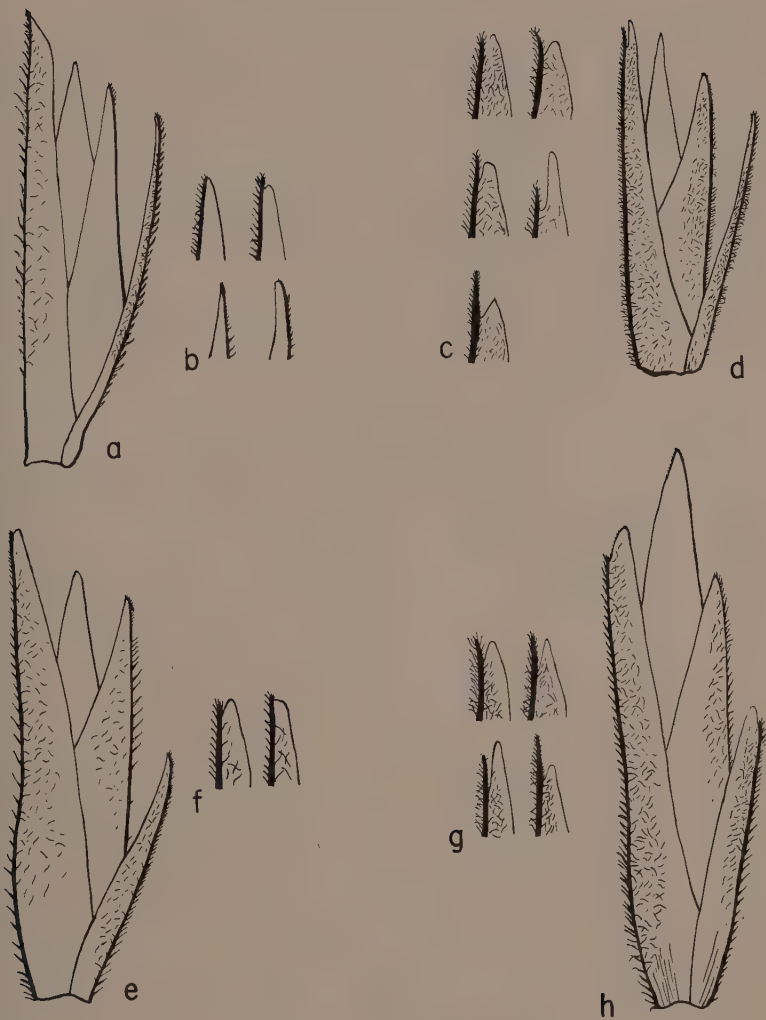


PLATE III

- a-b. *Spartina longispica*. a. spikelet 3.75X, b. second glume tips 3.75X.
 c-d. *Spartina maritima*. c. second glume tips 7.5X, d. spikelet 3.75X.
 e-f. *Spartina neyrautii*. e. spikelet 3.75X, f. second glume tips 7.5X.
 g-h. *Spartina townsendii*. g. second glume tips 3.75X, h. spikelet 3.75X.

and the hybrid, Saint-Yves claimed to have found differences which further suggested hybrid affinities. In S. brasiliensis, spikes were long and loosely appressed against the smooth panicle axis. Spikes of S. montevidensis were shorter and more or less tightly appressed against the axis which was smooth below and scabrous above.

Parodi (1937) maintained that the three species are found quite close together. He stated that S. brasiliensis occupies the intertidal zone, S. montevidensis the dry beach and S. longispica the intermediate habitat. S. montevidensis is thought to be separable from the other two on the basis of its much shorter spikelets (said to be 8--12 mm long), its sessile spikes and its lack of extensive rhizomes. S. brasiliensis is supposedly separable from S. longispica on the basis of its longer spikelets. Spikelet lengths of 15--18 mm are given for S. brasiliensis while lengths of 12--14 mm are given for S. longispica. The rhizomes of the latter two are also shown to differ in accordance with Saint-Yves' distinctions given above.

The name S. brasiliensis and S. montevidensis by which Saint-Yves and Parodi have referred to the two putative parents have been shown in this paper to be synonyms of S. alterniflora and S. densiflora respectively. Hereinafter, the two parent species will be referred to by the latter binomials.

At first glance the hypothesis presented by Saint-Yves and Parodi seems quite plausible, but there are certain inconsistencies. Many specimens previously identified as either S. alterniflora (S. brasiliensis) or S. longispica do not answer to their criteria. The pubescence causing scabridity on the panicle axis is quite variable and does not seem to be correlated with appression of the spikes. Spikelet lengths from the same plants may show a variation from 13--16 mm in length. In addition, certain specimens identical with regard to spikelet length differ in the type of rhizomes. In some they are hard, short, and knotty, and in others long and somewhat flaccid, more or less typical of S. alterniflora. In short, identification of plants at hand is fraught with exceeding difficulty.

A possible explanation for the numerous plants which fail to fulfill the criteria proposed is that they are individual hybrid progenies which, because multiple factors are involved, segregate out along more or less of a gradient toward either parent, or that they are simply backcrosses to either parent and, as a result, stand somewhat intermediate between the putative F_1 hybrid and the original parent. A combination of the two preceding alternatives might also be involved. If these conditions were true one might expect a rather continuous range of variations from one parent to the other. Such, however, is not the case, for S. densiflora is discretely separable from the others on several counts, while the more puzzling specimens seem to be variously disposed between the putative hybrid and the intertidal S. alterniflora. Therefore some other explanation must be sought.

It became apparent to this investigator after considerable study of the problem that some plants previously identified as S. alterniflora (S. brasiliensis) were not actually S. alterniflora at all, but were members of another population altogether. This population is similar to S. alterniflora in many respects, but the panicle is obviously much more robust. In connection with geographic distribution, a curious fact was brought to

light. This group of more robust specimens, which included those previously identified as hybrids, were localized at or near the mouth of the River Plate, while virtually all of the less robust true S. alterniflora were distributed further northward along the coast.

The plants of the River Plate population are not easily separated from S. alterniflora by means of any single key character but, as will be shown in connection with the study of the hybrid origin of the population, certain quantitative indices may be successfully employed. Accordingly, this discrete population is named S. longispica.

The hypothesis of the hybrid origin of S. longispica advanced by Saint-Yves (1932) and corroborated by Parodi (1937 and in personal communication) is not without merit. However, it seems evident that what they were attempting to define as hybrids are perhaps ecological variants of what may be a larger hybrid population. The fact that plants with short, hard rhizomes are found in the intermediate habitat may be an indication that environmental conditions influence the ultimate size and conformation of rhizomes.

In an effort to explore more fully the conclusions drawn by Saint-Yves (1932) regarding the hybrid origin of S. longispica, a hybrid index was constructed according to methods described by Anderson (1936, 1949) and Larisey (1940). A number of characters were selected by which the putative parent populations may be distinguished. Several of these characters serve to separate the parents discretely but the remainder reflect only the general trends of the populations, the extremes overlapping somewhat. An index value of 0 was given to a particular character if it was typical of S. alterniflora, while a value of 2 was assigned if it more closely represented S. densiflora. In several of the features used to separate the parent populations, an intermediate condition was found. This intermediate was given a value of 1. A total of 14 characters were selected, the values of which, for any single specimen, when added together provided a minimum total value of 0 for S. alterniflora and a maximum total value of 28 for S. densiflora.

This index was constructed solely from features discernible in herbarium specimens; living material was not available. Thus, certain characters which may actually offer a means of separation of the putative parents were not used because they were not always present on the herbarium sheets. This was particularly true of the basal portion of the culms and the rhizomes.

The 14 characters used in the construction of the hybrid index follow:

1. The leaf blades of S. alterniflora are normally flat, becoming somewhat involute near the apex. This latter feature is no doubt accentuated by drying. On the other hand, leaves of S. densiflora are decidedly involute throughout the length of the blade. An intermediate condition is suggested in several of the putative hybrids by a flattened base becoming markedly involute a short distance above the base. Since field observations were not made on these specimens it is entirely possible that this suggested intermediate condition is purely an artifact.

2. A great majority of the specimens from South America referable to S. alterniflora have rather long-peduncled spikes. Some specimens

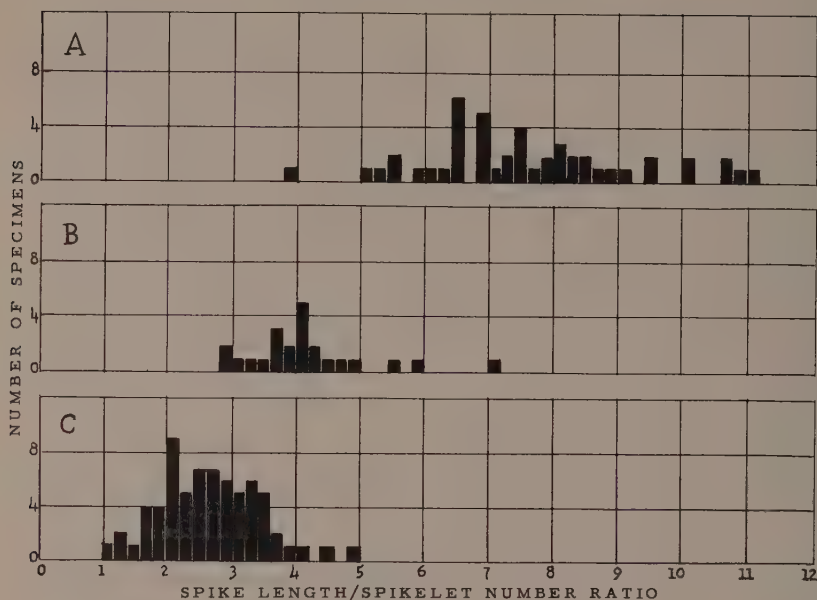


Fig. 7. Distribution of Spikelet Imbrication Values

A. *Spartina alterniflora*. B. *Spartina longispica*. C. *Spartina densiflora*.

may have lower spike peduncles as long as 5 cm. In the case of *S. densiflora* the spikes are sessile. The intermediate condition may be present in the putative hybrids in which the peduncles of some of the lower spikes vary from 0.5--1.5 cm in length.

3. Spikes of *S. alterniflora* are more or less the same length from base to apex of an individual panicle. In contrast, the panicles of *S. densiflora* usually have rather elongate basal spikes with a perceptible diminution in spike length toward the apex.

4. During the dissection of spikes and spikelets, it was observed that spikelets of *S. densiflora* disarticulate rather readily from the rachis, while those of *S. alterniflora* are somewhat persistent. The condition does not appear to be universally true among specimens of *S. alterniflora*, but reflects only the general trend of the population. Spikelets of all the specimens referred to the putative hybrid disarticulated readily.

5. In South American specimens of *S. alterniflora*, the spikelets are noticeably less crowded along the axis of the spike. A numerical evaluation of this imbrication of spikelets is presented in Fig. 7A. A similar evaluation of the more densely imbricate *S. densiflora* is presented in Fig. 7C. The values are obtained by calculating a ratio of the spike

length to the number of spikelets on the median spike. Comparison of the two histograms shows that there is some overlap making the feature an unsatisfactory key character, but it also shows that general trends in the population differ to a considerable extent. The values for most of the putative hybrids fall near the center of the range of overlap (Fig. 7B). Accordingly, any specimen which showed a value between 3.5 and 4.5 was scored as an intermediate.

6. Spikelets may be slightly pedicellate in S. alterniflora while they are sessile in S. densiflora. Again, this character does not appear to be constant throughout but is a more or less general trend in the specimens examined. Because the lengths of pedicels in S. alterniflora varied throughout a range of only 0--2 mm, no attempt was made to define an intermediate.

7. Somewhat spreading spikelets are typical of S. alterniflora while complete appression of spikelets characterizes S. densiflora. Most of the putative hybrids have spikelets rather tightly appressed to the rachis.

8. The relative length of first and second glumes may be used to separate the parent populations, although individual variation within both species precludes its use as a key character. S. densiflora commonly possesses first glumes that are shorter than one-half the length of the second, while S. alterniflora first glumes are usually longer than one-half the length of the second. No attempt was made to define an intermediate, although in several specimens first glumes exactly half as long as the second were observed.

9. The pubescence on both glumes has been found to be a useful taxonomic character separating the putative parents. That of S. alterniflora consists of trichomes that are rather soft and somewhat flexible and of varying lengths and distribution (Plate II, a-f). Trichomes of S. densiflora, on the other hand, are uniformly short and rigid (Plate IV, i-k). The trichomes of several putative hybrids seemed to be somewhat more rigid than those of S. alterniflora and slightly longer than those of S. densiflora. These were scored with a value of one.

10. The tip of the first glume in S. densiflora is rather obtuse while that of S. alterniflora is acuminate (Plates II, a-f; IV, i-k). The greater majority of the specimens examined show this condition to hold quite generally. The putative hybrids, for the most part, answered to the S. alterniflora type.

11. The same feature was observed at the tip of the second glume although the conditions were reversed. The second glume tip of S. densiflora is acuminate but not awned. That of S. alterniflora is obtuse, rounded or somewhat toothed with the mid-nerve projecting slightly beyond the toothed margin. Plates II, a-f and IV, i-k show the common type of second glume tips encountered. The putative hybrids exhibited about as much variability as S. alterniflora.

12. In most of the South American specimens of S. alterniflora, the lemmas were observed to be more or less glabrous on the keel. Occasionally a few trichomes are present near the tip. The keel of the lemma in S. densiflora is hispidulous on the upper two-thirds. Most of the putative hybrids are pubescent on the upper two-thirds of lemma keel, although the trichomes are, in general, somewhat longer than those encountered in S. densiflora. The distinction between putative parents was based solely on the distribution of the trichomes.

13. The texture of the glumes appears to be a feature in which the putative parents differ somewhat. S. alterniflora second glumes are somewhat thin and papery and tend to be rather pliable. Those of S. densiflora are slightly thicker and somewhat indurated. The putative hybrids agreed for the most part with S. densiflora in this respect.

14. The tip of the palea in S. densiflora is acuminate, occasionally possessing trichomes along the two rather closely set nerves which converge near the tip. On the other hand, the tip of the palea in S. alterniflora is obtuse to rounded and lacks pubescence. Attempts to define an intermediate failed here, although considerable raggedness was observed in some of the palea tips of the putative hybrids. They were scored, however, as typical of S. alterniflora.

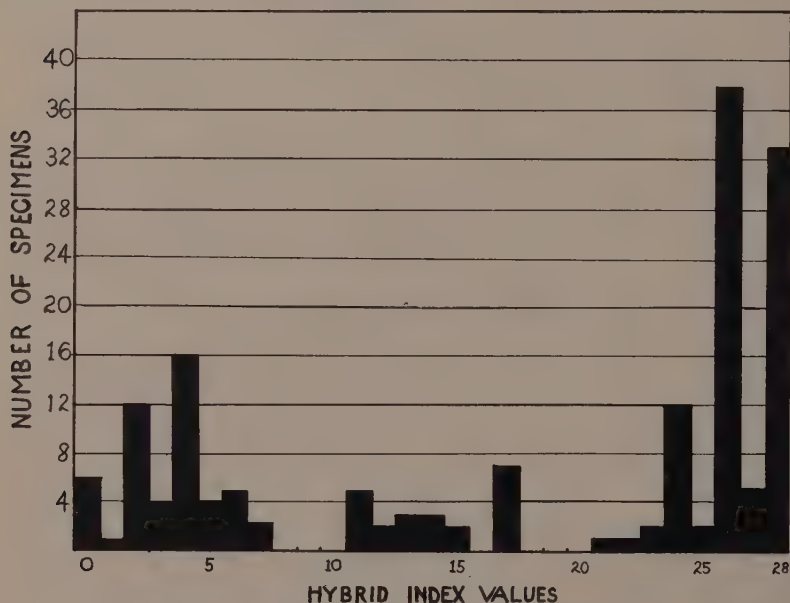


Fig. 8. Distribution of Hybrid Index Total Scores.

A total of 171 specimens were scored according to the features above. Results of the scoring are presented in Fig. 8. All the specimens within the intermediate cluster (11-17) were collected from the mouth of the

River Plate or near it. It should be noted that among plants within the intermediate cluster are specimens which had been identified by Parodi as *S. brasiliensis* (*S. alterniflora*) as well as several identified by him as *S. longispica*.

The specimens comprising this intermediate cluster are referable to *S. longispica*. From the evidence presented, they appear to be separable from both parents. Additional evidence showing that the *S. longispica* is discrete from *S. alterniflora* will be presented in this paper in the section dealing with distinctions between South American species of *Spartina*.

S. longispica, as defined in this paper, does have some characteristics which appear to relate it to *S. densiflora*. The ability to tolerate a dry land habitat is not known for any segment of *S. alterniflora*; yet it is shown by some specimens of *S. longispica*.

Two other features suggesting hybridization which do not lend themselves to treatment within the framework of the hybrid index are panicle and leaf length. Differences in these characters exhibited by *S. longispica* and *S. alterniflora* may be evidence of hybrid vigor. Measurements made on lengths of panicle and the second leaf from the summit are presented in Table 5. Panicle width measurements were found to be of little value, since pressing for herbarium storage often flattens and widens an otherwise narrow panicle.

Table 5. Lengths of panicle and blade in
S. alterniflora and *S. longispica*.

	<i>S. alterniflora</i>		<i>S. longispica</i>	
	Leaf	Panicle	Leaf	Panicle
Number of specimens	23	27	17	22
Minimum length (cm)	9.0	8.0	17.0	14.0
Average length "	17.6	15.6	29.5	28.0
Maximum length "	35.0	30.0	44.0	44.0

While the obvious overlap precludes use of the panicle and leaf length as key characters, there appears to be a significant difference in the average values for the two populations.

It is entirely possible that what is discussed here as a putative hybrid may not be a hybrid at all. There can be little doubt that the specimens from the area of the River Plate represent a population somewhat discrete from *S. alterniflora*. Further, it would appear from the evidence presented that some characteristics resembling *S. densiflora* actually appear in *S. densiflora*. This immediately suggests hybrid affinities or at least gene flow from *S. densiflora* to the extremely localized population.

It seems highly improbable, in view of the limited distribution, that hybridization is of common occurrence. Certainly concomitant distribution of the putative parents over extensive areas of the Brazilian coast offers ample opportunity for the frequent occurrence of hybridization. Yet, the putative hybrid population is extremely limited in its distribution.

A more likely explanation is that the entire population (both intertidal and intermediate zone plants) are the results of a single hybridization and that the resultant hybrid has proved fertile, or that the plant reproduces vegetatively so vigorously it has invaded territory which once may have been occupied by S. alterniflora and more or less obliterated it from the region. The case recalls that of the highly vigorous S. townsendii.

No specimens of S. alterniflora from the region occupied by the putative hybrid are deposited in any of the herbaria consulted during the course of this investigation. All specimens from that area had been previously identified as S. brasiliensis or S. alterniflora were distinctly referable to S. longispica.

The index described above, while useful for separating the three species involved, may not actually represent an accurate analysis of the supposed hybridization. If, as has been suggested, the hybridization occurred only once, then the index may be quite out of line in its utilization of trends in a variable population as bases for comparisons. Obviously, out of the two variable putative parent populations the individual plants which went into the formulation of the putative hybrid cannot be known.

Nomenclature

The population as delimited by Saint-Yves (1932) was thought to be of hybrid origin. It was designated X Spartina longispica. Parodi (1937) elected to regard it as a discrete species and dropped the hybrid symbol. However, he did retain the original author citation, viz. S. longispica Haum. and Par. apud Saint-Yves, but neglected to indicate that the rank of the taxon was altered from hybrid to species. Under articles 59 and 60 of the International Code of Botanical Nomenclature, which would seem to apply in this case, the name is correctly cited Spartina longispica (Haum. and Par. ex Saint-Yves) Par. (pro hybrid).

Distribution

S. longispica has a rather limited distribution. It exists only at the mouth of the River Plate in both Argentina and Uruguay.

The habitat of S. longispica also has been suggested in the preceding observations. While the intertidal zone appears to be the primary habitat, it nonetheless has been shown to invade the drier portions of the beach zone. Collecting dates indicate that the plants flower from January to May.

7. SPARTINA MARITIMA

Spartina maritima (Curt.) Fern. Rhod. 18:180. (1916)

Spartum essexianum, spica gemina clausa Petiver, Gram.

Concordia 1. (1716) (Type unknown)

Dactylis cynosuroides Loefl. Iter Hisp. 115. (1758) non Dactylis cynosuroides L. (1753) (Type from Lusitania)

Dactylis maritima Curt. Enum. Br. Gr. 4. (1787) non Dactylis maritima Walt. (1788) (Based on Spartum essexianum, spica gemina clausa Petiver)

Dactylis stricta Ait. Hort. Kew. 1:104. (1789) (Type from England)

Spartina stricta (Ait.) Roth, Neue Beitr. 101. (1802)

Paspalum cynosuroides (Loefl.) Brot. Fl. Lusit. 83. (1804)

(Based on D. cynosuroides Loefl.)

Trachynotia stricta (Ait.) DC. Fl. Fr. 3:74. (1805)

Limnethis pungens Rich. ex Pers. Syn. Pl. 1:72. (1897)

(Based on D. stricta Ait.)

Spartina pungens (Rich.) Beauv. Nouv. Agrost. 25. (1812)

Spartina capensis Nees, Fl. Afr. Austr. 260. (1841)

(Type from South Africa)

Spartina maritima (Curt.) Fern. subsp. stricta (Ait.) St. Y.

Cand. 5:45. (1932) (Based on D. maritima Curt.)

Description (Plate III, c-d)

Culms to 0.8 meter tall, erect, terete, glabrous, solitary or in small clumps from widely spreading, slender rhizomes, scales not closely imbricate; sheaths smooth to striate, the lower very often wrinkled, glabrous, margin of the orifice glabrous to very sparingly pilose; ligule a ring of hairs about 0.5 mm long; blades short, not exceeding 12 cm in length, more or less loosely involute, readily disarticulating from the sheath, glabrous to very sparingly scabrous on the adaxial surface, glabrous on the abaxial, margins glabrous, tip acuminate; panicle 4--14 cm long; panicle axis flattened, rounded or obscurely 3-angled, glabrous; spikes 1--7, commonly 2, 2--11 cm long, alternate, loosely appressed, moderately imbricate, sessile; rachis of the spikes glabrous, often prolonged beyond the terminal spikelet; spikelets 5--30, 10--15 mm long, alternate, loosely appressed to spreading, not closely imbricate; first glume narrow, 7--10 mm long, usually longer than one-half the second, densely pilose, keel pilose, margins glabrous, tip acuminate to obtuse or rounded; second glume 10--15 mm long, densely pilose, keel pilose, margins glabrous, tip acuminate, rarely obtuse; lemma densely pilose, keel pilose above, glabrous below, margins glabrous, tip obscurely lobed; palea thin and papery, glabrous, tip acuminate; anthers 4--6 mm long; chromosome number 56.

Observations

This species, limited to the Old World, is rather uniform. While it has attained a wide distribution ranging from the British Isles to South Africa, it is still remarkably uniform both vegetatively and in the inflorescence. The greatest variation is in the number of spikes, where as many as seven can be found. The predominant number is two--hence one of its common names, Two-spiked Cocksfoot Grass.

The octoploid chromosome number of 56 in plants from the British Isles has been reported by Huskins (1931). Specimens from the Mediterranean and South African segments of the range have not been cytologically analyzed.

Nomenclature

Curtis (1787) published the name Dactylis maritima and this epithet becomes the basis for the present combination of Fernald (1916), who clearly states the case for the ascription of the name. Although the publication of Curtis was in the briefest possible form, and did not itself contain a description of the species, it gave a clear reference to Ray

(1724) as the source for the description. Ray's description is based on Spartum Essexianum, spica gemina clausa of Petiver (1716)

Aiton (1789) described a specimen from England as Dactylis stricta. Roth (1802) recognized its affinities with the genus Spartina and accordingly published the combination Spartina stricta. This combination was in constant use for over a century. Only the most recent European works, viz. Clapham, Tutin, and Warburg (1952) have recognized the proper recombination of names as given by Fernald (1916).

Distribution

Spartina maritima is distributed exclusively in the Old World from the Netherlands and England south to the Cape of Good Hope along the Atlantic Seaboard. It has apparently become adventive at the head of the Adriatic Sea. Little is known of its distribution in Africa. Fig. 9 shows the distribution of the specimens examined.

The species appears to be limited to salt water habitats, on the strand or in tidal bays and lagoons. It does not seem able to withstand aggression by the more vigorous S. townsendii (Arber, 1934; Oliver, 1925), and in regions around Poole Harbor, England, it is being obliterated by S. townsendii. Collecting dates indicate that the species flowers from June to November in Europe and in February in South Africa.



Fig. 9. Distribution of Spartina maritima in Europe and Africa.

8. SPARTINA NEYRAUTII

Spartina neyrautii Fouc. Ann. Soc. Nat. Rochelle. 8. (1894)
(Type from Basse-Pyrenees)

Description (Plate III, e-f)

Culms to 1 meter tall, erect, terete, glabrous, usually solitary or in small clumps, from widely spreading, more or less slender rhizomes, scales not closely imbricate; sheaths striate, glabrous, margin of the orifice glabrous to short-pilose; ligule a ring of hairs 0.5--1 mm long; blades elongate, flattened, becoming involute toward the apex, glabrous on both surfaces, margins glabrous, tip acuminate; panicle 15--20 cm long, less than 5 mm wide; panicle axis 3-angled, glabrous; spikes 4--6, 8--12 cm long, alternate, tightly appressed, moderately imbricate, pedunculate, peduncles about 1 cm long; rachis of the spikes glabrous, not often prolonged beyond the terminal spikelet; spikelets 10--25, 14--16 mm long, alternate, tightly appressed, not closely imbricate, often pedicellate; first glume linear, 8--12 mm long, usually longer than one-half the second, pilose, keel sparingly pilose, margins glabrous, tip acuminate to obtuse, second glume 14--16 mm long, pilose, keel pilose, margins glabrous, tip usually bilobed; lemma glabrous to sparingly pilose above, keel pilose above, glabrous below, margins glabrous, tip acute, obtuse, rounded or obscurely lobed; palea thin and papery, slightly exceeding the lemma, tip acuminate to rounded; anthers 3--6 mm long.

Observations

Spartina neyrautii, like S. townsendii, has been adjudged a hybrid between the endemic European S. maritima and the introduced and variable S. alterniflora. Arber (1934) suggested that the two hybrids may have arisen in like manner, and that one may simply be the reciprocal of the other. Although the two populations have been considered identical by Saint-Yves (1932) and Chevalier (1923), Jovet (1941) has pointed to differences in the ligule and second glume tip by which the two may be effectively distinguished. Additional evidence in support of Jovet's contention is presented in a succeeding section of this paper dealing with means for separating and identifying species.

To date, cytological evidence supporting the hypothesis of the hybrid origin of S. neyrautii has not been published. In the absence of cytological findings, it is difficult to assess the origin of the species. That it combines characters of both S. maritima and S. alterniflora is evident. Pilosity of the glumes relates it to S. maritima while lack of early blade abscission may show it to be allied to S. alterniflora. Further evidence in support of hybrid origin for the species is found in its highly restricted distribution within a small segment of a larger area occupied by both putative parents.

Of particular importance to the resolution of the S. neyrautii problem is the nature of S. alterniflora distributed within the area now occupied by the hybrid. As shown in connection with observations on that species, all the specimens from southwestern France tend to be considerably less robust than those typical of the region around Southampton, England. That introduction of a different cytological segment of S. alterniflora

occurred suggests itself immediately. The less robust plants from southern France are not unlike those from America which have been shown by Church (1940) to have a somatic chromosome number of 56. It is therefore tentatively suggested that S. neyrautii, when cytologically examined, may be found to have a chromosome number quite different from that now known for S. townsendii. The fact that S. neyrautii is distinctly more slender, has generally more narrow spikes and shorter spikelets points to an obvious difference between the two species, somewhat more complex than that suggested to be due to reciprocal crosses.

Nomenclature

The description of S. neyrautii by Foucaud (1894) was written after study of specimens collected by E. J. Neyraut from Hendaye, France in 1892. No mention is made of a type specimen, and its whereabouts at the present time is unknown. However, duplicates of the Neyraut collection (E. J. Neyraut 3598, Hendaye, France, GH, US, and F) were examined.

Distribution

Specimens examined during the course of this study are shown by open circles in Fig. 10. The species is known only from a single station (Hendaye) in southwestern France. S. neyrautii occurs in salt marshes. While little is known of its ecology, it does not appear to be nearly so vigorous and aggressive as the closely related S. townsendii. The species has been observed to flower in August.



Fig. 10. Distribution of Spartina neyrautii and Spartina townsendii in Europe.

9. SPARTINA TOWNSENDII

Spartina townsendii H. and J. Groves. Bot. Exch. Club Rept. 37.
(1881) (Type from Southampton, England; isotype in K)

Description (Plate III, g-h)

Culms to 1.5 meters tall, erect, terete, glabrous, solitary or in clumps from widely spreading thickish rhizomes, scales not closely imbricate; sheaths smooth to striate, the lower often wrinkled, glabrous, margin of the orifice pilose; ligule a ring of hairs 1.5--3 mm long; blades elongate, flat, becoming involute toward the apex, glabrous on both surfaces, margins glabrous, tip acuminate; panicle 15--30 cm long, more than 8 mm wide, shorter and narrower in dwarf forms; panicle axis 3-angled, glabrous; spikes 3--10, 10--20 cm long, rarely longer, shorter in dwarf forms, alternate, loosely appressed, moderately imbricate, usually pedunculate, peduncle about 1 cm long; rachis of the spikes glabrous, prolonged beyond the terminal spikelet; spikelets 10--30, 16--22 mm long, alternate, not tightly appressed nor closely imbricate; first glume linear, 8--14 mm long, usually more than one-half as long as the second, pilose, keel pilose, margins glabrous to very sparingly hispidulous, tip acuminate; second glume 16--22 mm long, pilose, keel pilose, margins glabrous to sparingly hispidulous, tip acuminate to obtuse; lemma pilose, keel pilose above, glabrous below, margins glabrous, tip obtuse, rounded or obscurely lobed; palea thin and papery, glabrous, often exceeding the lemma by as much as 2 mm, tip acuminate; anthers 7--9 mm long; somatic chromosome number 126.

Observations

There now seems to be general agreement among European botanists that S. townsendii is a distinct species. The most recent treatment, that of Clapham, Tutin and Warburg (1952), recognizes it as discrete from other species of the genus occurring in England.

The origin of S. townsendii has received considerable attention in recent years as a striking example of a new species having arisen within the period of man's civilization. The actual mechanics of the origin of this species were worked out by Huskins (1931). The story is essentially complete except for certain facets which may never be known. Dobzhansky (1937) and Arber (1934) present excellent detailed reviews of the process.

Essentially, S. townsendii is regarded as a hybrid between the endemic European S. maritima and the introduced and variable American S. alterniflora. The haploid chromosome number of S. maritima is 28, while that of S. alterniflora is 35. Fertilization may have resulted in the formation of a zygote with 63 chromosomes, which then doubled immediately to produce a plant with a somatic number of 126 chromosomes. Huskins (1931) has shown that with the somatic chromosome number for S. townsendii is 126. Of the many possible alternatives explaining the mechanism of origin of polyploidy in this case, the one presented above seems the most likely. The rather uniform nature of S. townsendii leads to the conclusion that hybridization has probably taken place but once. Further light is shed by the fact that S. townsendii has been shown,

according to Oliver (1925) and Arber (1934), to be vigorously aggressive both by means of fertile seed production and vegetative reproduction.

As noted above in the discussion of *S. alterniflora*, two levels of polyploidy exist within that species; the form with fifty-six chromosomes may be somewhat less vigorous than that with seventy. Observations on the variability of specimens of *S. alterniflora* from England, indicate that plants of both polyploid levels may be present there. Huskins (1931) has shown that the form with seventy chromosomes is present in England. This form seems to have been the one involved in the hybrid origin of *S. townsendii*.

The apparently stable polyploid is remarkably uniform in virtually all morphological features. Several of these show great usefulness as taxonomic key characters. Among these are ligules over 1.5 mm, spikelets over 15 mm, palea exceeding the lemma by 2 mm or more, and median spikes usually exceeding 10 cm. Other features which have been found useful but somewhat variable are the length of that part of the median spike rachis prolonged beyond the apex of the terminal spikelet, and length and width of the panicle. These features will be discussed in greater detail in the section of this paper dealing with the distinctions between species.

Certain anomalous plants have been found within this otherwise stable population. Chater and Jones (1951) describe two such anomalies. One is a dwarf form with much reduced spike and panicle length, fewer spikelets and brown color. The other is somewhat of a dwarf, differing from the normal in that the mid-portions of leaves contain a band of yellow tissue along the length of the blade. Chater and Jones do not provide nomenclatural status for these anomalies. They suggest that the anomalies may represent "disintegration" of the stable polyploid.

Nomenclature

The whereabouts of the holotype of *Spartina townsendii* is unknown. An isotype in the Kew Herbarium was not available for study. However, the description (H. and J. Groves, 1881) verifies the accuracy of typification.

Distribution

Fig. 10 shows the distribution of specimens examined. That the species is spreading rapidly by natural means is borne out by Arber (1934). *S. townsendii* is perhaps the most economically valuable member of the genus as attested by its introduction into many parts of the world for use in tideland reclamation.

Considerable literature dealing with the ecological relationships of *S. townsendii* has been published. Among the papers are those of Oliver (1925), Arber (1934), and Stapf (1908). The plant is largely one of the intertidal zone with a six-hour tolerance of submergence under salt water. It flowers from July to November and in November may often be found, according to Arber (1934), with sprouting seeds still clinging to the spikes. Of particular interest is its very aggressive nature by means of which it has invaded the territory once occupied by *S. maritima* and *S. alterniflora* and has obliterated them from the immediate area.

10. SPARTINA BAKERI

Spartina bakeri Merr. Bull. U.S. Dept. Agric. Pl. Ind. 9:14. (1902)

(Type in US from Lake Ola, Tangerine, Florida)

Spartina juncea Willd. var. bakeri (Merr.) St. Y. Cand. 5:91. (1932)

(Based on S. bakeri Merr.)

Description (Plate IV, a-b)

Culms to 2 meters tall, erect, terete, indurate, glabrous, in large, dense clumps from hard, knotty bases, rhizomes wanting; culms often with shoots from the lower nodes; sheaths smooth to striate, glabrous, the margin of the orifice glabrous; ligule a ring of hairs 0.5--2 mm long; blades elongate, usually involute, rarely flat, scabrous on the adaxial surface, glabrous on the abaxial, margins scabrous, tip acuminate; panicle 8--25 cm long; panicle axis 2--4-angled, flat, scabrous on the angles; spikes 3--16, 2--6 cm long, alternate, appressed, rarely spreading, moderately imbricate, sessile, the lower often short pedunculate; rachis of the spikes glabrous to sparingly scabrous on the angles; spikelets 10--30, 6--9 mm long, alternate, tightly appressed, closely imbricate; first glume linear, 3--6 mm long, shorter than one-half the second, keel hispid, margins sparingly hispidulous, tip acuminate; second glume 6--9 mm long, hispidulous, keel hispid, margins hispidulous, 2--3 lateral nerves, prominent, on one side of the second glume, tip acuminate; lemma glabrous, keel hispid, margins glabrous to very sparingly hispidulous, tip acute to obtuse or obscurely lobed; palea thin and papery, glabrous, tip acute to rounded, slightly exceeding the lemma; anthers 5 mm long; somatic chromosome number 42.

Observations

From the standpoint of morphology of the inflorescence, Spartina bakeri is hardly separable from S. patens. However, because of its unique vegetative habit and tolerance of fresh water it is quite distinct. Rhizomes are not produced as they are in S. patens. The culms grow in dense tufts up to several feet across, a feature which serves to set this species apart from its closely related congeners. While the inflorescence exhibits some variability in its parts, it is overlapped in every respect by S. patens. Consequently, herbarium specimens are extremely difficult of identification unless the collector has noted the peculiar habit on the label.

A characteristic feature of the inflorescence is the rather tight appression of the spikes. This feature was observed on all the specimens examined and is doubtless typical of the species. Its use as a good taxonomic key character is precluded by the presence of the same degree of appression by spikes of immature specimens of S. patens.

The only chromosome count reported for the species is that of Church (1940) wherein he gives 42 as the diploid number.

Nomenclature

Merrill (1902) recognizes Spartina bakeri as separable from other species of the genus on the basis of the unique vegetative habit. Unfortunately, the type specimen is without the basal portion of the culm and

underground parts. However, Merrill's description, evidently based upon additional plants and field observations by the collector (Baker), leaves no doubt as to the accuracy of the typification.

Saint-Yves (1932) treated the species as a variety of *Spartina juncea*. Doubtless *S. bakeri* is closely related to *S. patens* (*S. juncea*) and were it not for the complete absence of rhizomes and the ability of the plant to tolerate and invade fresh water sites, there would be little argument for its retention as a separate species.

Distribution

The species is quite successful in fresh water habitats. It is found along the sandy beaches and shores of many of Florida's inland lakes. Distributional data also show that it grows equally as well in salt water sites in Florida and southern Georgia. Collecting dates point to a rather erratic flowering by this species. Unlike other North American *spartinas*, this species flowers for the most part from December to May, but is occasionally found in flower during the summer.

11. SPARTINA X CAESPITOSA

Spartina X caespitosa (A.A. Eaton) Fern. Gray's Man. Bot. ed. 8.

180. (1950) (Based on *S. caespitosa* A.A. Eaton)

Spartina caespitosa A.A. Eaton. Bull. Torrey Club. 25:338. (1898)

(Type in GH from Seabrook, N.H.)

Spartina patens var. *caespitosa* (A.A. Eaton) Hitchc. Rhod. 8:210.

(1906) (Based on *S. caespitosa* A.A. Eaton)

Trachynotia caespitosa Ait. in St. Y. Cand. 5:92. (Error for

S. caespitosa A.A. Eaton)

Description (Plate IV, c-d)

Culms to 1.2 meters tall, slender, erect, terete, glabrous, in dense caespitose clumps without rhizomes or with short rhizomes and with short, extra-vaginal shoots from the base, or solitary from widely spreading, thick rhizomes, scales closely imbricate; sheaths smooth to striate, glabrous, margin of the orifice glabrous to short-pilose; ligule a ring of hairs 0.5--1.0 mm long, rarely longer; blades elongate, involute or flat, narrow, glabrous or scabrous on the adaxial surface, glabrous on the abaxial, margins glabrous or scabrous, tip acuminate; panicle 9--20 cm long; panicle axis 3-angled, scabrous on the angles; spikes 3--9, 3--9 cm long, alternate, appressed or spreading, usually pedunculate; rachis of the spikes scabrous on the angles; spikelets 20--50, 10--17 mm long, alternate, tightly appressed, closely imbricate; first glume linear 4--9 mm long, glabrous to sparingly hispidulous, keel hispid, tip acuminate or awned; second glume 10--17 mm long, glabrous to sparingly hispidulous, keel hispid to pectinate, lateral nerves prominent, one on either side of the keel, or 2--3 on one side of the second glume, tip acuminate or awned; lemma glabrous to sparingly hispidulous, keel glabrous or hispid only at the tip or hispid to pectinate on the upper three-fourths, margins usually hispidulous, tip obtuse, rounded, obscurely lobed, or apiculate; palea thin and papery, glabrous, bilobed and obtuse or rounded

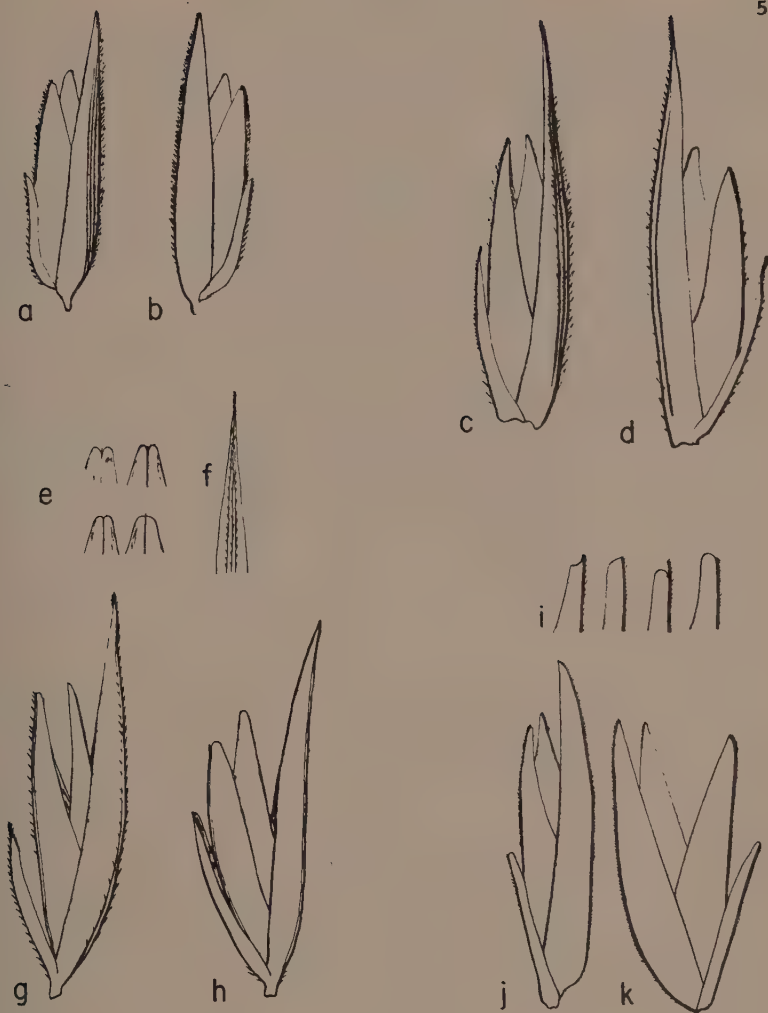


PLATE IV

a-b. *Spartina bakeri*.

c-d. *Spartina X caespitosa*.

e-h. *Spartina cynosuroides*.

i-k. *Spartina densiflora*.

a. spikelet showing lateral nerves 7.5X,

b. same spikelet, opposite view 7.5X.

c. spikelet showing two lateral nerves 7.5X,

d. spikelet showing one lateral nerve 7.5X.

e. lemma tips, keel view 3.75X,

f. second glume, keel view 3.75X,

g, h. spikelets 3.75X.

i. first glume tips 7.5X, j-k. spikelets 3.75X.

at the tip, slightly exceeding the lemma; anthers 3--6 mm long; chromosome number 42.

Observations

The taxonomic position of S. caespitosa has been the subject of some controversy since it was first described by Eaton (1898). Hitchcock (1906) reduced it to varietal rank under S. patens. Swallen (1939) retained it as a valid species as did Chase (1950). Fernald (1950) relegated the plant to a hybrid category, suggesting that it showed a combination of characters from both S. pectinata and S. patens.

Church (1940) had earlier hypothesized a hybrid origin based upon cytological analysis and apparent morphological similarities to both S. pectinata and S. patens. The cytological data presented by Church show identical chromosome numbers for the species involved. S. pectinata from New England was shown to have a somatic number of 42, identical to that found for the putative hybrid. From these data he concluded that crossing must have occurred between S. pectinata and the hexaploid segment of the S. patens population. While this hexaploid segment has been considered taxonomically separable from the tetraploid, evidence presented below in connection with S. patens shows that the two are not at all discrete, but that they grade imperceptibly into one another so as to defy separation. Similarly, within S. pectinata, the existence of a series of polyploids, not morphologically separable from one another, is reported.

In an effort to test the validity of Church's hybrid hypothesis a hybrid index was constructed according to methods described earlier in connection with S. longispica (page 502). Because the polyploids within each parent species are not morphologically separable from one another, all available specimens of both parents were used to ascertain ranges of variability in the characters utilized. The discussions of those characters possessing measurable variability are accompanied by tables showing the range of variability within both the parent species. The total number of specimens of each species introduced into the tables will be seen to vary from table to table. This variation occurs because certain structures were missing on some of the specimens. Many more specimens were used in assessing variability than were eventually scored by means of the hybrid index because some specimens, otherwise perfectly identifiable, did not possess all the morphological parts necessary for complete scoring.

The hybrid appears to have only a sporadic distribution along the east coast of the United States from New Hampshire to Maryland. Accordingly, only specimens from this particular region were used to determine ranges of variability within the parent populations. The twenty characters used in constructing the hybrid index are presented below:

1. The rhizomes in S. pectinata are thick and hard, possessing closely imbricated cales. Those of S. patens are thin and wiry with scales scarcely imbricate. An intermediate condition was not found. The hybrids either do not possess rhizomes or have rhizomes resembling those of S. pectinata. In cases where rhizomes are wanting, the intermediate index value of 1 is assigned.

2. The rhizomes of S. pectinata are distinctly brownish-purple in color while those of S. patens are whitish. Again, as in the preceding feature, the rhizomes of the hybrids, when present, are like those of S. pectinata. In instances where rhizomes are wanting, an index value of 1 is assigned.

3. Table 6 shows the distribution of flowering culm heights for specimens examined. Preliminary investigations in the hybrid swarm showed that heights of culms averaged about 1 meter. Accordingly, all specimens which were found in the range of overlap (81--120 cm) between the parents were assigned the index value of 1.

Table 6. Distribution of Culm Heights by Classes

Distribution classes (cm)	<u>S. pectinata</u>	<u>S. patens</u>	<u>S. X caespitosa</u>
0--10	0	0	0
11--20	0	2	0
21--30	0	18	0
31--40	0	40	0
41--50	0	29	0
51--60	0	28	0
61--70	0	20	2
71--80	0	17	6
81--90	0	12	5
91--100	3	3	6
101--110	2	1	6
111--120	9	6	5
121--130	3	0	4
131--140	11	0	0
141--150	5	0	0
151--160	6	0	0
161--170	1	0	0
171--180	0	0	0
181--190	2	0	0
191--200	0	0	0
201--210	0	0	0
211--220	0	0	0
221--230	1	0	0
231--240	0	0	0

4. The leaf blades of S. pectinata are usually flat, becoming involute toward the apex. Those of S. patens are commonly involute throughout. An intermediate condition was not discernible.

Table 7. Distribution of Second Blade Widths and Lengths by Classes

Distribution Classes	<u>S. pectinata</u>	<u>S. patens</u>	<u>S. X caespitosa</u>
<u>Second Blade Widths</u>			
0--0.9 mm	0	65	0
1--1.9	0	67	0
2--2.9	0	27	7
3--3.9	0	6	11
4--4.9	0	3	13
5--5.9	2	0	1
6--6.9	6	2	2
7--7.9	23	0	0
8--8.9	10	0	0
9--9.9	11	0	0
10--10.9	8	0	0
11--11.9	1	0	0
12--12.9	3	0	0
13--13.9	1	0	0
<u>Second Blade Lengths</u>			
0--3.9 cm	0	0	0
4--7.9	0	28	0
8--11.9	0	49	1
12--15.9	0	32	1
16--19.9	0	14	2
20--23.9	0	18	7
24--27.9	0	8	5
28--31.9	0	3	7
32--35.9	1	3	4
36--39.9	4	1	2
40--43.9	3	0	2
44--47.9	1	0	0
48--51.9	7	0	1
52--55.9	7	0	2
56--59.9	5	0	0
60--63.9	5	0	0
64--67.9	8	0	0
68--71.9	3	0	0
72--75.9	2	0	0
76--79.9	0	0	0
80--83.9	0	0	0
84--87.9	2	0	0
88--91.9	1	0	0
92--95.9	1	0	0

5. Leaf width in the putative parents shows considerable variation. Table 7 shows the distribution of leaf width in the specimens examined. All specimens in the range 4--6 mm were scored with the intermediate value of 1. Measurements were made on the second leaf from the summit.

6. Length of the blades does not separate the parents discretely. However, Table 7 shows that considerable difference exists between the species with respect to blade length. All specimens in the range 24--39.9 cm were assigned the intermediate value of 1. Measurements were made on the second leaf from the summit.

7. Length of the ligule serves to separate the parent species discretely. Ligules in *S. pectinata* vary from 1--3 mm. *S. patens* has ligules which rarely exceed 0.5 mm. Those specimens with ligules exceeding 0.5 mm but less than 1 mm were assigned the intermediate value of 1.

Table 8. Distribution of Panicle Lengths by Classes

Distribution classes (cm)	<i>S. pectinata</i>	<i>S. patens</i>	<i>S. X caespitosa</i>
0--1.9	0	0	0
2--3.9	0	4	0
4--5.9	0	32	0
6--7.9	0	47	0
8--9.9	0	29	3
10--11.9	0	19	4
12--13.9	0	21	3
14--15.9	0	20	7
16--17.9	1	3	8
18--19.9	2	5	5
20--21.9	6	3	3
22--23.9	8	0	1
24--25.9	11	0	0
26--27.9	11	0	0
28--29.9	10	0	0
30--31.9	15	0	0
32--33.9	7	0	0
34--35.9	11	0	0
36--37.9	4	0	0
38--39.9	2	0	0
40--41.9	0	0	0
42--43.9	1	0	0

8. Considerable difference in panicle length is evident between the parent species. Panicles in S. pectinata vary from 16--44 cm in length, while those of S. patens range from 2--22 cm in length. Table 8 shows the distribution of panicle lengths in both species. Specimens with panicle lengths within the range of 16--22 cm were assigned a value of 1.

9. Table 9 shows the distribution of spike numbers in both parent species. In cases where spike number was 7--9, these specimens were assigned a value of 1.

Table 9. Distribution of Spike Numbers by Classes

Distribution classes	<u>S. pectinata</u>	<u>S. patens</u>	<u>S. X caespitosa</u>
1	0	1	0
2	0	76	0
3	0	52	4
4	0	24	11
5	0	19	8
6	0	6	2
7	3	3	5
8	5	0	2
9	6	1	2
10	5	1	0
11	9	0	0
12	7	0	0
13	9	0	0
14	8	0	0
15	7	0	0
16	5	0	0
17	4	0	0
18	4	0	0
19	4	0	0
20	3	0	0
21	1	0	0
22	3	0	0
23	3	0	0
24	0	0	0
25	0	0	0
26	3	0	0
27	3	0	0

10. Mean spike lengths of the two parent species differ somewhat although the variability within the specimens examined shows some overlap. Consequently, delimitation of an intermediate does not seem advisable here. Table 10 shows the distribution of spike lengths for each parent species. A value of 55 mm was selected as an arbitrary division point. Accordingly, specimens with spike lengths of 55 mm or less (typical of S. patens) were assigned the index value of 2, while those exceeding 55 mm (typical of S. pectinata) were given a value of 0.

Table 10. Distribution of Spike Lengths by Classes

Distribution classes (mm)	<u>S. pectinata</u>	<u>S. patens</u>	<u>S. X caespitosa</u>
0--5	0	0	0
6--10	0	0	0
11--15	0	1	0
16--20	0	4	0
21--25	0	19	0
26--30	0	28	0
31--35	0	32	1
36--40	0	19	6
41--45	2	20	6
46--50	2	21	4
51--55	3	10	4
56--60	5	9	5
61--65	8	2	4
66--70	10	3	2
71--75	10	1	2
76--80	10	1	0
81--85	14	0	0
86--90	8	0	0
91--95	7	0	0
96--100	2	0	0
101--105	2	0	0
106--110	0	0	0
111--115	1	0	0
116--120	2	0	0
121--125	2	0	0
126--130	0	0	0
131--135	0	0	0
136--140	1	0	0

11. Peduncle lengths appear to differ significantly between the two parent species. Lengths of the peduncles of median spikes in S. patens were not found in excess of 5 mm while the value in S. pectinata commonly exceed 5 mm. No intermediate was defined here. The hybrids exhibited considerable variability in this respect.

12. Longer spikelets with a range of 11--21 mm appear to be more typical of S. pectinata while shorter spikelets with a range of 6--14.9 mm characterize S. patens. Data presented in Table 11 show an overlap between 11 and 14.9 mm. Consequently, any spikelet having a length within that intermediate range was assigned a value of 1.

Table 11. Distribution of Spikelet Lengths by Classes

Distribution classes (mm)	<u>S. pectinata</u>	<u>S. patens</u>	<u>S. X caespitosa</u>
6--6.9	0	3	0
7--7.9	0	20	0
8--8.9	0	40	0
9--9.9	0	50	0
10--10.9	0	28	4
11--11.9	2	17	7
12--12.9	3	7	9
13--13.9	7	3	4
14--14.9	9	2	6
15--15.9	12	0	3
16--16.9	20	0	0
17--17.9	13	0	1
18--18.9	11	0	0
19--19.9	8	0	0
20--20.9	2	0	0
21--21.9	1	0	0
22--22.9	0	0	0
23--23.9	0	0	0
24--24.9	0	0	0
25--25.9	0	0	0
26--26.9	1	0	0

13. The first glume of S. patens is commonly shorter than one-half the length of the second glume. In contrast, first glumes of S. pectinata are almost always longer than one-half the length of the second. Only in cases where the length of the first glume was found to be exactly one-half that of the second was the intermediate value of 1 assigned.

14. Considerable difference in length of first glumes was found between the parents. Table 12 shows variation from 2--5 mm for S. patens and from 5--12 mm for S. pectinata. Values from 5--5.9 mm are assigned the intermediate value of 1.

Table 12. Distribution of First Glume Lengths by Classes

Distribution classes (mm)	<u>S. pectinata</u>	<u>S. patens</u>	<u>S. X caespitosa</u>
2--2.9	0	5	0
3--3.9	0	36	1
4--4.9	0	19	6
5--5.9	2	1	15
6--6.9	10	0	8
7--7.9	13	0	4
8--8.9	17	0	0
9--9.9	14	0	0
10--10.9	7	0	0
11--11.9	2	0	0
12--12.9	2	0	0

15. The tip of the first glume in S. pectinata is typically awned, while that of S. patens is merely acuminate. Because of the difficulty in determining how much of the tip is actually awn, no intermediate was defined here.

16. As with the preceding, the same differences are noted between the parents with respect to the tip of the second glume.

17. The second glume of S. pectinata is typically 1--3-nerved. When three nerves are evident, the two lateral nerves are disposed, one on either side of the prominent mid-nerve (keel). S. patens second glumes are 3-nerved and the two lateral nerves are both disposed on only one side of the prominently pubescent mid-nerve (keel).

18. Pubescence on the keel of the second glume in S. pectinata is very strongly pectinate while that of S. patens is hispid.

19. As with the preceding, the same differences are noted between the parents with respect to pubescence on the keel on the lemma.

20. The tip of the lemma in S. pectinata is bilobed and each lobe is distinctly acuminate, while in S. patens the tip is obtuse, rounded or obscurely bilobed (Plate V, j-m).

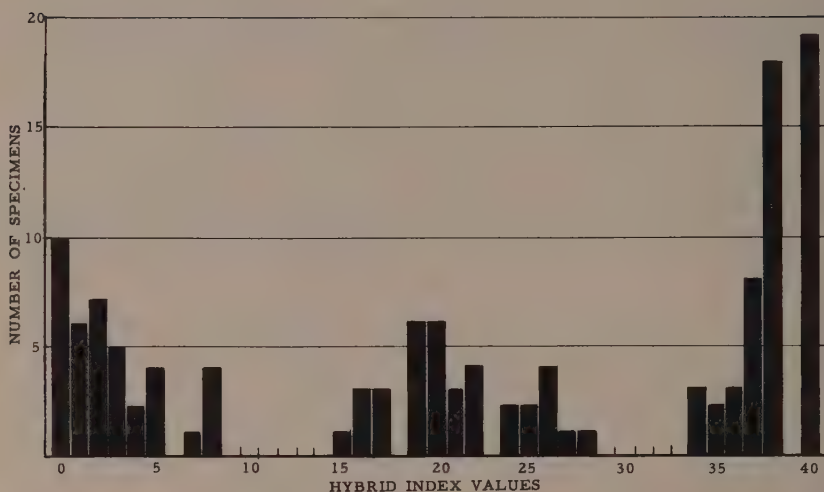


Fig. 11. Distribution of Hybrid Index Total Scores

Results of the scoring of 128 specimens are shown in Fig. 10. The putative hybrids are represented by the intermediate cluster, values 15 to 28. It appears from the data presented that the hypothesis of hybrid origin of *S. caespitosa* is further substantiated.

For items 3, 5, 6, 8, 9, 10, 12, and 14, tables are presented which show the variability within the putative parent populations. After the scoring of parents and hybrids was completed, measurement data from plants of the hybrid swarm were tabulated. These data are shown in the fourth column of each of the tables.

While the intermediate specimens appear to be members of a population which is discrete from either putative parent, there exists within this intermediate cluster considerable variability especially with regard to rhizomes. Field observations show that the plants without extensive rhizomes grow in large, dense caespitose clumps often as much as 50 cm across. From observations on herbarium specimens which fall within the intermediate cluster on Fig. 11 but possess extensive rhizomes, it appears that the plants do not have a true caespitose habit. Quite to the contrary, they appear to arise as solitary upright culms, much as in *S. pectinata*. Certain plants of caespitose habit were observed to possess occasional short rhizomes in addition to the numerous, short, extra-vaginal shoots.

As Blomquist (1948) has suggested, perhaps the caespitose habit may be merely an ecological form. However, field observations do not bear this conclusion out. A visit to the type locality, an abandoned seaside pasture one mile east of the town of Seabrook, New Hampshire, proved quite rewarding. At that station, *S. caespitosa* of the true caespitose habit was found abundantly. Growing very near it on the same disturbed soil were large numbers of rhizome-producing specimens of *S. pectinata*

and S. patens. If the production of rhizomes was a mere response to environmental conditions, one would certainly have expected to find rhizomatous S. caespitosa at that station. That only the caespitose form occurs there was borne out by extensive observations that required the uprooting of virtually every clump of S. caespitosa in the immediate vicinity.

A more likely explanation for the variability of underground structures is that hybridization between S. pectinata and S. patens may occur repeatedly and that the capacity for rhizome production may be controlled by hereditary mechanisms. Certainly, conditions favoring hybridization are present throughout the known range of S. caespitosa. The flowering periods of the putative parents are coincident. They grow in the same habitat, often together, and they both vary with regard to polyploidy. These factors, coupled with the results of scoring by means of the hybrid index and the sporadic distribution, lend considerable support to the hypothesis that the intermediate plants are actually members of a polyphyletic hybrid swarm standing intermediate between S. patens and S. pectinata. That crossing of the putative parents does produce viable seeds has been demonstrated experimentally by Church (1940). However, his seeds failed to develop beyond the seedling stage.

If such crosses be of frequent natural occurrence, opportunity might be offered for a rather wide variety of character segregation. The genetic mechanisms which may control the production of rhizomes are not known, but it is entirely possible that such mechanisms may account for the variability evident in regard to the rhizomes.

The true caespitose forms may themselves be polyphyletic. The fact that some caespitose clumps seem to produce rhizomes might indicate that a separate cross was involved in their formation.

Among the clumps examined at the type locality, a distinct difference was noted in one. It was unlike any of the others in that its leaves were completely involute, quite as leaves of S. patens, while the others possessed flat leaves more typical of S. pectinata. The presence of this unique clone may further indicate that hybridization is of frequent occurrence. These specimens (Mobberley 249, ISC) are very similar to the type specimens of Eaton.

Further evidence supporting that polyphyletic origin theory may be found in the differences manifested by the hybrid specimens in regard to certain features typical of the putative parents. For example, the character in item 17 of the hybrid index above is perfectly stable for each parent, but the hybrids vary considerably, some resembling S. patens, others S. pectinata. While there is no evidence supporting a view that the hybrid population is capable of maintaining itself by breeding true, field observations indicate that the caespitose form without rhizomes is capable of extensive vegetative reproduction. Several clumps distributed across a small abandoned dump at the type locality are virtually identical. Considerable vegetative reproduction is evident in the caespitose clumps themselves where new culms are seen to arise from the many short, extra-vaginal shoots.

While the hybrid swarm doubtless has some difficulty increasing its distribution in competition with its more aggressive congeners, there appears to be little doubt that it is persistent.

Nomenclature

The species was first described by Eaton (1898) from a specimen collected in a seaside pasture near Seabrook, New Hampshire. The type specimen is of the caespitose form and shows no evidence of rhizomes.

Hitchcock (1906) reduced the species to S. patens var. caespitosa on the grounds that it was identical with S. patens except for the caespitose habit. However, as shown in the hybrid index above, S. patens and S. caespitosa are separable on more counts than simple habit. Eaton's type, upon which Hitchcock's variety was based, was scored according to the hybrid index and was found to have a total score of 21.

Fernald (1950), having seen morphological features which obviously related S. caespitosa to both S. patens and S. pectinata, reduced the species to a hybrid category. Evidence presented above would appear to support the hybrid hypothesis, but it is apparent that the members of the hybrid swarm are not all of the same ancestry. In accordance with

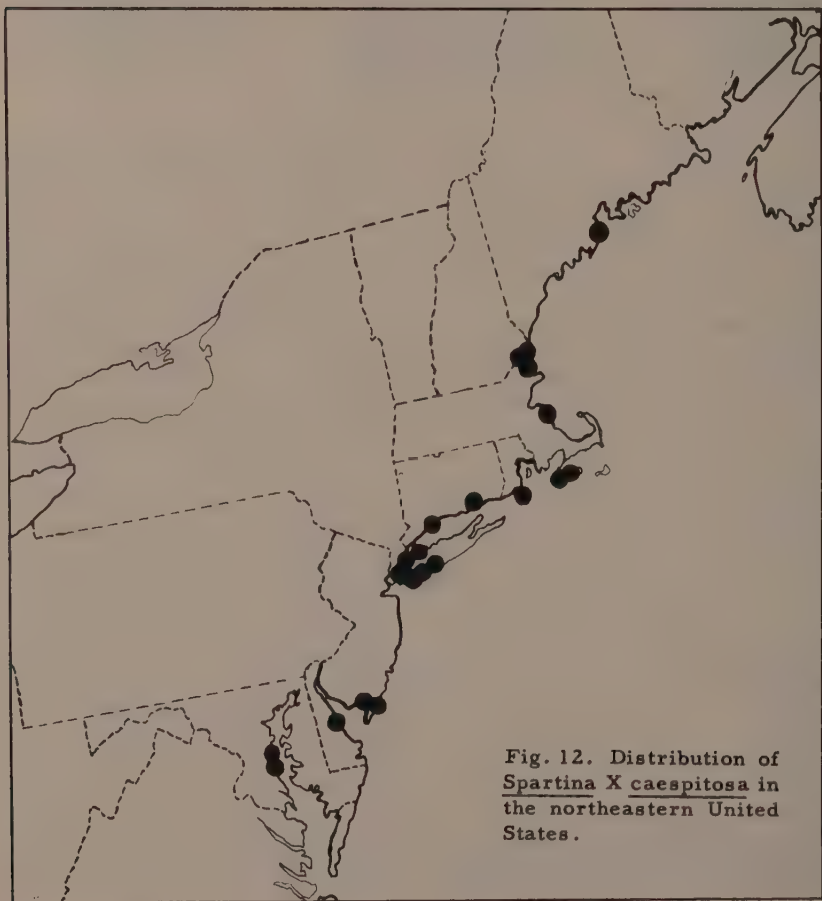


Fig. 12. Distribution of Spartina X caespitosa in the northeastern United States.

Article H.1 of the International Code of Botanical Nomenclature (1952), the hybrid swarm is designated Spartina X caespitosa until more information is available. It may be possible to show that individual segments of the hybrid swarm are capable of maintaining themselves as discrete taxonomic entities and might therefore be regarded as distinct species.

Distribution

Fig. 12. shows the distribution of specimens examined. Specimens referable to the hybrid swarm by means of the hybrid index seem to occur only where the ranges of S. pectinata and S. patens are coincident.

That Spartina X caespitosa grows in the same habitat as its putative parents is significant. The habitats are mostly the drier portions of salt and brackish marshes, in areas some distance above the intertidal zone. At the type locality, clumps of the hybrid were found within 50 feet of the water's edge at high tide. Perhaps the most significant feature concerning this hybrid is that it was not found on any but disturbed soil. At the type locality, the part of the seaside pasture in which these plants were found had obviously been used as a dump, for the author encountered all sorts of household refuse during the collection of underground portions of the caespitose clumps. None of the hybrids were found on ground that had not been disturbed, although the parents seemed to flourish on this dump as well as on the undisturbed ground nearby. An illuminating note is appended to the label on a hybrid specimen (Eames 4071, Bridgeport, Conn., GH) to the effect that the specimens was collected from "made ground" at the edge of a salt marsh. The prevalence of a number of intermediate specimens in and near New York City, where little if any ground has likely remained undisturbed, might be considered further evidence supporting the hybrid hypothesis. As Anderson (1949) has pointed out, the success of naturally occurring hybrids seems to be enhanced when the environment particular to the parent species is in some way disturbed.

12. SPARTINA CYNOSUROIDES

Spartina cynosuroides (L.) Roth. Cat. Bot. 3:10. (1806)

Dactylis cynosuroides L. Sp. Pl. ed.1. 71. (1753)

(Type in BM from Virginia)

Trachynotia polystachya Michx. Fl. Bor. Amer. 1:64. (1803)

(Type in P from South Carolina)

Limnethis polystachia Rich. in Pers. Syn. Pl. 1:72. (807)

Spartina cynosuroides (L.) Willd. Enum. Pl. 81. (1809) pro parte.

Spartina polystachya (Michx.) Beauv. Nouv. Agrost. 25. (1812)

Cynodon cynosuroides (L.) Rasp. Ann. Sci. Nat. Paris I. 5:303. (1825)

Spartina cynosuroides Willd. var. polystachya (Willd.)

Scribn. Bull. Torrey Club. 10:86. (1883)

Spartina cynosuroides (L.) Roth var. polystachya (Michx.) Beal ex

Fern. Rhod. 49:113. (1947)

Description (Plate IV, e-h)

Culms to 3.5 meters tall, erect, stout, terete, glabrous, solitary from stout, widely spreading rhizomes, scales closely imbricate; sheaths smooth to striate, the lower often wrinkled, glabrous, the margin of the orifice often densely pilose; ligule a ring of hairs 1--3 mm long; blades elongate, flattened, glabrous on both surfaces, margins scabrous, tip acuminate; panicle 15--40 cm long; panicle axis 3--4-angled, prominently contorted on the more robust specimens, glabrous to scabrous on the angles; spikes 5--67, 6--15 cm long, alternately to irregularly arranged, spreading, occasionally appressed, imbricate, sessile or pedunculate, the peduncles as much as 4.5 cm long; rachis of the spikes glabrous to scabrous on the angles; spikelets 10--70, 9--14 mm long, alternate, tightly appressed, closely imbricate; first glume linear, 3--7 mm long, usually less than one-half the length of the second, glabrous, keel strongly hispid, margins hispidulous, tip acute; second glume 9--14 mm long, glabrous to hispidulous, keel hispid, lateral nerves prominent, one on either side of the keel, nerves hispid, margins of the glume hispidulous, tip acuminate; lemma glabrous to sparingly hispidulous, often only on the upper half, keel hispid, tip obtuse or rounded; palea thin and papery, often hispidulous near the summit of the keel, tip acuminate, exceeding the lemma; anthers 4--6 mm long; somatic chromosome numbers 28, 42.

Observations

S. cynosuroides is a rather uniform species, although less robust plants may resemble, superficially at least, vigorous specimens of *S. patens*. The plant is essentially a coarse halophyte which was observed to reach a height of 3.5 meters in one locality (Pungo, Va. Mobberley 209, ISC). The vegetative parts of the plant, in general, exhibit much more vigor than similar parts in the other species within this complex. Considerable variation was found in measured values for certain vegetative parts commonly used as key characters. Ligule length was found to be quite variable, not differing significantly in this respect from most other species of *Spartina*.

The more important characteristics from the standpoint of plant identification are spike number, spike length, peduncle length, spikelet length, relative glume lengths, and tips of glumes and lemmas.

Reports concerning spike number in this species are somewhat at variance. Fernald (1947, 1950) states that in the maritime variety, up to 100 spikes may occur on a single panicle. However, during the examination of over three hundred panicles from across the range of distribution, the greatest number of spikes encountered on any one panicle was sixty-seven. On the other hand, Small (1933) reported the maximum number of spikes to be about fifty. This value is shown to be slightly less than the number encountered in specimens collected by the author from southeastern United States.

A character of this species which appears to be of some taxonomic significance is the position of the nerves on the second glume. Two lateral nerves are usually prominent, one on either side of the keel. Very often the lateral nerves are hispid. This character is illustrated in Plate IV, f.

Two specimens (Chapman, Fla. MO) geographically identified only as "Florida" are almost completely glabrous along nerves and keels of all parts of the spikelet (Plate IV, h). However, this feature is not sharply delimited, for a number of specimens, particularly those from the Gulf Coast area, show a tendency to be less copiously hispid than others from more northerly stations.

One chromosome count has been previously reported. Church (1940) found this species to have the hexaploid number of 42 from a station in East Dennis, Massachusetts. During the present investigation, chromosome numbers of both 28 and 42 were found. These findings are presented in Table 13.

Table 13. Chromosome Numbers in S. cynosuroides

Mobberley 55	Cypremort Point, Louisiana	2n = 42
Mobberley 158	St. Mary's, Georgia	2n = 28

Plants found to have the tetraploid number of chromosomes were critically compared with the hexaploids. No morphological differences were found that would permit a satisfactory taxonomic separation of the two. Differences may exist in certain anatomical features such as guard cell size, epidermal cells, and the like. However, these were not studied during the present investigation. Not nearly enough specimens with chromosome numbers known are available to make a more satisfactory disposition of the apparent differing levels of polyploidy.

Nomenclature

This species, named by Linnaeus (1753) as Dactylis cynosuroides, was based upon a plant collected in Virginia by Clayton. The original description was that of Gronovius (1743). Linnaeus gave this plant the specific epithet cynosuroides and appended a variety β which is clearly identified as S. alterniflora. Subsequently, a number of European authors, among them Loeffling (1758) and Michaux (1803), took up the specific epithet and applied it mistakenly to other species within the genus. Loeffling misapplied the epithet to the European S. maritima and Michaux to the American S. pectinata. Michaux then settled on the epithet polystachya for the present species.

Richard (1807), apparently without the knowledge of Schreber's earlier description of Spartina, transferred Michaux's Trachynotia and the spartinoid segment of Linnaeus' Dactylis to the genus Limnethis. Limnethis polystachya was obviously an orthographic variant of Limnethis polystachya. Hitchcock (1953) gives Limnethis cynosuroides in synonymy for Spartina cynosuroides, but it appears that Limnethis cynosuroides of Richard is actually Michaux's Trachynotia cynosuroides, now shown to be a synonym of Spartina pectinata.

Roth (1806) correctly identified the true cynosuroides of Linnaeus with the genus Spartina. His combination seems to merit recognition as the valid one. Roth does not cite polystachya at all and thus it remained for Beauvois (1812) to effect the transfer of Trachynotia polystachya to Spartina. It stands now as a synonym of S. cynosuroides.

Scribner (1888) concluded upon examination of several anomalous spikelets that the present species was merely a variety of the more widely distributed transcontinental *S. cynosuroides* Willd. The erroneous usage of the epithet *cynosuroides* for *S. pectinata* is reviewed in connection with the synonymy of that species. Scribner held that because some spikelets of the more southern coarse halophyte were found with atypically long first glumes, the entire species should be regarded as a variety.

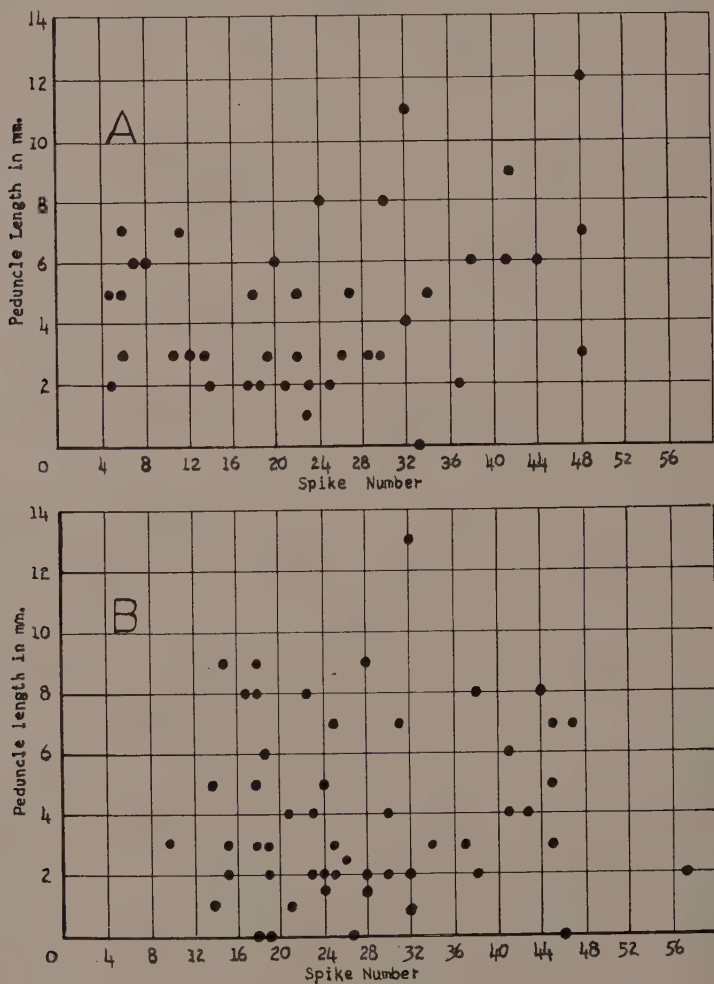
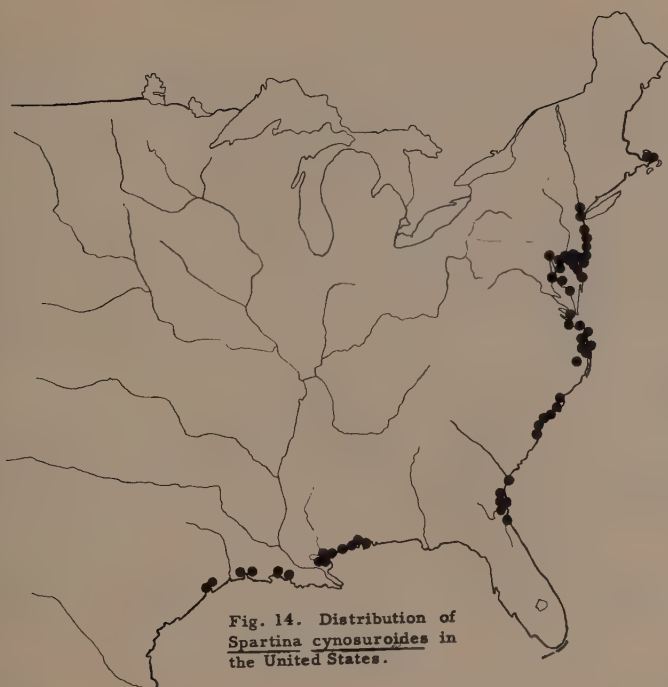


Fig. 13. Comparison of Specimens of *Spartina cynosuroides* from Maritime and Brackish Water Habitats by Spike Number and Peduncle Length.

A. Maritime Habitat.

B. Brackish Water Habitat.



Beal (1896) favored Scribner's varietal rank for the present species. However, Fernald (1947) used *Spartina cynosuroides* (L.) Roth var. *polystachya* (Michx.) Beal to name what he regarded as a distinct variety of the southern halophyte. Fernald sought to distinguish var. *polystachya* from the typical variety on the basis of its exclusively maritime habit, and 30--100 short-pedunculate to subsessile spikes. The typical variety is supposed to have 6--50 spikes, longer peduncles (no values are given) and a fresh to brackish water habitat.

Fernald's observations were made principally in Princess Anne County, Virginia, where he found the two varieties supposedly quite selective of habitat. He also quotes Bayard Long to the effect that the two varieties are similarly disposed in New Jersey, Delaware, and eastern Pennsylvania. It is significant to note that Pohl (1947) in the most recent treatment of Pennsylvania grasses does not cite the present species as occurring anywhere in Pennsylvania. Chase (1950) states that habitat is not satisfactorily correlated with morphological distinctions.

In an effort to determine whether the variety is actually distinct from the typical form, extensive collections from both brackish water and maritime habitats were made by the author, particularly in Princess Anne County, Virginia. The collections were studied from the standpoint of spike number and peduncle lengths. Peduncle measurements were made on a median spike. Results of the investigation are presented in Fig. 13, where spike number is plotted against peduncle length. Two

diagrams were constructed, one for fresh to brackish water plants and the other for strictly maritime plants.

The scatter diagram shows that scarcely one-third of the maritime specimens examined answered to Fernald's spike number criterion. Many specimens with fewer than 30 spikes were found in maritime habitats. The diagram also shows that a considerable number of plants of brackish to fresh water habitat possess peduncles much shorter than those found on many of the strictly maritime specimens. The failure of dots, each representing a single specimen, to become segregated into two discrete clusters would seem to cast doubt upon the presence of two distinguishable populations. In view of the foregoing evidence, it appears desirable to regard the varietal name as a synonym of S. cynosuroides.

Distribution

Distribution of Spartina cynosuroides is shown in Fig. 14. While the plant has been observed in strictly maritime habitats bordering the strand and intertidal zones, the primary habitats seem to be the edges of brackish water estuaries and borders of tidal lagoons and bays. A particularly favored site for the species is the margin of the intercoastal waterway. Collection dates indicate that the plant flowers from June to October. Quite in contrast to the more succulent S. alterniflora, the inflorescence of this species is remarkably free from insect predation, doubtless because of the harsh, indurate texture of the spikelets.

13. SPARTINA DENSIFLORA

Spartina densiflora Brong. Du Perr. Bot. Voy. Coq. 14. (1829)

(Type in P from Concepción, Chile)

Chauvinia chilensis Steud. Syn. Pl. Glum. 362. (1855)

(Type in P from Concepción, Chile)

Spartina montevidensis Arech. Anal. Mus. Nac. Montevideo. 4:378.

(1896) (Type from Rio de La Plata shore in Uruguay)

Spartina patagonica Speg. Rev. Agron. La Plata. 579. (1897)

(Type from Santa Cruz, Argentina)

Spartina densiflora Brong. var. typica St. Y. Cand. 5:74. (1932)

Spartina densiflora Brong. var. typica St. Y. subv. brongniartii St. Y.

Cand. 5:74. (1932) (Based on S. densiflora Brong. and Chauvinia chilensis Steud.)

Spartina densiflora Brong. var. typica St. Y. subv. pauper St. Y.

Cand. 5:77. (1932) (Based on S. patagonica Speg.)

Spartina juncea Willd. var. montevidensis (Arech.) St. Y. Cand.

5:89. (1932) (Based on S. montevidensis Arech.)

Spartina juncea Willd. var. laxiflora St. Y. Cand. 5:88. (1932)

(Type in G from Jacarehy, Brazil)

Spartina platensis Mez. ex St. Y. Cand. 5:86. (1932) nomen, in obs.

Description (Plate IV, i-k)

Culms to 1.5 meters tall, erect, terete, indurate, glabrous, caespitose from hard knotty bases, rarely with short rhizomes, commonly with short extra-vaginal shoots appressed to the culm; sheaths smooth to

somewhat striate, glabrous, the lower indurate and shining, the upper dull and somewhat striate, margin of the orifice glabrous to moderately pilose; ligule a ring of hairs 1--2 mm long; blades elongate, 3--8 mm wide, involute when fresh, commonly scabrous on the adaxial surface, glabrous on the abaxial, margins scabrous, tip acuminate; panicle 10--30 cm long, 4--8 mm wide; panicle axis 3-angled, scabrous on the angles, often twisted; spikes 2--15, 1--11 cm long, 2--3 mm wide, the longer narrow, the shorter broad and often flattened dorsiventrally, alternate, tightly appressed, moderately imbricate, sessile; rachis of the spikes glabrous to scabrous on the angles, not prolonged beyond the terminal spikelet; spikelets 10--30, 8--14 mm long, 1--2 mm wide, the lower usually the longer, alternate, tightly appressed, moderately to closely imbricate; first glume linear, 4--7 mm long, glabrous to sparingly hispidulous, keel hispidulous, margins glabrous to sparingly hispidulous, tip usually obtuse; second glume 8--14 mm long, sparingly hispidulous, keel hispidulous, margins sparingly hispidulous, tip usually acuminate; lemma minutely hispidulous, keel glabrous below, hispidulous above, tip acuminate to obtuse; palea thin and papery, keel glabrous below, hispidulous above, tip acuminate, exceeding the lemma; anthers 3--5 mm long.

Observations

The above description is based upon herbarium specimens from Chile and Argentina which had previously been referred to S. densiflora and upon specimens from Brazil, Uruguay, and Argentina which had been referred to S. montevidensis.

The vegetative parts of the plant are rather uniform throughout the population. The plants were not found to exceed 150 cm in height on any of the sheets. The only vegetative characters of taxonomic significance are the short, slender, upturned shoots arising from the hard, knotty culm bases. The ligule of this species does not differ significantly from that present in most of its congeners.

The important characters of the inflorescence are lengths of the panicles, spikes and spikelets as well as the number of spikes and spikelets. Chilean specimens commonly referred to S. densiflora are distributed across the range of variation in all features except spike lengths and spike number. These two features will be discussed in connection with geographic distribution.

Several other features were somewhat characteristic for the population but these are of doubtful taxonomic significance. They were, however, useful in connection with studies on the putative hybrid origin of S. longispica. Among these features are the relative lengths of the two glumes, the first generally being shorter than one-half the length of the second. As will be pointed out in the discussion of synonymy, this feature was not found to be consistent, but represents only a general trend in the population. The same thing may be said with regard to the glume tips. Plate IV, i-k, illustrates the more common condition found within the specimens examined. The acuminate tip of the palea, although not entirely consistent among the specimens examined, offers a distinctive feature. Spikelets are always readily disarticulated from the rachis. Perhaps not too much regard should be paid to this as a feature of taxonomic significance, since the condition may depend upon maturity and drying.

Nomenclature

The type specimen upon which the name S. densiflora is based was collected by Du Perry from the vicinity of Concepción, Chile. A fragment, deposited in the U.S. National Herbarium, was examined. Although the type specimen has no basal parts according to Hitchcock's note on the herbarium sheet, spikelet morphology clearly relates the type and the fragment to the original description by Brongniart (1829).

Chauvinia chilensis, described by Steudel (1855) is also based upon a plant collected from the vicinity of Concepción. It is difficult to see why Steudel chose to erect the genus Chauvinia when the type specimen of S. densiflora was accessible to him. From his published descriptions of both nomenclatural elements, the only apparent difference is the size of the plants, Chauvinia being slightly the smaller. Comparison of fragments of the C. chilensis type specimen with those of S. densiflora shows that the two undoubtedly belong to the same taxon.

Descriptions of the type specimen of S. montevidensis by Arechvaleta (1896) show it to be unquestionably S. densiflora. In addition to the description, there is presented a full-page, carefully executed drawing showing that the plant is strikingly similar to S. densiflora. Examination of a number of specimens referred to S. montevidensis shows no clear-cut morphological criterion by which a separation of S. montevidensis from S. densiflora may be effected. Parodi (1919) suggested that S. montevidensis might possibly be nothing but a variety of Brongniart's S. densiflora. Critical study of these specimens reveals nothing that would enable one to separate these plants in such a way that even varietal rank would be justified.

Spegazzini (1897) described a plant from the Argentine territory of Santa Cruz, giving it the name S. patagonica. The description of the plant clearly identifies it as S. densiflora although Parodi (1919) in his treatment of the Chlorideae of Argentina recognizes it as discrete from its several congeners occurring in Argentina.

Parodi's criteria for separating the three species which have, in this paper, been reduced to S. densiflora Brong. are presented in tabular form (Table 14). They are based solely on features of the inflorescence.

Table 14. Criteria for Speciation in Certain Spartinas in Argentina (Parodi, 1919)

Character	<u>S. densiflora</u>	<u>S. patagonica</u>	<u>S. montevidensis</u>
Panicle length	15--20 cm	6--15 cm	15--40 cm
Spike length	3--4 cm	2--4 cm	3--8 cm
Spike number	--	3--6	6--14
Spikelet length	12--13 mm	10--12 mm	10--12 mm
Relative lengths of glumes	1st longer than one-half the 2nd	1st shorter than one-half the 2nd	1st shorter than one-half the 2nd

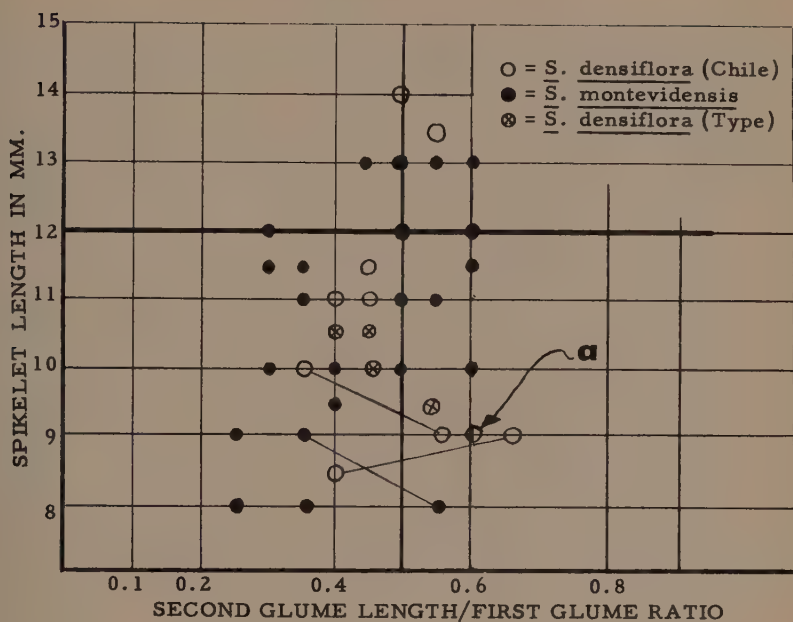
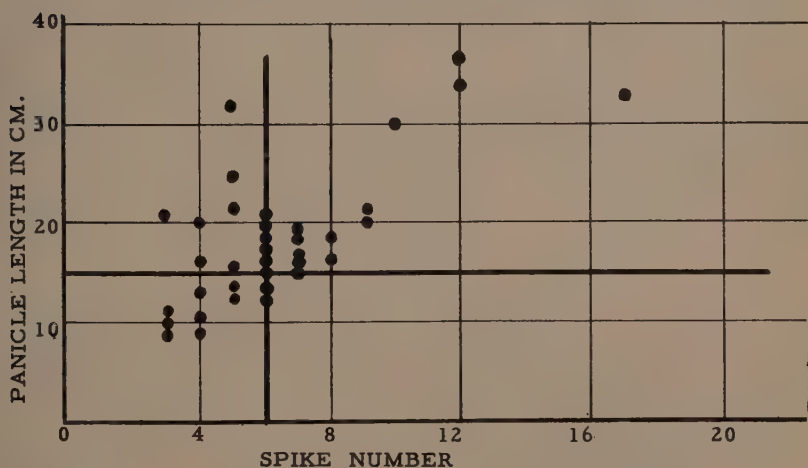


Fig. 15. Comparison of specimens of *Spartina densiflora* by ratio of second to first glume and spikelet length. ("a" represents a spikelet from the type specimen of *Chauvinia chilensis*.)



It is apparent that S. densiflora should be separable from the other two species on the basis of spikelet length and the ratio of glume lengths. Accordingly, a scatter diagram (Fig. 15) was constructed to learn whether these differences were actually valid and useful. By plotting the numerical value of the ratio of first glume to second glume against the length of the spikelet, it should be possible to show that all the plants examined fall into the first and third quadrants of the diagram, were the criteria valid. However, quite a different picture results. Not only is there a failure of the values to fall into place, but also, in certain situations where more than one spikelet per plant was measured, the scores for two spikelets from the same plant fell into different quadrants. These spikelets are connected by lines on the diagram. In general, an average spikelet was considered to be one from near the middle of a near-median spike. A number of spikelets from the type specimen also were introduced into the diagram. These are marked by an X in the open circles, which show the Chilean specimens. The black dots represent plants referred to S. montevidensis. From the evidence presented, the "species" are not separable from each other on the bases suggested. Further evidence supporting the view that these three "species" are not distinct is presented in Fig. 16, a scatter diagram based upon features which Parodi used to separate S. montevidensis from S. patagonica. By plotting the number of spikes per panicle against the length of the panicle, four quadrants are erected which should have specimens falling into only the first and third, were the criteria valid. Chilean specimens are excluded from this diagram; only specimens previously referred to S. montevidensis were used. No specimens were available which had been referred by others to S. patagonica. The scattering of dots representing specimens throughout all the quadrants but one, and the overlap which appears to exist, suggests that there is little basis for regarding S. patagonica and S. montevidensis as anything but synonyms of S. densiflora.

Saint-Yves (1932) described two subvarieties of S. densiflora var. typica. His variety typica is synonymous with S. densiflora, and, as noted in the discussion of synonymy of S. spartinae, is held by him to be discrete from his variety junciformis which is synonymous with S. spartinae. The two subvarieties of S. densiflora var. typica are separable, according to Saint-Yves, on the basis of the number of spikes and the length of the panicle. His criteria for separating the two taxa are presented in Table 15.

Table 15. Subvarietal criteria within S. densiflora var. typica. Saint-Yves (1932)

	Panicle length	Spike number
<u>S. densiflora</u> var. <u>typica</u> subv. <u>Brongniartii</u>	12--15 cm	12--15
<u>S. densiflora</u> var. <u>typica</u> subv. <u>pauper</u>	10 cm	4--8

Examination of twelve specimens previously identified as S. densiflora shows that only four answer the criteria proposed by Saint-Yves (1932). The remainder have panicles in excess of 10 cm in length and fewer than twelve spikes. It is at once apparent that these subvarietal taxa are ill-defined and should accordingly be treated as synonyms of S. densiflora.

Spartina montevidensis, shown in this paper to be a synonym of S. densiflora, has been considered by Saint-Yves (1932) as S. juncea var. montevidensis and var. laxiflora. His reasons for inclusion of these South American taxa within S. patens (S. juncea) are not at all clear. The vegetative appearance of S. montevidensis is altogether different than that of S. patens in that extensive rhizomes, characteristic of S. patens, are never found in S. densiflora. In addition, spikelets are usually one-nerved in S. densiflora and three-nerved in S. patens.

The varieties laxiflora and montevidensis can hardly be considered discrete taxa. S. juncea var. laxiflora is based upon a specimen collected and identified by P. Dusen as Spartina alterniflora Lois. A specimen from the same collection (P. Dusen 14264, MO) was examined. Criteria set up by Saint-Yves for the separation of the two varieties were the lax or gracile appearance of the spikes and the degree of remoteness of the spikelets from one another. Visual inspection of all the plants referred to S. densiflora by this author revealed that the lax attitude of the spikes was an extremely variable characteristic occurring occasionally among the specimens examined. Examination of Fig. 7, p. 506, will show that the degree of remoteness of the spikelets when reduced to a numerical basis (ratio of spike length to spike number) is disposed as a unimodal feature across the range of the population sample. The specimen referred to S. densiflora var. laxiflora occurs near the upper end of the population sample.

Spartina juncea var. montevidensis is clearly synonymous with S. densiflora for the reasons discussed in connection with the transfer of S. montevidensis to S. densiflora. This variety is held to be distinct from var. laxiflora because of the closely imbricate spikelets and the appressed spikes. As pointed out above, the occasional occurrence of a rather loosely appressed spike is rather sporadic throughout the population sampled, and does not appear to be correlated with other features such as the degree of remoteness of the spikelets. Attempts to delimit the varieties laxiflora and montevidensis by Saint-Yves are not warranted. All such specimens examined during the course of this study are distinctly referable to S. densiflora.

Distribution

S. densiflora is distributed across a considerable area of southern South America. Collection stations are plotted in Fig. 17. A pictorial diagram is attached to each station dot by means of a broken line. The pictorial diagrams show the relative median spike length and spike number of the specimens.

From the description of the species it is evident that considerable variation occurs in the number as well as the length of the spikes. Viewed geographically, a curious fact becomes apparent. Plants with fewer and longer spikes are more or less characteristic of the northern portion of the east coast range. Diminution in spike length and increase in number

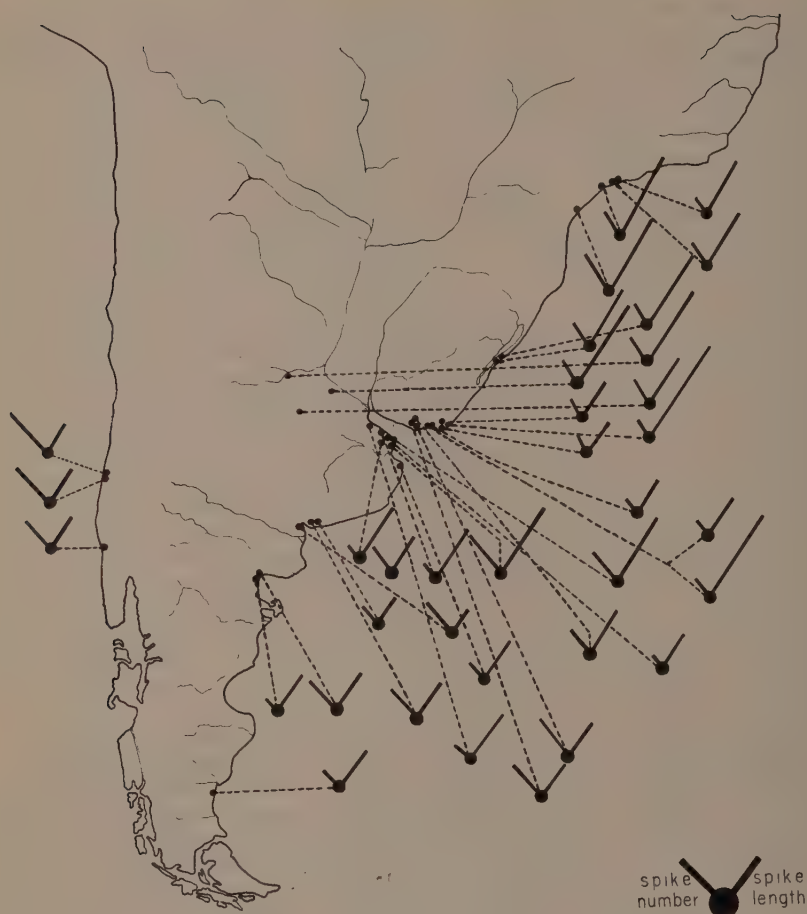


Fig. 17. Distribution of Spartina densiflora in South America.
(For discussion of pictorial diagrams, see text.)

are evident as one proceeds southward and across to the coastal areas of Chile. Very often diminution in spike length is accompanied by a perceptible increase in width. These features are not at all consistent but represent only the general trends within the population. In all other characters, no such distribution phenomena were observed. The primary habitat of the species seems to be the drier parts of the coastal marshes. However, several collections have been made at inland stations. Collection records show that the plant flowers from December to May.

14. SPARTINA GRACILIS

Spartina gracilis Trin. Mem. Acad. St. Petrsbg. Sci. Nat. IV.
6:110. (1840) (Type in LE from North America)

Description (Plate V, a-c)

Culms to 1 meter tall, erect, terete, indurate, glabrous, solitary, from widely spreading rhizomes, scales closely imbricate; sheaths smooth to striate, glabrous, the margin of the orifice usually glabrous, rarely short pilose; ligule a ring of hairs 0.5--1 mm long; blades elongate, flat, becoming involute, scabrous on the adaxial surface, glabrous on the abaxial, margins scabrous, tip acuminate; panicle 8--25 cm long; panicle axis 3--4-angled, flattened, scabrous on the angles; spikes 4--12, 1.5--8 cm long, 2--6 mm wide, alternate, usually appressed, not closely imbricate, sessile; rachis of the spikes scabrous on the angles; spikelets 10--30, 6--11 mm long, alternate, appressed, rarely spreading, moderately imbricate; first glume narrow, 3--7 mm long, glabrous to sparingly pilose, keel glabrous to pilose, margins sparingly hispidulous, tip acute or often mucronate; second glume 6--10 mm long, glabrous to sparingly pilose, keel pilose, margins glabrous to sparingly hispidulous, tip awnless, acute or often mucronate; lemma glabrous to sparingly pilose, keel pilose at least on the upper one-half, margins sparingly hispidulous, tip obtuse, rounded or obscurely lobed; palea thin and papery, glabrous, sparingly hispid above, tip obtuse to slightly rounded; anthers 2.5--5 mm long; somatic chromosome number 42.

Observations

This species is one of the more uniform ones within the genus despite the fact that it is distributed from south-central Mexico to the McKenzie Territory in Canada and occurs at altitudes from about sea level in the Great Basin to 7200 feet. Comparison of specimens from widely separated geographic localities shows little variability other than in spike number and culm height. None of the variations could be correlated with geographic distribution. Similarly, comparison of specimens collected at varying altitudes revealed no marked differences. Some variability exists in spikelet shape, grading from ovate to lanceolate (Plate V, a-c), but spikelet shape could not be correlated with other morphological features or geographical or ecological distribution. The ovate spikelets are shorter, in general, than are the lanceolate. The second glumes of the more ovate spikelets may be acute to obtuse at the tip while the more lanceolate are usually acuminate. Of particular importance taxonomically is the presence of two (rarely one) prominent lateral nerves on the second glume. Both are disposed on one side of the densely pilose keel.

A single chromosome count has been reported. Church (1940) found the species to have the hexaploid number of 42.

Nomenclature

A photograph of the type specimen, received from the Komarov Botanical Institute of Leningrad, clearly establishes that Spartina gracilis is properly typified. The photograph is deposited in the herbarium of the Iowa State College.

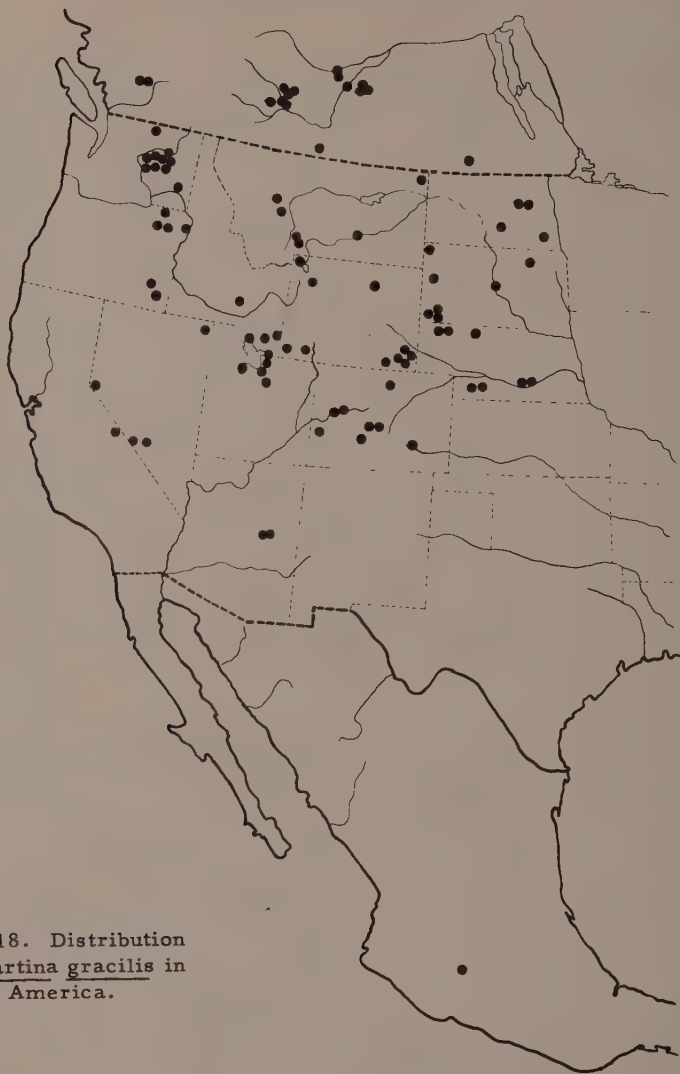


Fig. 18. Distribution of *Spartina gracilis* in North America.

Distribution

Distribution of this species is shown in Fig. 18. Two specimens collected from the McKenzie Territory north of Saskatchewan have been omitted from the map. These two specimens and the one from Mexico are similar in every respect to the majority from the central portion of the range.

The plants grow in a somewhat specialized habitat, usually at the margins of alkali lakes and along stream margins and river bottoms. The plants flower from June to October.

15. SPARTINA PATENS

- Spartina patens (Ait.) Muhl. Descr. Gram. 55. (1817)
Dactylis patens Ait. Hort. Kew. 1:104. (1789) (Type grown in
 England from seed from New York; isotype in K)
Trachynotia juncea Michx. Fl. Bor. Amer. 1:64. (1803)
 (Type in P from Carolina)
Limnethis juncea (Michx.) Rich. in Pers. Syn. Pl. 1:72. (1807)
Spartina pumila Roth. Cat. Bot. 3:10. (1806) (Type from New York)
Spartina juncea (Michx.) Willd. Enum. 81. (1809)
Limnethis juncea (Michx.) Rich. var. monogyna M.A. Curtis
 Bost. Jour. Nat. Hist. 1:136. (1835) (Type from Cape Fear,
 North Carolina; isotype in GH)
Spartina americana Roth in Trin. Mem. Acad. St. Petersburg.
 Sc. Nat. VI. 6:109. (1840) pro syn.
Spartina duriaei Parl. Fl. Ital. 1:230 (1848)
Spartina versicolor Fabre, Ann. Sci. Nat. Paris III. 13:123. (1849)
 (Type from Southern France)
Spartina patens (Ait.) Muhl. var. juncea (Michx.) Hitchc.
 Rhod. 8:210. (1906) (Based on Trachynotia juncea Michx.)
Spartina juncea (Michx.) Willd. var. juncea St. Y. Cand. 5:82. (1932)
Spartina juncea (Michx.) Willd. var. juncea (Michx.) St. Y. subv.
europaea St. Y. Cand. 5:82. (1932) (Based on S. duriaei Parl.)
Spartina juncea (Michx.) Willd. var. juncea (Michx.) St. Y. subv.
americana St. Y. Cand. 5:84. (1932) (Based on S. juncea Willd.)
Spartina juncea Willd. var. patens (Ait.) St. Y. Cand. 5:86. (1932)
 (Based on Dactylis patens Ait.)
Spartina patens (Ait.) Muhl. var. monogyna (M.A. Curtis) Fern.
 Rhod. 49:114. (1947) (Based on Limnethis juncea var. monogyna
 M.A. Curtis)

Description (Plate V, d-k)

Culms to 1.5 meters tall, slender, erect, terete, glabrous, usually solitary or in small clumps from widely spreading, slender, wiry rhizomes, scales not closely imbricate; sheaths smooth to striate, glabrous, margin of the orifice glabrous, rarely short pilose; ligule a ring of hairs 0.5 mm, rarely to 1 mm long; blades short, involute, rarely flat, narrow, scabrous on the adaxial surface, glabrous on the abaxial, margins scabrous, tip acuminate; panicle 3--15 cm long; panicle axis 3--4-angled, scabrous on the angles; spikes 2--15, 1--7 cm long, alternate, appressed or spreading, often distant, commonly sessile, occasionally pedunculate; rachis of the spikes scabrous on the angles; spikelets 10--30, 7--12 mm long, alternate, tightly appressed, closely imbricate; first glume linear, 3--8 mm long, glabrous to sparingly hispidulous, keel hispid, margins sparingly hispidulous, tip acuminate; second glume 7--12 mm long, glabrous to sparingly hispidulous, keel hispid, lateral nerves commonly hispid, disposed on one side of the keel, margins glabrous to hispidulous, tip acuminate; lemma glabrous to sparingly hispidulous, keel hispid at the tip, margins glabrous to very sparingly hispidulous, tip obtuse, rounded or often obscurely lobed; palea thin and papery, glabrous, tip obtuse, rounded, exceeding the lemma; anthers 3--5 mm long; chromosome numbers 28, 35, 42, 56.

Observations

Spartina patens, a highly variable species, has been found to possess several levels of polyploidy (Church, 1940). Specimens from Massachusetts, Virginia, and Florida were shown to possess the tetraploid, hexaploid, and octoploid number of chromosomes respectively. According to Church, comparison of morphological differences among the specimens showed that the one from Florida was considerably more robust than the others, and the one from Massachusetts had spikelets much less closely imbricate on the spike. The Massachusetts specimen corresponded to S. patens while the Virginia and Florida specimens answered to the description of S. patens var. junceae.

The several specimens whose chromosome numbers were determined as a part of the present investigation do not answer satisfactorily to the descriptions of the two taxa nor do they agree with the geographic distribution of polyploid levels suggested by Church.

Table 16. Chromosome numbers in S. patens

Collection	Location	Chromosome number
Mobberley 115	Fort Walton, Florida	2n = 35
Mobberley 121	Destin, Florida	2n = 42
Mobberley 193	Cape Hatteras, North Carolina	2n = 28

The pentaploid does not seem to be morphologically different from either of the others. It is presumed that it originated from a cross involving tetraploid and hexaploid parents.

Several authors (Fernald 1947, 1950; Hitchcock 1906, 1935; and Merrill 1902) have held the view that this species is distinctly separable into two well-defined taxa. Chase (1950) recognized the two taxa but stated that intermediate specimens occurred frequently. Fernald (1947), having rectified the earlier nomenclatural errors, recognized S. patens and S. patens var. monogyna. The two are supposedly distinguished by a number of morphological criteria, geographic distribution which overlaps somewhat and differing ecological preferences. That occurrence in differing habitats is not correlated with morphological differences will be shown in the section of this paper dealing with distribution.

The several morphological characters by which the two taxa are supposedly separated are tabulated in Table 17.

In some cases, specimens could be assigned to one of the two taxa on the basis of the criteria provided. However, the degree of overlap in most of the criteria precluded the accurate identification of many.

In an effort to learn whether the population was actually divisible into the two taxa suggested, a method of numerical analysis was invoked. The range of variability was determined for each of the above criteria amenable to numerical analysis. These data were accumulated from the examination of 235 specimens. Simple measuring or counting was sufficient for most of the criteria. Blade angle was measured with a protractor in degrees from the longitudinal axis of the culm. Spikelet imbrication was

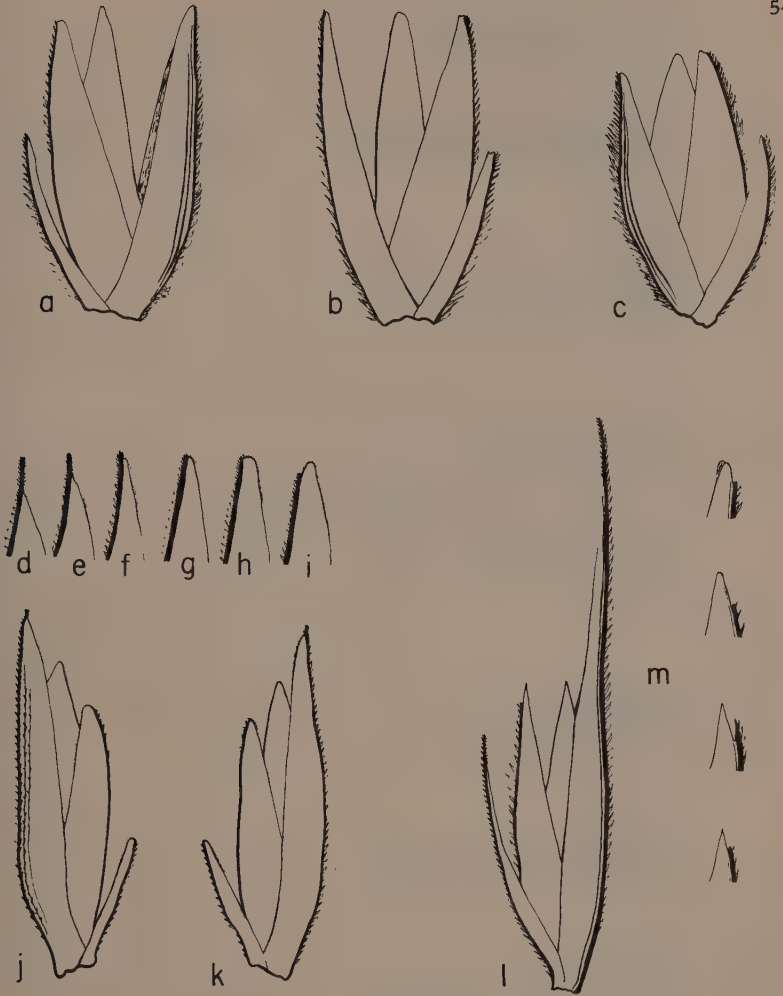


PLATE V

- a-c. Spartina gracilis. a. spikelet showing lateral nerves 3.75X,
b. same spikelet, opposite view 3.75X,
c. spikelet, 3.75X.
- d-k. Spartina patens. d-i. variation in second glume tips 7.5X,
j. spikelet showing lateral nerves 3.75X,
k. same spikelet, opposite view 3.75X.
- l-m. Spartina pectinata. l. spikelet 3.75X,
m. variation in lemma tips 7.5X.

Table 17. Criteria of Merrill (1902) and Fernald (1950) for separating S. patens from var. monogyna

Character	<u>S. patens</u>	<u>S. patens</u> var. <u>monogyna</u>
Rhizome diameter (mm)	1--3	2--6
Culm diameter (mm)	1--2.5	1--6
Culm height (dm)	1.5--8	2--15
Leaf number	2--5	5--9
Blade length (dm)	0.5--2	1--5
Blade angle	spreading	appressed
Spike number	1--4	2--9
Spikelet length (mm)	9--13	7--10
Spikelet imbrication	loose	tight
Second glume tip	acuminate	acute to blunt

reduced to numerical terms simply by dividing spikelet number into spike length. The second glume tip was found to vary considerably, grading from acuminate (nearly aristate) to blunt. Its range of variability was arbitrarily broken up into six specific types which served as models by which the tips could be numerically scored (Plate V, d-i). Once determined, the range of variability of each character was divided into six classes as shown in Table 18.

Table 18. Class values for morphological criteria of S. patens

Criteria	0	1	2	3	4	5
Rhizome diameter (mm)	0--1	1.1--2	2.1--3	3.1--4	4.1--5	5.1--6
Culm diameter (mm)	0--1	1.1--2	2.1--3	3.1--4	4.1--5	5.1--6
Culm height (cm)	14--36	37--59	60--82	83--105	106--128	129--150
Leaf number	3--4	5	6	7	8	9--12
Blade length (cm)	3--8	9--14	15--20	21--26	27--32	33--41
Leaf blade angle	77--65°	64--52°	51--39°	38--26°	25--13°	12--0°
Spike number	1--2	3	4	5--6	7--9	10--13
Spikelet length (mm)	11--14	10--10.9	9--9.9	8--8.9	7--7.9	5--6.9
Spikelet imbrication value	2.7--3.4	2.3--2.6	1.9--2.2	1.5--1.8	1.1--1.4	0.7--1.0
Second glume tip	Pl. V, d	Pl. V, e	Pl. V, f	Pl. V, g	Pl. V, h	Pl. V, i

The values are arranged so that classes zero and five characterize typical S. patens and var. monogyna, respectively. Distribution by classes for each of the ten characters is shown in Fig. 19. In all the characters the distribution is unimodal.

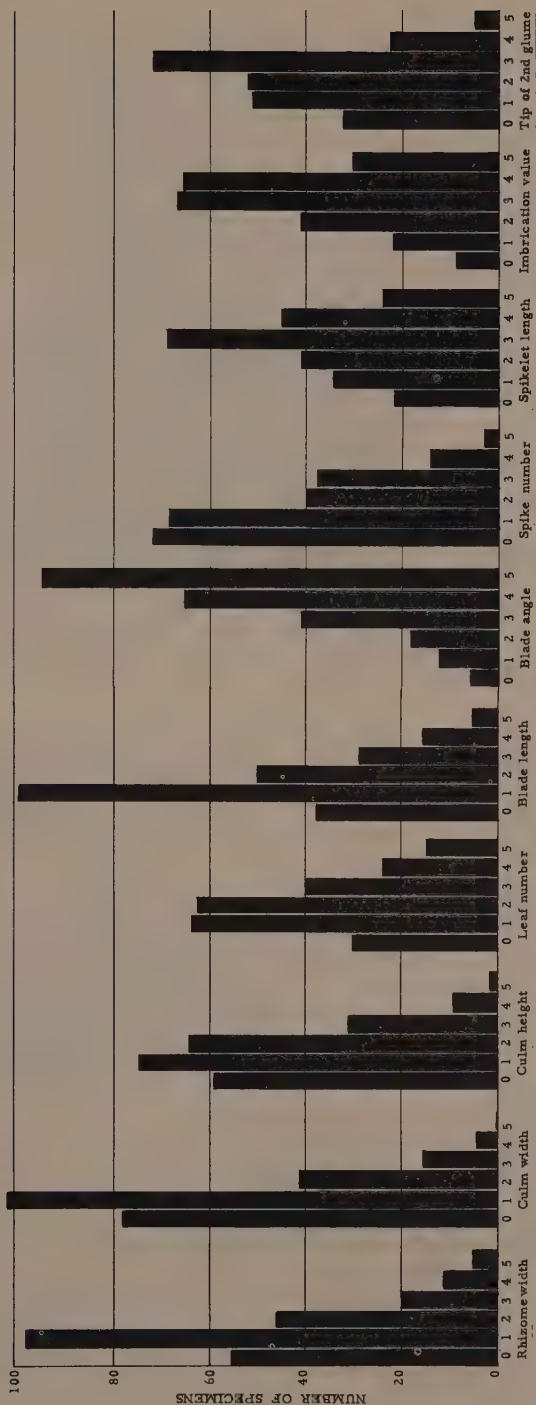


Fig. 19. Distribution by classes (0--5) for morphological features of *Spartina patens*.

Each of the specimens was scored according to the values shown in the above table. The scores for all characters of each specimen were summed and the total introduced into a histogram (Fig. 21). Specimens scored on these bases do not fall into two separable groups.

Fernald (1947) suggested that plants from the region north of New Hampshire were the typical S. patens and that plants south of Virginia were variety monogyna. The distribution of the two taxa was thought to be coincident from New Hampshire to Virginia. None of the criteria utilized in the present study exhibited variability that could be correlated with geographic distribution beyond the fact that smaller plants appear to be more typical of the northern part of the range. However, when the total scores for the specimens were introduced into histograms in which the three geographic sections of the range were treated independently, some differences in the population became evident. Fig. 21, A, B, and C show that higher total scores were found among specimens from Texas to North Carolina while the lower total scores were found among specimens from Maine northward. However, the large degree of overlap provided by total scores of specimens from the intermediate region and the slight overlap between the southern and northern extremes seem to preclude taxonomic separation.

Seven European specimens were scored according to the methods described above. The total scores for these specimens ranged from twenty-eight to thirty-three. These specimens are not separable from the North American material on the basis of the above criteria.

Nomenclature

Spartina patens was first described by Aiton (1789) as Dactylis patens. The type specimen was not seen. However, Aiton's detailed description offers evidence supporting the accuracy of typification. Muhlenberg (1817) properly effected the transfer of this species to Spartina.

Curtis (1835) recognized two taxa within the species. Limnietis juncea var. monogyna was held to be distinct from L. juncea which he evidently took to be a less robust form. Hitchcock (1906) also recognized two taxa and provided the name Spartina patens var. juncea for the more robust form of the species. Realizing that var. monogyna should have received priority, Fernald (1947) provided the combination S. patens var. monogyna. In the light of the evidence presented above, effective separation of the two taxa is not possible. It therefore seems desirable to regard the entire population as a single species that possesses several indistinguishable levels of polyploidy.

In the recent monographic treatment of the genus Spartina by Saint-Yves (1932), this species was subdivided into varietal and subvarietal categories under the name Spartina juncea. The segment of Saint-Yves' S. juncea recognized as S. patens in this paper was subdivided into var. patens and var. juncea. The latter, in turn, was subdivided into subv. americana and subv. europaea. Variety juncea was held to be separable from variety patens by virtue of a difference in spike number (three or more in juncea, three or less in patens) and appressed or spreading blades. The present investigation shows that many plants with less than three spikes have leaf blades tightly appressed to the culm. The subvarieties of var. juncea were claimed by Saint-Yves to be separable on

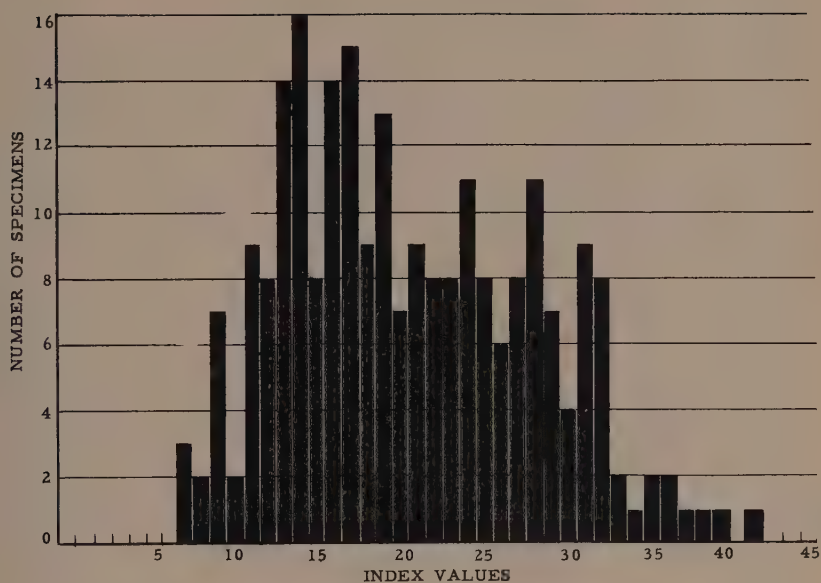


Fig. 20. Distribution of Index Total Scores.

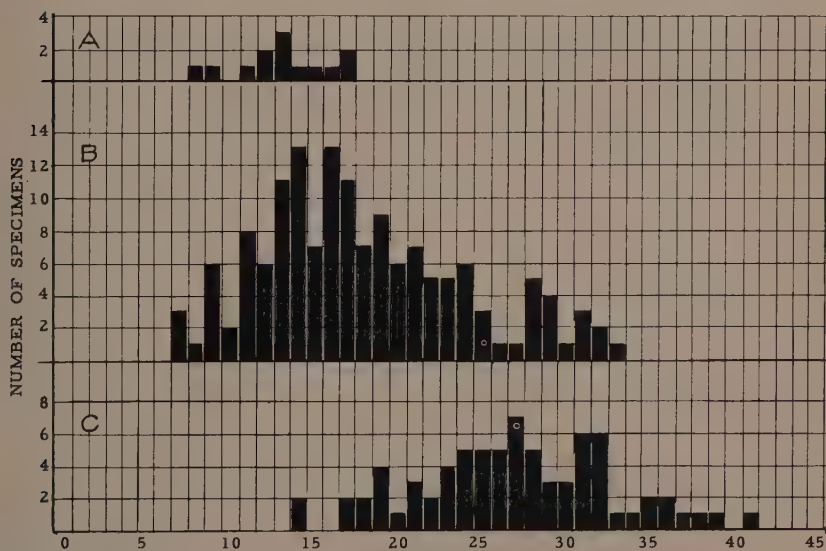


Fig. 21. Distribution of Index Total Scores for specimens of differing geographic localities.

A. Maine to the Maritime Provinces.

B. Virginia to New Hampshire

C. Texas to New Hampshire.



Fig. 22. Distribution of *Spartina patens* in North America and the Caribbean.

the basis of spikelet length which was found to be 6 mm or less in var. europaea and 7--9 mm in var. americana. The present study shows that spikelets of American material may be less than 6 mm in length. Five of the seven European specimens examined have spikelets in excess of 6.5 mm. In view of the obstacles to accurate identification of plants by means of the criteria provided by Saint-Yves, and in the light of the results of the investigation presented in the preceding section, these varieties and subvarietal names are regarded as synonyms of S. patens.

Distribution

Distribution of Spartina patens in North America is shown in Fig. 21. The presence of this species in Europe is probably a result of introduction of rhizomes or seeds from North America by means of ship ballast. It is known from southern France, Corsica, and Italy. A distribution map is not provided for the species in Europe because geographic data on all but one of the sheets were too meager for accurate location of the sites.

While principally a denizen of the coastal salt and brackish water habitats in North America, the species has been found in inland regions in New York and Michigan. These plants from the inland stations are doubtless relics of a maritima flora once common in the Great Lakes region.

Typically, the species is one of the dominant components of the coastal salt marshes. It is frequently found in unbroken stands from the dry, sandy beach above the intertidal zone well up into the drier portions of the marshes. No consistent visible differences could be found in plants that had been collected from the beaches and those from the salt marshes. Occasionally, plants from the beach zone were found to be somewhat glaucous. However, the presence of the bloom could not be correlated with the other morphological characters by which Fernald (1947) sought to set apart S. patens of the salt marshes from S. patens var. monogyna of the strand.

While the time of blooming is somewhat erratic, flowering usually occurs from June to September.

16. SPARTINA PECTINATA

Spartina pectinata Link, Jahrb. Gewachsk. I. 3:92. (1820)

(Type from North America in B)

Trachynotia cynosuroides Michx. Fl. Bor. Amer. 1:64. (1803)

(Not D. cynosuroides of Linnaeus) (Type in P from Illinois)

Limnethis cynosuroides Rich. in Pers. Syn. Pl. 1:72. (1807)

(Based on T. cynosuroides Michx.)

Spartina pectinata Bosc. Ind. Kew. 2:952. (1895) (nomen; error for S. pectinata Link)

Spartina michauxiana Hitchc. Contr. U.S. Nat. Herb. 12:153. (1908)

(Based on T. cynosuroides Michx.)

Spartina michauxiana var. suttiei Farw. Rep. Mich. Acad. 21:352.

(1920) (Type in BLH from Orchard Lake, Michigan)

Spartina michauxiana var. tenuoir Farw. Rep. Mich. Acad. 21:352.

(1920) (Type in BLH from River Rouge, Michigan)

Spartina cynosuroides Willd. var. michauxiana St. Y. Cand. 5:58.
(1932) (Based on T. cynosuroides Michx.)

Spartina cynosuroides Willd. var. michauxiana f. major St. Y.

Cand. 5:62. (1932) (Based on S. pectinata Link)

Spartina cynosuroides X gracilis St. Y. Cand. 5:66. (1932) Type not designated; based upon four specimens from Cusick 221, Oregon, 1890. Ballard's Landing, Snake River (error for 2221 in 1899)

Spartina pectinata Link var. suttiei (Farw.) Fern. Rhod. 35:260.

(1933) (Based on S. michauxiana var. suttiei Farw.)

Description (Plate V, 1-m)

Culms to 2.5 meters tall, erect, terete, indurate, usually solitary from widely spreading, elongate, indurate, brownish rhizomes, scales closely imbricate; sheaths smooth to slightly striate, often keeled above, glabrous, margins glabrous to scabrous, margin of the orifice often pilose; ligule a ring of hairs 1--3 mm long; blades flat, becoming involute in drying, glabrous on both surfaces, margin scabrous, tip acuminate; panicle 10--50 cm long; panicle axis 3-angled, glabrous to scabrous on the angles; spikes 5--50, 1.5--15 cm long, the lower usually slightly the longer, 2--15 mm wide, appressed to somewhat spreading, mostly pedunculate, occasionally sessile, alternately to irregularly arranged; rachis of the spikes glabrous to scabrous on the angles, not prolonged beyond the terminal spikelet; spikelets 10--80, 10--25 mm long, alternate, appressed, closely imbricate, rarely subremote; first glume narrow, 5--10 mm long, glabrous to sparingly hispidulous, keel hispid, tip awned; second glume 10--25 mm long, including the awn, glabrous to sparingly hispidulous, keel pectinate, tip produced into a scabrous awn up to 8 mm in length; lemma glabrous, keel glabrous below, pectinate above, margins hispidulous, tip apiculate, deeply bilobed; palea thin and papery, bilobed and rounded at the tip, slightly exceeding the lemma; anthers 4--6 mm long; somatic chromosome numbers 42, 70, 84.

Observations

Spartina pectinata is a variable species that has been shown to possess at least three levels of polyploidy. Church (1940) reported a hexaploid from Massachusetts and a dodecaploid from Nebraska. In the present investigation, a hexaploid and a decaploid were found from two stations in Iowa.

Table 19. Somatic chromosome numbers for S. pectinata

Collection	Location	Chromosome number
Mobberley 7	Boone County, Iowa	2n = 42
Morrill, John B.	Monona County, Iowa	2n = 70

A comparison of the two specimens shows very little morphological difference. In fact, a comparison of over one hundred specimens from across the range of distribution of the species reveals nothing that would in any way suggest morphological differences that might be correlated

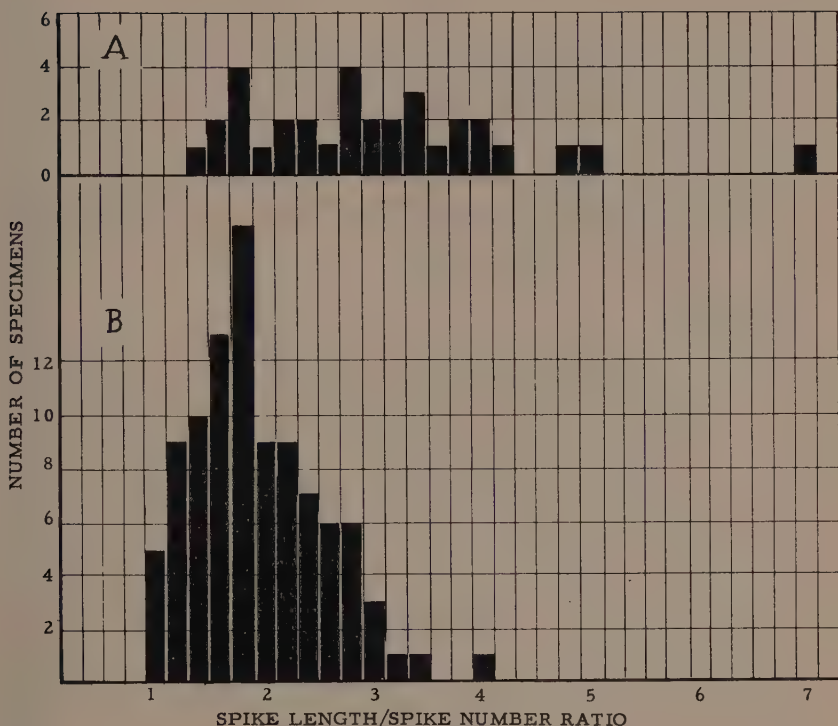


Fig. 23. Comparison of specimens of *Spartina pectinata* from Eastern and midwestern North America by Spikelet imbrication value.
A. Eastern North America. B. Midwestern North America.

with varying levels of polyploidy. A number of specimens were found that had spikelets less closely imbricate along the spike rachis. These specimens seemed to be much more common in material from the maritime provinces of Canada and New England. The imbrication values of a number of specimens were plotted in the accompanying histogram (Fig. 23). The values for specimens from New England and eastern Canada show a much greater variability than do those of the remainder of the range, but nothing in the data presented would enable one to effect a satisfactory taxonomic separation.

The vegetative features of the species offer very little that is taxonomically useful. However, several excellent characters are found in the inflorescence. The extremely long awn, the sharply toothed lemma and the nature of the pubescence are all found useful for separating this species from others of the genus.

Two minor forms of *S. pectinata* have been described. Irving (1905) published a description for *S. cynosuroides* (Willd.) var. *aureo-marginata*. The plant agrees in all particulars with *S. pectinata* except that the margins of the leaves are striped with a golden band running the length of the

the leaf. It appears that this unusual color form arose as a mutation from an otherwise typical clone. In keeping with the most recent interpretations of taxonomic hierarchy, the variety thus described is reduced to a form and becomes Spartina pectinata Link forma aureo-marginata (Ilv.) comb. nov.

A second such form was described by Marie-Victorin (1944). It differs from the preceding in that the leaves are marked with white along the margins and the median nerve. The type specimen is deposited in the herbarium of Montreal University. Nomenclaturally, it is Spartina pectinata Link forma variegata M.-Vict.

Nomenclature

The case for ascription of the name S. pectinata to the present taxon is carefully stated by Fernald (1933). The first description is that of Link (1820) from a plant sent to him from North America by Bosc. In the original paper, Link cites "Nord-Amerika" as the type locality.

The authors of Index Kewensis (Hooker and Jackson, 1895) have erroneously ascribed the name S. pectinata to Bosc. Fernald (1933) showed that Link's use of the word "von" (S. pectinata, von Bosc) was not a citation of a description by Bosc but was intended only to show that his (Link's) description was based upon a specimen supplied by Bosc. Fernald states that a fragment of the type in the Gray Herbarium, given to Asa Gray many years ago, agrees in all particulars with this present transcontinental species. Thus, there can be little doubt of the validity of the name and the correctness of the author citation.

Considerable confusion has arisen concerning the name Spartina cynosuroides Willd. Correctly cited by Willdenow (1809) in synonymy of the true S. cynosuroides is Dactylis cynosuroides of Linnaeus. Incorrectly cited, however, is Trachynotia cynosuroides of Michaux which has been shown to be synonymous with S. pectinata Link. Michaux (1803) published a description of Trachynotia cynosuroides erroneously basing the specific epithet upon D. cynosuroides of Linnaeus. The plant he described, however, was collected in Illinois and belongs to the inland species; in reality, the true S. pectinata. At the same time, Michaux also described the coarse halophytic species, now known as S. cynosuroides (L.) Roth, as Trachynotia polystachya.

The citation of Michaux's T. cynosuroides by Willdenow undoubtedly led later workers into erroneous usage of the combination S. cynosuroides Willd. or S. cynosuroides (L.) Willd. for S. pectinata. Among those presumably misled by Willdenow and Michaux was Asa Gray (1848). The first edition of The Manual of Botany incorrectly referred the transcontinental inland species to S. cynosuroides Willd. The proper combination was used in a succeeding edition for the first time by Fernald (1950).

Hitchcock (1908) rejected the name S. pectinata on the grounds that the type was collected "probably in South Carolina, where (it) does not grow." In its stead he proposed the name Spartina michauxiana, basing it upon Trachynotia cynosuroides of Michaux. There is no indication that Bosc's collecting expedition was limited to South Carolina or that the type specimen was not obtained by Bosc from some other North American source. The plant is known from Virginia and North Carolina, although

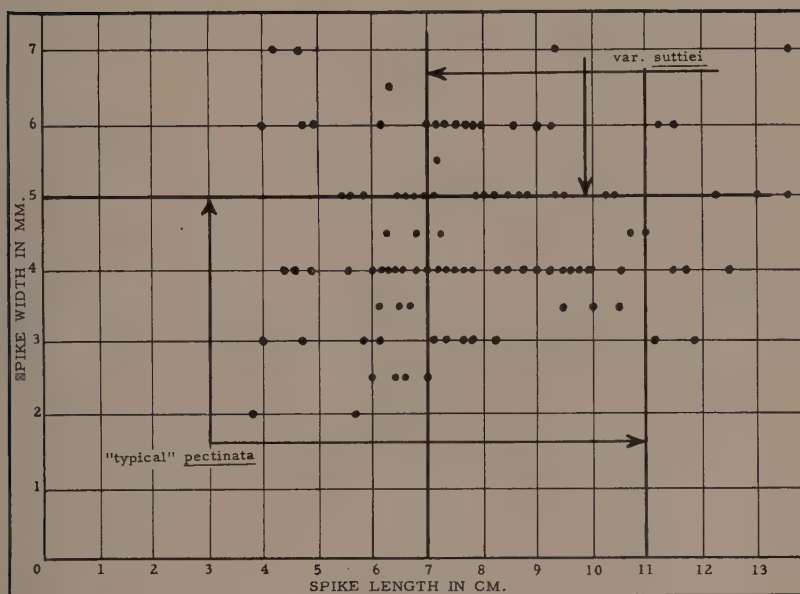


Fig. 24. Comparison of specimens of *Spartina pectinata* by spike length and spike width.

as Blomquist (1948) has shown, its incidence in North Carolina is restricted to two mountainous localities in the extreme western part of the state.

Several puzzling varieties of *S. pectinata* have been described. Two of those were published by Farwell (1920) as *S. michauxiana* var. *suttiei* and var. *tenuior*. Holotypes of both of these varieties from the herbarium of the Cranbrook Institute of Science were examined. Varietal rank was assigned the specimens by Farwell on the basis of panicle, spike, and peduncle lengths. Var. *suttiei* was delimited by panicles 1.5 feet long, spikes 5--7 inches long, and peduncles 1.5--2 inches long. Var. *tenuior* is delimited by peduncles three-eighths of an inch long or less.

Farwell's variety *suttiei* does not appear to be separable on the bases offered. Transposed to metric values, the minimum limits for panicle and spike length are 45.7 cm and 12.7 cm, respectively. Of the several hundred panicles of *S. pectinata* examined during the course of this investigation, only three were found in excess of 45 cm in length, and in each case the median spike length was found to be less than 10 cm. Several specimens were found in which the median spike length exceeded 12.7 cm, but in each case the panicle length did not exceed 43 cm. The type specimen has a panicle 41 cm long with the median spike 14 cm long. Peduncles in excess of 3.8 cm are supposed to characterize variety *suttiei*, but of all the specimens examined, only one, a highly aberrant

specimen (Aiton, G.B. 4795, Minneapolis, Minn. WISC), has a peduncle in excess of 3 cm. This specimen had only one elongate spike, the others having aborted. Further, in Farwell's holotype peduncle lengths do not exceed 2.4 cm.

Similarly, Farwell's variety tenuior does not appear to be distinct. In all but one of the features by which Farwell sought to distinguish the variety, it is overlapped by the typical variety. He did, however, set apart variety tenuior on the basis of peduncles which were said not to exceed 0.9 cm. Spikes of the variety were not supposed to exceed 7.5 cm in length. Examination of over one hundred specimens shows that many have spikes in excess of 7.6 cm and peduncles much less than 0.9 cm.

Fernald (1933) took up the variety suttiei and recombined it as S. pectinata Link var. suttiei but ignored Farwell's var. tenuior. This variety is supposedly separable from the typical variety because of its long, narrow spikes. Ranges given for length and width of spikes are: "typical" variety 2--11 cm long, 5--8 mm wide, including the salient awns, var. suttiei 7--15 cm long, 3--5 mm wide. In the variety suttiei the awns are supposed to be appressed and the spikes somewhat more peduncled. No values for peduncle lengths are given. Hitchcock (1935), Swallen (1939), and Chase (1950) all cite this variety as a synonym but state no reasons. Fassett (1951) states that among Wisconsin specimens the spikes vary from 3.5--13 cm in length and from 2--10 mm in width, but that the short spikes are often narrow and the long ones often broad. From this, he concludes that there can be no division of the species into discrete varieties.

In an effort to clarify the picture within this species, spike length is plotted against spike width in a scatter diagram (Fig. 24). Were variety suttiei distinct from the "typical" variety, specimens should fall discretely into the marked sections of the diagram. It appears that instead of a clear-cut separation, dots are found scattered throughout the diagram. The foregoing evidence suggests that both Farwell and Fernald were dealing with selected extremes of a somewhat variable population rather than with discrete infraspecific taxa.

Saint-Yves (1932) described what he thought was an intermediate between S. cynosuroides Willd. and S. gracilis Trin. A type specimen was not designated from among the four specimens of the single Cusick collection 2221 at hand. His description clearly indicates that a somewhat less robust form of S. pectinata is involved. Specimens from this same collection (Cusick 2221, Ballard's Landing, Snake River, Oregon, 1899, F and UC) were examined. There is little doubt that the collection is S. pectinata for the specimens agree in all particulars with that species.

Spartina cynosuroides var. michauxiana f. major has been proposed by Saint-Yves (1932) as an intermediate form between his varieties michauxiana and polystachya. As stated above in the lists of synonyms, the variety polystachya is synonymous with Spartina cynosuroides (L.) Roth, while michauxiana is synonymous with S. pectinata. Saint-Yves seems to have satisfactorily separated his two varieties on the basis of their nearly complete geographic discontinuity. However, plants assigned by him to the intermediate forma major are largely from the range of true S. pectinata and are doubtfully separated from it on the basis of a greater number of spikes. The rather unsatisfactory statement that the two



Fig. 25. Distribution of *Spartina pectinata* in North America.

varieties grade imperceptibly into one another through the intermediate form concludes his argument. The forma *major* is therefore referred to *S. pectinata* as a synonym. That the two "varieties" are discrete species and rather easily separated will be shown in another section of this paper.

Distribution

Distribution of the widely-ranging *S. pectinata* is shown in Fig. 25. In addition to the stations shown on the map, the species also occurs in a restricted locality at the head of the Adriatic Sea in northern Italy and Trieste. It has presumably been introduced from North America.

The species, in eastern United States and Canada, is an inhabitant of marshes, sloughs, and flood plains. It seems to thrive equally well in both salt and fresh water habitats. Both *S. pectinata* and *S. patens* were found in salt marsh habitats in New England often growing intermingled with each other.

In the midwestern United States, *S. pectinata* has been found in wet

swampy places, but, contrary to most of the statements concerning habitat in extant keys and manuals, the primary habitat seems to be open, dry prairie or high ground along railroad rights-of-way and roadsides. Parcels of native prairie in parts of Iowa show dense growths of this species, often in large, pure stands. That it flourishes on disturbed ground is evident from its abundance on railroad embankments and highway slopes in the Midwest.

In North America, the species flowers from late June to September.

ANALYTICAL KEYS TO THE SPECIES

KEY TO SPECIES OF SPARTINA OCCURRING IN EUROPE AND AFRICA

1. Spikes closely imbricate, several often overlapping; leaf blades over 5 mm wide at the base; spikelets exceeding 10 mm . . . 2
1. Spikes not closely imbricate, not overlapping or only the upper part of one spike overlapping the next above; leaf blades less than 5 mm wide at the base; spikelets not exceeding 9 mm in length; western Mediterranean region. 15. Spartina patens
2. Second glume long-awned, strongly hispid on the keel; culms and rhizomes hard and firm in texture; plants of Trieste and the surrounding region at the head of the Adriatic Sea. 16. Spartina pectinata
2. Second glume not awned, pilose on the keel; culms and rhizomes soft and succulent in texture. 3
3. Spikelets exceeding 15 mm in length. 4
3. Spikelets not exceeding 15 mm in length 5
4. Panicles exceeding 7 mm in width; ligule 1.5--3 mm long. 9. Spartina townsendii
4. Panicles not exceeding 5 mm in width; ligule about 1 mm long; plants of southwestern France. 8. Spartina neyrautii
5. Leaf blades readily disarticulating, not exceeding 12 cm ; spikelets densely pilose. 7. Spartina maritima
5. Leaf blades not disarticulating, exceeding 17 cm in length; spikelets not usually densely pilose. 6
6. Rachis of a median spike not less than 1 mm in width; plants of southwestern France. 8. Spartina neyrautii
6. Rachis of a median spike not more than 1 mm wide; not plants of southwestern France if over 1 mm. 4. Spartina alterniflora

Discussion

The European species of Spartina are not too difficult to identify. The introduced S. patens is easily distinguished from the other European species on the basis of its short spikes and spikelets, densely imbricate spikelets and narrow leaves. S. pectinata, also introduced, may be distinguished from its congeners on the basis of its strongly hispid second glume keel, long awn, and restricted distribution at the head of the Adriatic Sea.

That the introduced S. alterniflora is discrete from the indigenous S. maritima was shown by Fernald (1916), although some earlier authors considered the two identical. Fernald cited the readily disarticulating leaf blades and the fact that the rachis is not prolonged beyond the terminal spikelets as features by which S. maritima differed from S. alterniflora. The disarticulating leaf blades of S. maritima appear to be a constant feature throughout the population, in about one-half of the eighty specimens of S. maritima examined by this investigator, the rachis of the spikes was actually prolonged beyond the terminal spikelet, often by as much as 1 cm.

Jovet (1941) showed that S. alterniflora was distinct from S. maritima on the basis of ligule length. He gives ligule length values as 0.5 mm for S. maritima and 1.0 mm for S. alterniflora. The present investigation revealed considerably more variation than indicated by Jovet. Ligule lengths were not found to exceed 1 mm in S. maritima, while in the European S. alterniflora they varied from 0.7--2 mm.

Length of leaf blade offers an unusually effective means of separation of the two species. Lengths of the blade of the second leaf from the summit in the European S. alterniflora were found to vary from 17 to 44 cm. Frequently, the second leaf blade in S. maritima was missing. In this event, a blade lower down on the culm was measured. In no case was the length of the blade found to exceed 12 cm.

The degree of pubescence on the glumes also serves to separate the two species. The glumes of S. maritima are densely pubescent, while even in the most pilose specimens of S. alterniflora the amount of pubescence is much less than that encountered in S. maritima (Plates II, a-d and III, c-d).

Pubescence on the keel of the lemma seems to merit some attention. European specimens of S. alterniflora were observed to have an essentially glabrous lemma keel while that of S. maritima is pilose at least on the upper two-thirds. The lemma keel of S. alterniflora may possess a few soft trichomes near the tip.

The nature of the second glume tip may also offer an effective taxonomic criterion. While quite variable in both species, there appears to be a rather definite separation of midvein from the lamina of the glume in S. maritima. Usually no such separation is present in S. alterniflora. Plates II, b, and III, c illustrate the variety of second glume tips encountered in both species.

S. alterniflora has not always been considered discrete from S. townsendii. Chevalier (1923) held S. townsendii to be identical with S. glabra var. pilosa of Merrill. Saint-Yves (1932) recognized none of the features by which the two species may be distinguished. Actual lengths of the leaf blades were not found to be a useful key character, but the fact that the

uppermost blade in S. townsendii only rarely exceeds the panicle may have some value. In almost all the European specimens of S. alterniflora the uppermost leaf blade exceeds the panicle.

Jovet (1941) found that ligule lengths of the two species were quite different. His findings showed a value of about 1 mm for S. alterniflora and 2.5 mm for S. townsendii. This feature appears to vary somewhat more than Jovet has indicated. Ligule lengths for S. alterniflora were found to vary between 0.7--2 mm, while those of S. townsendii vary between 1.5--3 mm.

Length of spikelets has been used successfully as a key character. Spikelets of the European S. alterniflora do not exceed 15 mm in length, while those of S. townsendii are always 16 mm or more in length.

The degree of pubescence on the glumes also serves to separate S. alterniflora from S. townsendii, which resembles S. maritima in this regard. Similarly, comparison of the keel of the lemmas offers a fair degree of differentiation. The lemma keels of S. townsendii are characteristically pilose at least on the upper two-thirds, while those of S. alterniflora are essentially glabrous except for the few soft trichomes near the tip.

Comparison of anther lengths shows a discrete difference between the two populations. Those of S. alterniflora are not found in excess of 6 mm while those of S. townsendii exceed 7 mm.

Stapf (1908) used the nature of the second glume tip as a taxonomic key character. He states that the keel of the second glume is not prolonged beyond the glume tip in S. townsendii while it is prolonged in S. maritima and S. alterniflora. Examination of a number of spikelets of S. townsendii shows that this feature is not constant. Plate III, g-h show the variety of second glume tips encountered. It is evident that these glume tips are, in a large measure, similar to those of S. maritima by virtue of the distinct separation of keel and glume proper. Hence, this feature may be used to separate S. townsendii from S. alterniflora but not from S. maritima.

Separation of S. townsendii from S. maritima is easily accomplished, however. Spikelets of S. maritima do not exceed 15 mm, anthers do not exceed 6 mm, and leaf blades do not exceed 12 cm. On the other hand, spikelets of S. townsendii vary from 16--22 mm, anthers are always in excess of 7 mm, and leaf blades exceed 12 cm. Median spikes of specimens of S. townsendii examined during the course of this investigation were found to vary from 10--20 cm in length; those of S. maritima with three or more spikes per panicle varied between 3--10 cm. In several cases involving the two-spiked S. maritima, spike lengths up to 13 cm were found.

Of some taxonomic significance is the length by which the palea exceeds the lemma in S. townsendii. This length varies in S. townsendii from 1.5--3 mm and in all but two of the twenty-five specimens examined was 2 mm or more. Such lengths measured in the other European species only rarely exceeded 1 mm.

Perhaps the seat of greatest difficulty within the European spartinas is the separation of S. neyrautii from the other species. It has a number of characteristics typical of both S. alterniflora and S. maritima. That it is discrete from S. maritima is evident from a comparison of the

leaves. The blade of the second leaf from the apex exceeds 18 cm in length and is not caducous, quite in contrast to that of S. maritima which does not exceed 12 cm and is readily disarticulated from the sheath.

Comparison of S. neyrautii with S. alterniflora shows that they are indeed similar. At first glance it appears that S. neyrautii is somewhat more robust but quantitative evaluation of certain parts of the inflorescence failed to show any discrete differences. Spike and spikelet length are quite variable in both but are predominantly longer in S. neyrautii. Pubescence of the glumes appears to be somewhat more dense in S. neyrautii but this is an extremely unsatisfactory key character. The only feature which was found useful in effecting a separation of these two related species is the width of the rachis of a median spike. Measurements were made on the flattened adaxial surface of the triangular rachis of a median spike at the approximate center of the spike. Since S. neyrautii has such a restricted distribution, it was compared in this respect only with specimens of S. alterniflora from southwestern France. Values of rachis widths in S. alterniflora varied between 0.5--0.9 mm, while those of S. neyrautii were all in excess of 1.3 mm.

Jovet (1941) has shown that the second glume tips of S. neyrautii and S. alterniflora differ somewhat, those of S. neyrautii agreeing for the most part with S. maritima and S. townsendii. However, the variability encountered in the course of this investigation shows this feature to be a rather unsatisfactory key character. Second glume tips for S. neyrautii are shown in Plate III, e-f. The ligule of S. neyrautii is about 1 mm in length and does not differ significantly from that found in S. alterniflora.

With the exception of panicle width and ligule length, no other characters were found that would enable one to separate S. neyrautii from S. townsendii. At best, panicle width is a rather unsatisfactory criterion since it depends to a large extent upon the degree of appression of the spikelets. Spikelets of S. townsendii showed some variation in this respect. A number of specimens with distinctly spreading spikelets were encountered as were some with spikelets rather tightly appressed. Spikelets of S. neyrautii were found to be tightly appressed. The ligule length in S. neyrautii appears to be quite variable in that values from 0.7--1.3 mm were found. In contrast those of S. townsendii vary from 1.5--3 mm in length. Since ligule length has been shown to vary considerably throughout the genus, diagnosis on this feature alone does not seem advisable.

Perhaps geographic separation is the simplest way in which identification may be made. The two species are not known to overlap as yet, although S. townsendii has shown startling abilities in the matter of migration and subsequent establishment.

KEY TO SPECIES OF SPARTINA
OCCURRING IN SOUTH AMERICA

1. Spikes more than 15; panicle spikelike, cylindrical. 2
1. Spikes less than 15; panicle not spikelike, not usually cylindrical. 3
2. Spikelets 8 mm or less in length, lanceolate; panicle narrow, not exceeding 1 cm in diameter; pubescence on glumes and lemmas hispid. 3. Spartina spartinae
2. Spikelets 8 mm or more in length, ovate; panicle thick, exceeding 1 cm in diameter; pubescence on glumes and lemmas long-pilose. 2. Spartina ciliata
3. Plants lacking rhizomes; spikelets 8--12 mm in length; pubescence on keels of glumes and lemmas of short, rigid, slender trichomes; spikes usually diminishing in length from base to apex of the panicle. 13. Spartina densiflora
3. Plants with rhizomes; spikelets usually exceeding 12 mm in length; pubescence on keels of glumes and lemmas of long, soft trichomes, or rarely wanting; spikes usually subequal in length. 4
4. Median spike rachis exceeding 1 mm in width; ratio of spike length to spikelet number usually 3.5--4.5. . 6. Spartina longispica
4. Median spike rachis seldom exceeding 1 mm in width; ratio of spike length to spikelet number usually exceeding 4.5. 4. Spartina alterniflora

Discussion.

Identification of South American members of the genus may be accomplished with relative ease in most cases. The more uniform of the species found in South America are S. ciliata and S. spartinae. They are easily distinguished from the other species on the bases of numerous short spikes, always in excess of 15 per panicle and not exceeding 8 cm in length. The spikes are densely imbricate and usually twisted, giving the entire panicle a smooth, cylindrical contour. The two species are separable from each other on the basis of spikelet shape and pubescence. Spikelets in S. ciliata are ovate and the keels of glumes and lemmas possess soft trichomes which may exceed 1.0 mm in length. Spikelets of S. spartinae are lanceolate and hispid with rigid trichomes which do not exceed 0.5 mm in length.

S. ciliata appears to be rather closely related to S. arundinacea which occurs on two widely separated island groups in the South Atlantic and Indian Oceans. Superficially, the two are somewhat alike but they are separable on the basis of spikelet shape and pubescence. S. arundinacea has lanceolate spikelets with hispid pubescence.

S. spartinae in South America is rather easily separated from the other

species by virtue of its geographic distribution. Limited to Paraguay and the inland regions of Argentina, its range of distribution is not coincident with that of the other species. While specimens of S. spartinae from South America examined during the course of this study all possessed panicles with spikes in excess of 15, a number of North American specimens were found to have fewer than 15 spikes. It is entirely possible that such depauperate specimens might be found in South America. If this be true, some confusion could result in attempting to distinguish between S. spartinae and S. densiflora. That some confusion did exist is evident from the perfunctory treatment given the two species by Saint-Yves (1932). He considered them only as varieties of a single species, S. densiflora. His separation of S. densiflora varieties typica and junci-formis (our S. spartinae) was based upon the cylindrical appearance of S. spartinae and the length of the panicle and spikelet. The cylindrical aspect of the panicle is certainly distinctive for the more typical specimens of S. spartinae, but it is less apparent in panicles with fewer than 20 spikes. S. densiflora, on the other hand, is not typically cylindrical except where a near maximum number (15) of spikes occurs. Saint-Yves further states that the panicle lengths do not exceed 15 cm in S. densiflora and that the spikelets are 10--11 mm in length. On the other hand, in S. spartinae, he sets 20--35 cm as the limits for panicle length and 6--7 mm for spikelet length. Examination of over one hundred specimens shows that these criteria of Saint-Yves are not answered. A specimen of S. spartinae (Rojas 2269, Lima Pora Chaco, Par. US) was observed to possess two panicles, one 14 cm long, and the other 39 cm long.

S. densiflora and S. spartinae, as defined in this paper, can be shown to be separable. The spikelets of S. densiflora are in excess of 8 mm in length, but in none of the South American specimens of S. spartinae were spikelets longer than 7 mm encountered. Further, the pubescence on the keels of glumes and lemmas differs. In S. densiflora, the pubescence on the spikelets is made up of short, rigid, slender trichomes which are about one-half as long as the thicker trichomes of S. spartinae. Most commonly, in S. densiflora, the first glume is slender and about one-half as long as the second. On the other hand, the first glume of S. spartinae is only rarely less than 2 mm shorter than the first.

Perhaps the source of greatest difficulty within the South American species is the separation of S. longispica from S. alterniflora and S. densiflora. That the latter two are discrete and easily identified has been shown in connection with the hybrid index evaluating the hybrid origin of S. longispica. Hard culms, involute leaves, lack of rhizomes and shorter spikelets all serve to aid in distinguishing S. densiflora from the more succulent-culmed, flat-leaved, rhizomatous S. alterniflora with its longer spikelets. On the same bases, S. densiflora seems to be separable from S. longispica.

However, S. longispica is distinguished from S. alterniflora only with considerable difficulty. As noted in the discussion of the hybrid origin of S. longispica, it was observed to be considerably more robust than the related S. alterniflora. While mean values of panicle and leaf length appear to vary significantly (see Table 5), the overlap encountered renders these characters useless as key criteria.

The only feature which seemingly sets S. alterniflora apart from S.

longispica is the width of the rachis. This structural feature may be perfectly satisfactory, but it would seem inadvisable to consider a specimen properly diagnosed on this basis alone. Perhaps evaluation by means of the characters enumerated in the hybrid index offers the most suitable method of identification.

KEY TO SPECIES OF SPARTINA
OCCURRING IN NORTH AMERICA

1. Culms soft and fleshy; rhizomes flaccid, the scales closely imbricate and inflated; pubescence when present usually pilose; blades usually flat, glabrous, margins glabrous, occasionally scabrous. 2
1. Culms indurate and firm; rhizomes when present indurate and firm, the scales closely imbricate or remote; pubescence usually hispid (long-pilose in S. gracilis); blades flat or involute, glabrous or scabrous, margins scabrous. 3
2. Median spikes usually less than 5.5 cm in length; glumes commonly curved; spikes tightly appressed and often twisted, usually more crowded toward the apex of the panicle; California and Baja California. 5. Spartina foliosa
2. Median spikes usually more than 5.5 cm in length; glumes straight; spikes not tightly appressed nor twisted, not more crowded toward the apex of the panicle; eastern North America, introduced in Oregon and Washington. . . . 4. Spartina alterniflora
3. Rhizomes wanting; panicle spikelike; spikes commonly more than 15, densely imbricate, tightly appressed and twisted, giving the panicle a smooth cylindrical contour; spikelets not exceeding 10 mm in length. 3. Spartina spartinae
3. Rhizomes present or absent; panicle not spikelike nor smoothly cylindrical; spikes few to many, remote or closely imbricate, appressed or spreading; spikelets 5--25 mm in length. . . . 4
4. Second glume with prominent lateral nerves, the nerves all disposed on one side of the keel; spikes fewer than 15; rhizomes when present slender, whitish. 5
4. Second glume with or without prominent lateral nerves, if prominent, the nerves disposed one on either side of the keel; spikes 3--70, rhizomes when present, thick and purplish-brown in color. 8
5. Rhizomes wanting; culms caespitose, in clumps to 1 meter across; culms and sterile shoots arising from hard, knotty bases. 6

5. Rhizomes present; culms usually solitary, occasionally in small clumps of several culms; sterile shoots arising from the rhizomes. 7
6. Culms exceeding 1.3 meters in height; spikes appressed; Florida and Georgia; anthesis usually December to May. 10. Spartina bakeri
6. Culms less than 1.3 meters tall; spikes spreading; New England to Maryland; anthesis July to October. 11. Spartina X caespitosa
7. Spikelets ovate to lanceolate, conspicuously ciliate (pilose) on the keels; floret commonly equalling the second glume; spikes appressed; mountains and plains of western North America. 14. Spartina gracilis
7. Spikelets linear to lanceolate, hispid on the keels; floret shorter than the second glume; spikes spreading, rarely appressed; salt marshes of eastern North America. 15. Spartina patens
8. Second blade from the summit usually involute, not exceeding 7 mm in width; spikes fewer than 10; rhizomes present or absent. 11. Spartina X caespitosa
8. Second blade from the summit flat, usually exceeding 7 mm in width; spikes mostly more than 10; rhizomes present. 9
9. Second glume long-awned, strongly pectinate on the keel, glabrous or rarely hispid on the lateral nerves; lemma tip acuminate, sharply bilobed. 16. Spartina pectinata
9. Second glume not awned, hispid on the keel and usually on the two lateral nerves; lemma tip obtuse or rounded, not sharply bilobed or only shallowly lobed. 12. Spartina cynosuroides

Discussion

The North American species of Spartina, in some cases, present rather formidable obstacles to accurate identification. For some of the specimens at hand, the extant keys seem to serve quite well but very often specimens are found which do not answer to key criteria.

Specimens of S. spartinae are rather readily distinguished from other species on the basis of the dense, cylindric inflorescence. However, in cases where the spikes are few in number, this species approaches S. bakeri, S. gracilis, and S. patens, at least superficially. It is, however, immediately distinguished from all of these by the absence of prominent lateral nerves on the second glume. If lateral nerves are at all evident, they are always disposed on opposite sides of the keel. The two or more lateral nerves on second glumes of S. bakeri, S. gracilis, and S. patens are found on one side of the keel.

S. gracilis may be readily separated from the other North American

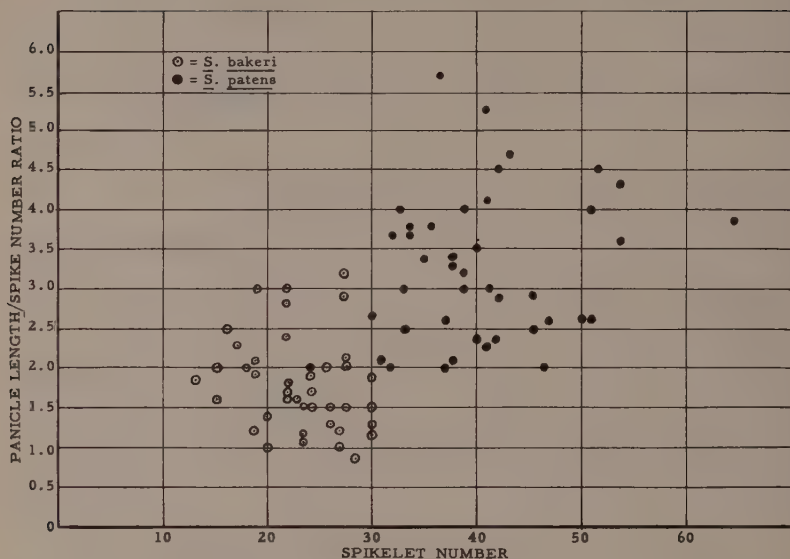


Fig. 26. Comparison of specimens of *Spartina bakeri* and *Spartina patens* by ratio of panicle length to spike number and spikelet number.

species by virtue of the copious pubescence in the form of long pilose or villous trichomes. The range of distribution of *S. gracilis* is coincident only with that of *S. pectinata* from which it may be distinguished not only by the pubescence but also by the disposition of lateral nerves on the second glume.

A close relationship between *S. foliosa* and *S. alterniflora* is suggested by the difficulty encountered in separating them. No single character seems to be perfectly satisfactory. Length of the median spike appears to offer the best criterion, although a median spike length of less than 5.5 cm is characteristic of depauperate specimens of *S. alterniflora*. Spike number is rarely less than six in *S. foliosa*. Two of the more than one hundred specimens examined had three elongate spikes. In *S. alterniflora*, spike number commonly varies from three to fifteen. In *S. foliosa*, the spikes are noticeably more crowded toward the apex of the panicle; in general, the two lower spikes are somewhat remote. This arrangement of spikes is not particularly evident among specimens of *S. alterniflora*. The curving of the glumes, especially the first, is a general trend in most of the specimens of *S. foliosa* examined; this curving was not observed among specimens of *S. alterniflora*.

Spartina bakeri is completely separable from its congeners only upon the basis of its unique vegetative habit. Very often, information on herbarium sheet labels is lacking concerning this particular feature and recourse must be made to other characters which are taxonomically less certain. The only species with which *S. bakeri* is likely to be confused

is the highly variable S. patens. Samples of both populations within the geographic range of S. bakeri were studied. Panicle length-spike number ratios were plotted against the number of spikelets from a median spike in Fig. 26. The specimens of S. patens used in the study were largely those of the author's Florida, Georgia, and South Carolina collections, while those of S. bakeri were largely herbarium specimens from the Florida University Herbarium at Gainesville, Florida. Only those specimens of S. bakeri which were satisfactorily identified by means of vegetative habit, time of flowering and geographic location were used in this numerical analysis.

It is evident from the scatter diagram that the two populations tend to differ regarding the characters employed, although some overlap does occur. It should be noted, however, that these criteria hold only for S. patens from the southeastern United States for, as shown in connection with S. patens as a whole, a considerable number of specimens from the more northerly segment of the range possess many fewer than thirty spikelets per spike.

Spartina pectinata is rather readily distinguished from other species of the genus except perhaps S. cynosuroides. These two are somewhat similar and are doubtless closely related. Their range of distribution coincides along the eastern seaboard of North America from Massachusetts south to Delaware. In this area, the two species are usually ecologically segregated. S. cynosuroides inhabits the tidal marshes and lagoons while S. pectinata is ordinarily restricted to the drier parts of the salt marshes, well above the intertidal zone.

Morphologically, the two are found to be separable on several counts. Perhaps the most prominent feature is the awned or aristate second glume of S. pectinata. That of S. cynosuroides is, by contrast, merely acuminate. While it is difficult to draw a fine line of distinction between aristate and acuminate, most specimens of S. pectinata possess enough of an awn to facilitate identification. In the more difficult cases, recourse may be had to several other fairly distinctive character differences. The base of the rachis of the spikes in S. cynosuroides is somewhat contorted and knotty, while that of spikes of S. pectinata is perfectly straight and smooth or scabrous. The lemma tip in S. pectinata is acute, and distinctly bilobed, while that in S. cynosuroides is obtuse or rounded and not markedly bilobed. A comparison of the pubescence on the keels and nerves of the second glumes usually shows a difference. The pubescence is strongly pectinate and usually limited to the keel in S. pectinata, but in S. cynosuroides it is hispid, shorter in length, and distributed along the lateral nerves as well as the keel.

Perhaps the seat of greatest taxonomic difficulty in the North American species concerns S. X caespitosa. Because of its polyphyletic hybrid origin, it exhibits various combinations of Characters of both S. patens and S. pectinata. The absence of rhizomes and the caespitose habit are adequate criteria for distinguishing some members of the hybrid swarm. However, in dealing with those that possess rhizomes, no single character allows a satisfactory separation from the S. pectinata parent. These rhizomatous members of the hybrid swarm may best be identified by evaluation of the characters listed in the hybrid index constructed for S. X caespitosa.

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REFERENCES

- Aiton, William. *Hortus Kewensis*. Vol. 1. London. 1789.
- Anderson, Edgar. Hybridization in American *tradescantias*. I. A method for measuring species hybrids. II. Hybridization between *T. virginiana* and *T. canaliculata*. *Ann. Missouri Bot. Gard.* 23:511-525. 1936.
- _____. Introgressive hybridization. New York, J. Wiley. 1949.
- Arber, Agnes. *The Gramineae*. New York, The MacMillan Co. 1934.
- Arechavaleta, J. *Las Gramineas Uruguayas*. *Anal. Mus. Nac.* Montevideo 4:213-581. 1896.
- Beal, W. J. *Grasses of North America*. Vol. 2. New York, Henry Holt and Company. 1896.
- Beauvois, Palisot de. *Essai d'une nouvelle agrostographie*. Paris. 1812. Original not seen: cited in Niles, Cornelia D. A bibliographic study of Beauvois' agrostographie. *Contr. U.S. Natl. Herb.* 24:162. 1925.
- Bentham, George. Botany of the voyage of H. M. S. *Sulphur*. Edited by Hinds, Richard Brindley. London, Smith, Elder and Company. 1844.
- _____. Notes on *Gramineae*. *Jour. Linn. Soc.* 19:14-134. 1881.
- Bews, J. W. *The World's Grasses*. New York, Longmans, Green and Company. 1929.
- Blomquist, H. L. *The Grasses of North Carolina*. Durham, N. C., Duke University Press. 1948.
- Britton, N. L., E. E. Sterns, and Justus F. Poggenburg. Preliminary Catalogue of *Anthophyta* and *Pteridophyta* reported as growing spontaneously within one hundred miles of New York City. Compiled by a committee of the Torrey Botanical Club. Columbia College Herbarium. 1888.
- Brongniart, Adolphe. Botanique (Phanerogamie) in: Du Perry, Louis Isidore. *Botanique voyage la Coquille autour du monde*. Paris, A. Bertrand. 1829.

- Brotero, Felix. Flora Lusitanica. Olissipone. 1804. Original not seen: cited in Swallen, J.R. Tribe Chlorideae. North Amer. Fl. 17:583. 1939.
- Candolle, Augustin de. See de Candolle.
- Carmichael, Dugald. Some account of the Island of Tristan de Cunha and of its natural productions. Trans. Linn. Soc. 12:483-513. 1819.
- Chapman, A.W. Flora of the Southern United States. New York, Ivison, Phinney and Company. 1860.
- _____. Flora of the Southern United States. 3rd ed. New York, American Book Company. 1897.
- Chase, Agnes. Manual of the grasses of the United States. 2nd ed. U.S. Dept. Agric. Misc. Publ. 200. Washington, U.S. Government Printing Office. 1950.
- Chater, E.H. and H. Jones. New forms of Spartina townsendii. Nature 168:126. 1951.
- Chevalier, M. Aug. Note sur les Spartina de la flore française. Bull. Soc. Bot. France 70:54-63. 1923.
- _____. Nouvelles observations sur les Spartina et spécialement sur le S. townsendii. Bull. Soc. Bot. France 80:779-788. 1933.
- Christophersen, Erling. Plants of Gough Island. Scientific results of the Norwegian Antarctic expeditions. 13:1-13. 1934.
- Church, George L. Cytological studies in the Gramineae. Amer. Jour. Bot. 23:12-15. 1936.
- _____. Cytotaxonomic studies in the Gramineae. Spartina, Andropogon and Panicum. Amer. Jour. Bot. 27:263-271. 1940.
- Clapham, A.R., T.G. Tutin and E.F. Warburg. Flora of the British Isles. Cambridge, University Press. 1952.
- Curtis, Moses A. Enumeration of plants growing spontaneously around Wilmington, North Carolina. Boston Jour. Nat. Hist. 1:82-141. 1835.
- Curtis, William. Enumeration of the British grasses. Lambeth-Marsh, England. 1787. Original not seen: cited in Fernald, M.L. Some notes on Spartina. Rhodora 18:179. 1916.
- de Candolle, Augustin Pyramus and Jean Baptiste Lamarck. Flore française. Vol. 3. Paris, H. Agasse. 1805.
- _____. Flore française. Vol. 6. Paris, H. Agasse. 1815.
- Dobzhansky, Theodosius. Genetics and the Origin of Species. New York, Columbia University Press. 1937.
- Du Petit-Thouars, Louis Marie Aubert. Esquisse de la flore de Tristan d'Acugna. Mélanges de botanique et des voyages. Paris. 1811.
- Eaton, A.A. Two new species. Bull. Torrey Bot. Club 25:338-341. 1898.
- Englemann, George and Asa Gray. Plantae Lindheimerianae. Boston Jour. Nat. Hist. 5:210-264. 1847.
- Fabre, Esprit. Description d'une nouvelle espèce de Spartina. Ann. Sci. Nat. Bot. Paris. III. 13:122-124. 1849.
- Farwell, Oliver A. Notes on the Michigan flora II. Rept. Mich. Acad. 21:345-371. 1920.
- Fassett, Norman C. Grasses of Wisconsin. Madison, University of Wisconsin Press. 1951.
- Fernald, M.L. Some notes on Spartina. Rhodora 18:177-180. 1916.
- _____. The slender-spiked Spartina pectinata. Rhodora 35:258-260. 1933.

- _____. Another century of additions to the flora of Virginia. *Rhodora* 43:485-553. 1941.
- _____. Additions to and subtractions from the flora of Virginia. *Rhodora* 49:86-115. 1947.
- _____. Gray's Manual of Botany. 8th ed. New York, American Book Company. 1950.
- Foucaud, M. Un Spartina nouveau. *Ann. Soc. Sci. Nat. Rochelle* 31:8. 1894.
- Fournier, Eugene. *Mexicanarum Plantarum*. Vol. 1. Part 2. Parisiis, Reipublicae. 1886.
- Gmelin, Johann Friedrich. *Systema Naturae*. 13th ed. Lipsiae, Georg Emanuel Beer. 1791.
- Gray, Asa. A Manual of Botany of the Northern United States. Boston and Cambridge, James Munroe and Company. 1848.
- _____. Manual of the Botany of the Northern United States. 2d ed. New York, George P. Putnam and Company. 1856.
- Gronovius, Jan Fredrik. *Flora Virginica*. Lugduni Batavorum, Luchtmans. 1743. Original not seen: cited in Hitchcock, A.S. Types of American grasses. *Contr. U.S. Natl. Herb.* 12:129. 1908.
- Groves, H. and J. Notes on plants received. *Bot. Exch. Club Rept.* for 1880. pp. 29-38. 1881.
- Hackel, Eduard. The True Grasses. A translation from a section of *Die natürlichen Pflanzenfamilien* by F. Lamson-Scribner and Effie Southworth. New York, Henry Holt and Company. 1890.
- _____. Neue Gräser. *Oesterr. Bot. Zeits.* 52:237-240. 1902.
- _____. Ex herbario Hassleriano. *Novitates paraguariensis*. I. Fedde *Rep. Spec. Nov. Reg. Veg.* 6:341-352. 1909.
- Hasskarl, Justus Karl. *Catalogus plantarum in Horto botanico Bogoriensi cultarum alter. Bataviae, officinae publicae*. 1844.
- Hemsley, William Botting. Report on the botany of the Bermudas and various other islands. *Rept. on the scientific results of the voyage of H.M.S. Challenger* 1:133-185. 1884.
- Hitchcock, A.S. Notes on grasses. *Rhodora* 8:205-212. 1906.
- _____. Types of American grasses. *Contr. U.S. Natl. Herb.* 12:113-158. 1908.
- _____. Mexican grasses in the United States National Herbarium. *Contr. U.S. Natl. Herb.* 17:181-389. 1913.
- _____. Manual of grasses of the United States. U.S. Dept. Agric. Misc. Publ. 200. Washington, U.S. Govt. Printing Office. 1935.
- Hooker, Joseph D. and B. Daydon Jackson. *Index Kewensis*. Vol. II. Oxford, The Clarendon Press. 1895.
- Huskins, C. Leonard. The origin of Spartina Townsendii. *Genetica* 12:531-538. 1931.
- Hutchinson, J. The Families of Flowering Plants. II. Monocotyledons. London, Macmillan and Company, Ltd. 1944.
- Irving, W. Spartina cynosuroides var. aureo-marginata. *Gard. Chron.* III. 38:372. 1905.
- Jovet, Paul. Notes systématiques et écologiques sur les Spartines du Sud-Ouest. *Bull. Soc. Bot. France* 88:115-123. 1941.
- Kunth, C.S. Révision de graminees. in Humboldt, Alexander von. *Voyage de Humboldt et Bonpland*. Part 6. Vol. 5. Botanique. Paris, F. Schoell. 1829.

- _____. Enumeratio Plantarum. Agrostographia synoptica. Stutgardiae et Tubingae, J.G. Cottae. 1833.
- Lanjouw, J. and F.A. Stafleu. Index Herbariorum. Utrecht, Kemink and Zoon. 1952.
- _____, et al. International Code of Botanical Nomenclature. Utrecht, Kemink and Zoon. 1952.
- Larisey, Mary Maxine. Analysis of a hybrid complex between Baptisia leucantha and Baptisia viridis in Texas. Amer. Jour. Bot. 27:624-628. 1940.
- Link, H.F. Der botanische Garten bei Berlin und die Willdenowsche Kräutersammlung. Jahrb. Gewächsk. I. 3:13-93. 1820.
- Linnaeus, Carolus. Species Plantarum. Vol. 1. Holmiae, Laurentii Salvii. 1753.
- Loefling, Peter. Iter Hispanicum, Stockholm. 1758. Original not seen: cited in Fernald, M.L. Some notes on Spartina. Rhodora 18:179. 1916.
- Loiseleur-Deslongchamps, Jean. Flora Gallica. Lutetiae, Migueret. 1807.
- Marie-Victorin, Frere. Quelques entités phanerogamiques mineures de la flore du Quebec. Nat. Canad. 71:201-209. 1944.
- Merrill, Elmer D. The North American species of Spartina. Bull. U.S. Dept. Agric. Pl. Ind. 9:1-15. 1902.
- Michaux, Andre. Flora boreali-americana. Vol. 1. Paris, fratres Leversault. 1803.
- Muhlenberg, Henry. Descriptio uberior graminum et plantarum calamariarum Americae septentrionalis. Philadelphia, Solomon W. Conrad. 1817.
- Nees von Esenbeck, Christian Gottfried Daniel. Florae Africae australioris. I. Gramineae. Glogaviae, Prausnitzianis. 1841.
- Niles, Cornelia D. A bibliographic study of Beauvois' agrostographie. Contr. U. S. Natl. Herb. 24:135-214. 1925.
- Nuttall, Thomas. The Genera of North American Plants. Philadelphia, D. Heartt. 1818.
- Oliver, F.W. Spartina Townsendii; its mode of establishment, economic uses and taxonomic status. Jour. Ecol. 13:74-91. 1925.
- Parlatore, Filippo. Flora Italiana. Vol. 1. Firenze, le Monnier. 1848.
- Parodi, L.R. Las Chlorideas de la República Argentina. Rev. Fac. Agron. Vet. Buenos Aires 2:231-335. 1919.
- _____. Algunas gramineas nuevas para la flora Uruguay. Rev. Sudam. Bot. Montevideo 4:177-178. 1937.
- _____. Personal communication. 1953.
- Petiver, James. Graminum concordia. Londini. 1716. Original not seen: cited in Saint-Yves, Alfred. Monographia Spartinarum. Cand. 5:96. 1932.
- Pohl, Richard W. A taxonomic study on the grasses of Pennsylvania. Amer. Midland Nat. 38:513-604. 1947.
- _____. A rapid softening agent for dried plant structures. Proc. Iowa Acad. Sci. 61:149-150. 1954.
- Poiret, Jean. In Lamarck, M. Encyclopedie methodique botanique. Suppl. IV. Paris, Agasse. 1816.
- Prat, Henri. La systematique des graminees. Ann. Sci. Nat. Bot. X. 18:165-258. 1936.

- Raddi, Guiseppe. *Agrostografia Brasiliensis*. Atti-Della Reale Accad. 2:326-384. 1823. Reprinted separately with change of pagination as *Agrostografia Brasiliensis*. pp. 1-58. 1823.
- Raspail, M. *Classification générale des Graminees*. Ann. Sci. Nat. Bot. Paris. I. 5:287-311. 1825.
- Ray, John. *Synopsis methodica stirpium britannicarum*. 3d ed. Londini, Innys. 1724. Original not seen; cited in Fernald, M. L. Some notes on *Spartina*. *Rhodora* 18:179. 1916.
- Richard, Claude. In Persoon, Christiaan Hendrik. *Synopsis Plantarum*. Vol. 1. Parisiis, Lutetiorum, C.F. Cramerum. 1807.
- Rickett, H.W. Personal communication. 1953.
- Roth, Albrecht Wilhelm. *Neue Beitrage zur Botanik*. Frankfurt am Main, Wilmans. 1802.
- _____. *Catalecta Botanica*. Lipsiae, Gleditsch. 1806.
- Saint-Yves, Alfred. *Monographia Spartinarum*. Cand. 5:19-100. 1932.
- Schreber, Johann Christian Daniel. *Genera Plantarum*. Francofurti ad Moenum, Varrentrapii et Wenneri. 1789.
- Scribner, F.L. Notes on *Spartina*. Bull. Torrey Club 10:85-86. 1883.
- Small, John Kunkel. *Manual of the Southeastern Flora*. New York, published by the author. 1933.
- Spegazzini, Carlo. Description of *Spartina patagonica*. Rev. Fac. Agron. Vet. La Plata. p.579. 1897.
- Stapf, Otto. *Spartina Townsendii*. Gard. Chron. III. 43:33-35. 1908.
- Steudel, Ernst Gottlieb. *Nomenclator Botanicus*. 2d ed. Stuttgartiae et Tubingae, J.G. Cottae. 1841.
- _____. *Synopsis Plantarum Glumacearum*. Part I. *Gramineae*. Stuttgartiae, J.B. Metzler. 1855.
- Swallen, Jason R. The grasses of the Yucatan Peninsula. Contr. Amer. Arch. 2:323-355. 1932.
- _____. Tribe *Chlorideae*. North Amer. Fl. 17:581-638. 1939.
- Thouars, See Du Petit-Thouars.
- Trinius, C.B. *Genera Graminearum*. V. *Agrostideae*. Mem. Acad. St. Petersburg Sci. Nat. VI. 6:23-134. 1840.
- Vasey, George S. A neglected *Spartina*. Bot. Gaz. 16:292. 1891.
- Walter, Thomas. *Flora Caroliniâna*. London, Fraser. 1788. Original not seen: cited in Swallen, J.R. Tribe *Chlorideae*. North Amer. Fl. 17:582. 1939.
- Willdenow, Karl Ludwig. *Enumeratio Plantarum*. Berolini, in taberna libraria Scholae realis. 1809.

PUBLICATIONS OF MEMBERS OF THE STAFF OF THE IOWA STATE COLLEGE FOR THE ACADEMIC YEAR 1954-55

Certain summaries and indices are of interest in a survey of the publications of members of the staff of an educational and research institution such as the Iowa State College. The publications are listed in alphabetic order under the names of the senior authors. Junior authors are also listed alphabetically with cross reference to senior author.

SUMMARY

Number of individuals listed.	701
Number of publications.	725
Number of publications with single author	308
Number of publications with joint authorship.	417
Number of departments or fields represented in publications	45
Number of individuals who serve as editors or on the editorial staff of one or more scientific or technical periodicals.	42

The individuals thus serving are:

Ayres, J.C., Ayres, Q.C., Becker, Biester, Bolton, Brozier, Buchanan, Burroughs, Dahm, Davies, A.W., Diehl, Douglas, Dwelle, Eppright, Fassel, Getty, Gilman, Gowen, Graebner, Harris, Hartley, Hazel, Heady, Heath, Heer, Hendrickson, Iverson, Johnson, I.J., Jones, W.P., Kirkham, Keutemeyer, Loomis, Lunden, Parks, W.R., Riecken, Ross, Snedecor, Sprague, Swanson, Tintner, Wenkert, Werkman.

INDEX TO PUBLICATIONS BY DEPARTMENTS OR FIELDS

The numbers which follow the names of the departments refer to the index number of the alphabetic list by authors.

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Agricultural Engineering: Total 18 - Numbers 7, 11, 67, 103, 104, 149, 266, 284, 285, 341, 400, 401, 402, 403, 557, 598, 664, 680.	Botany: Total 67 - Numbers 20, 34, 42, 43, 44, 73, 75, 91, 92, 129, 159, 172, 182, 217, 218, 220, 240, 241, 242, 253, 254, 262, 298, 307, 310, 312, 335, 341, 384, 390, 391, 395, 396, 408, 420, 469, 470, 472, 488, 489, 490, 491, 515, 529, 561, 566, 597, 604, 617, 618, 619, 620, 621, 622, 655, 658, 668, 669, 697, 698, 699, 700, 701, 702, 703, 704, 720.
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AUTHORS AND PUBLICATIONS

1. Aandahl, A.R., W.G. Murray, and W.H. Scholtes. Economic rating of soils for tax assessment. *Jour. Farm Econ.* 36:483-499. 1954.
Acker, D.C., joint author. *See under Ashton, Jensen, A.H.*
2. Albert, W.W., U.S. Garrigus, R.M. Forbes, and W.H. Hale. Modified urea supplements with corn silage for wintering ewe lambs. *Jour. Animal Sci.* 14:143-152. 1955.
Alexander, L.T., joint author. *See under Muckenhirn.*
Allaway, W.H., joint author. *See under Hammond.*
3. Allen, R.S., P.G. Homeyer, and N.L. Jacobson. Use of activated glycerol dichlorohydrin for estimating vitamin A in dairy calf blood plasma. *Iowa State Coll. Jour. Sci.* 29:721-734. 1955.
4. _____, and N.L. Jacobson. Effect of massive dosage of vitamin A on the blood plasma fat and vitamin A levels in dairy calves. *Proc. Iowa Acad. Sci.* 61:205-209. 1955.
_____, joint author. *See under Jacobson, Lambert, Martin, T.G., Picken, Porter, A.R., Voelker.*
- 4a. Allos, H.F., and W.V. Bartholomew. Effect of available nitrogen on symbiotic fixation. *Soil Sci. Soc. Amer. Proc.* 19:182-184. 1955.
5. Almfeldt, M.W., and Kenneth E. Haughton. *Engineering Drawing Problem Book - Book I.* 70 pp. Wm. C. Brown Co. 1955.
6. _____, _____ *Engineering Drawing Problem Book - Book II.* 90 pp. Wm. C. Brown Co. 1955.
7. Altman, Landy B. Jr., and Emil H. Jebe. Load characteristics of southeastern Iowa farms using electric ranges. *Agric. Expt. Sta. Res. Bull.* 420. 16 pp. 1955.
8. Altshuler, S., and J.F. Carlson. Time dependent variational principle. *Phys. Rev.* 95:546-548. 1954.
9. Anderson, V.L., and O. Kempthorne. A model for the study of quantitative inheritance. *Genetics* 39:883-898. 1954.
10. Anfinson, C.B., W.F. Harrington, A. Hviot, K. Linderström-Lang, M. Ottesen, and J. Schellman. Studies on the structural basis of ribonuclease activity. *Biochim. et Biophys. Acta.* 17:141-142. 1955.
Angus, R., joint author. *See under Willis.*
11. Armstrong, Ray E., and Dale O. Hull. Farm custom rates for 1955. *Iowa Farm Sci.* 9:13-14. 1955.
12. Arnold, L.K., and H.C. Arvidson, Jr. The solvent extraction of fish offal. *Jour. Amer. Oil Chemist's Soc.* 32:163-166. 1955.
13. _____, _____. Solvent extraction of meat offal. *Jour. Amer. Oil Chemist's Soc.* 32:25-29. 1955.
14. _____, and George Burnett, Jr. The determination of small amounts of trichloroethylene in soybean oil meal. *Proc. Iowa Acad. Sci.* 61:210-216. 1954.
15. _____, W.G. Juhl, and H.C. Arvidson, Jr. Densities and viscosities of tri-chloroethylene miscellas of cottonseed oil, fish oil, and beef tallow. *Jour. Amer. Oil Chemist's Soc.* 31:393-394. 1954.
16. _____, _____. The reduction of free gossypol in cottonseed flakes during solvent extraction. *Jour. Amer. Oil Chemist's Soc.* 32:151-152. 1955.
17. _____, _____. Solvent extraction of cottonseed meals. *Jour. Amer. Oil Chemist's Soc.* 31:613-618. 1954.
18. Arnrich, Lotte. The effect of hypothyroidism in the metabolism of carotene in dogs. *Jour. Nutrition* 56:35-50. 1955.
19. _____, and Agnes Fay Morgan. The utilization of carotene by hypothyroid rats. *Jour. Nutrition.* 54:107-120. 1954.
20. Aronoff, S. Translocation from soybean leaves. II. *Plant Physiol.* 30:184-185. 1955.
_____, joint author. *See under Graf, Racusen.*
Arvidson, H.C. Jr., joint author. *See under Arnold.*

21. Ashton, G.C., J. Kastelic, D.C. Acker, A.H. Jensen, H.M. Maddock, E.A. Kline, and D.V. Catron. Different protein levels with and without antibiotics for growing-finishing swine: Effect on carcass leanness 1,2,3. Jour. Animal Sci. 14:82-93. 1955.
_____, joint author. See under Catron, Jensen, A.H.
- Atkins, D.F., joint author. See under Dooley.
- Atkins, R.E., joint author. See under Finkner, Taylor.
- Austrheim, H.A., joint author. See under Bureau.
- Avakian, Souren, joint author. See under Gilman.
22. Ayres, John C. Stability of dehydrated eggs - a symposium II. Salmonella problems. Methodology for isolating Salmonella from dried egg products. Natl. Acad. Sci. - Natl. Res. Coun. pp.74-82. 1954.
23. _____. Quality and stability of canned meats - a symposium II. Microbiological factors affecting canned meats. Natl. Acad. Sci. - Natl. Res. Coun. pp.15-25. 1954.
24. _____. Comparison of fermentation and enzyme treatment methods for the removal of glucose from egg albumen. Bact. Proc. 55th General Meet. New York City. pp.20-21. 1955.
_____, joint author. See under Carlin.
- _____. Associate Editor, Food Technology.
- Ayres, Quincy C., Consulting Editor, McGraw-Hill Agric. Engr. Ser. (12 books)
25. Babbar, M.M., Gerhard Tintner, and Earl Heady. Programming with consideration of variations in input coefficients. Jour. Farm Econ.37:333-341. 1955.
26. Babbitt, Harold E. and E. Robert Baumann. Effect of body feed on the filtration of water through diatomite. Univ. of Illinois Engr. Expt. Sta. Bull. No.425. pp. 1-40. 1954.
_____, joint author. See under Baumann.
- Baker, C.B., joint author. See under Heady.
- Baker, M.L., joint author. See under Dickerson.
- Baker, M.P., joint author. See under Fortney.
27. Balloun, S.L. The effect of quaternary ammonium derivatives in chick diets. Poultry Sci. 34:191-196. 1955.
_____, joint author. See under Donovan.
- Bancroft, T.A., joint author. See under Brindley.
28. Banks, C.V., and R.J. Davis. Analytical applications of the reaction of thorium with benzenephosphonic acid. Analyt. Chim. Acta 12:418-431. 1955.
29. _____. and R.E. Edwards. Separation and determination of thorium and aluminum. Analyt. Chem. 27:947-949. 1955.
30. _____. R.W. VanderHaar, and R.P. VanderWal. Magnetic studies of nickel (II) complexes with some vic-dioximes. Jour. Amer. Chem. Soc. 77:324-325. 1955.
_____, joint author. See under Klingman.
31. Barger, Gerald L. The climate of Iowa. Introduction. Iowa Agric. Expt. Sta. Spec. Rept. No. 7. 7 pp. 1954.
_____, joint author. See under Shaw, R.H.
- Barnes, K.K., joint author. See under Frevert.
32. Barnes, L.E. Oxytetracycline in bovine mastitis 1. Treatment of mastitis. Amer. Jour. Vet. Res. 16:386-390. 1955.
33. _____. Four cases of bovine mastitis caused by Klebsiella pneumoniae. Jour. Amer. Vet. Med. Assoc. 125:50-54. 1954.
_____, joint author. See under Collier.
- Bartholomew, W.V., joint author. See under Allos, Fitts, Johnson, I.J., Kirkham.
34. Bass, Louis N. Factors affecting germination of Kentucky bluegrass seed. Iowa State Coll. Jour. Sci. 28:503-519. 1954.
_____, joint author. See under Delouche, Grabe.
35. Baumann, E. Robert. Iowa sewage works operators' basic training course. League of Iowa Municipalities Monthly Magazine 9:9-12. 1955.
36. _____. and Harold E. Babbitt. The removal of Entamoeba histolytica cysts from water by porous filter septums either with or without filter aid. Univ. of Illinois Engr. Expt. Sta. Bull. No. 431. 52:1-40. 1955.

37. _____, and Charles S. Oulman. The specific heat of garbage. Publ. Health Repts. 80:805-809. 1955.
38. _____, and M.T. Skodje. Garbage cooking in Iowa. Publ. Works Mag. 86:158. 1955.
39. _____, _____. Garbage cooking in Iowa. Publ. Health Repts. 70:314-318. 1955.
_____, joint author. See under Babbitt, Morgan, P.E., and Skodje.
40. Baumann, Ross V., and Earl O. Heady. Costs and returns for soil-conserving farming systems on Ida-Monona soils. Iowa Agric. Expt. Sta. Bull. 429. 1955.
_____, joint author. See under Jensen, H.R.
41. Bauriedel, Wallace R., Joseph C. Picken Jr., and Leland A. Underkofler. Reaction of cyanocobalamin and aquocobalamin with proteins. Fed. Proc. 14:(1), 1955.
42. Baxter, John W. Diseases of forage legumes in Iowa in 1954. Plant Dis. Reporter 39:239. 1955.
43. _____. Species of Puccinia on salvia in Europe, Asia, and Africa. Mycologia 47:225-232. 1955.
44. Beal, Ernest O. and Paul H. Monson. Marsh and Aquatic Angiosperms of Iowa. State University of Iowa Studies in Natural History. 95 pp. 1954.
45. Beal, George M. Are farmers satisfied with their co-ops? Iowa Farm Sci. 9:7-10. 1955.
46. _____. Farmer co-operatives and member satisfactions. Midwest Sociologist 17:47-53. 1955.
47. _____. What makes a satisfied co-op member? News for Farmer Cooperatives 22:12-13. 1955.
48. _____. How can we get more member participation? News for Farmer Cooperatives 22:14-15. 1955.
49. _____, and Richard Phillips. What does participation mean. News for Farmer Cooperatives 22:10-11. 1955.
50. Becker, Elery R. The host affinities of Isospora bigemina - type Coccidia. Proc. Iowa Acad. Sci. 61:463-467. 1954.
51. _____, and William J. Zimmermann. Influence of alcoholic extract of horse kidney on Eimeria tenella infection in chicks. Proc. Iowa Acad. Sci. 60:574-578. 1953.
_____, joint author. See under Pattillo.
_____, Board of Editors. Journal of Protozoology; and Journal of Morphology.
52. Beegle, R.M., P.H. Roberts, N. Howard, J. Smith, I. Pesek, M.H. Britton, et al. Weights of foods eaten per meal by 242 women, 30 to 92 years of age. Michigan Agric. Expt. Sta. North Central Regional Bull. No.37. 32 pp. 1954.
- Beel, John A., joint author. See under King, W., Bernard.
- Bell, M.R., joint author. See under Lambert.
53. Benbrook, Edward A. A life cycle approach to parasite control. Proc. 91st Meet. Amer. Vet. Med. Assoc. pp.247-249. 1954.
54. Beneke, Raymond R., Richard Phillips, and Roy M. Kottmann. Training tomorrow's co-op manager. Co-op Grain Quart. 13:55-61. 1955.
55. Benkeser, R.A., and H. Gilman. Copper in organic chemistry. A chapter in A.C.S. Monograph No.122, "Copper", A. Butts, Ed. Reinhold Publ. Corp. 1954.
_____, joint author. See under Gilman.
56. Bennett, F.W., and F.E. Nelson. Action of aerosols of certain viricidal agents on lactic streptococcus bacteriophage. Jour. Dairy Sci. 37:840-846. 1954.
57. _____, _____. Action of certain viricidal agents on lactic streptococcus bacteriophage in liquids. Jour. Dairy Sci. 37:847-856. 1954.
Bennett, Paul C., joint author. See under Picken,
Beveridge, Elizabeth, joint author. See under Dudley.
58. Beyer, G.H., and R.B. Edwards. Flooding characteristics of a pulse extraction column. Ames Lab. Rept. ISC-553. 28 pp. 1954.
59. _____, and F.M. Jacobsen. Operating characteristics of a centrifugal extractor. Ames Lab. Rept. ISC-548. 39 pp. 1954.
60. _____, D.R. Spink, J.B. West, and H.A. Wilhelm. Caustic treatment of zircon sand. Chem. Engr. Progr. Symposium Ser. 50:67-71. 1954.
Biester, H.E., joint author. See under Picken, Zimmermann.
_____, Associate Editor, The Cornell Veterinarian.

62. Bird, E.W., and O.M. Ystgaard. Some aspects of the detection of watering in milk. *Jour. Dairy Sci.* 38:453-456. 1955.
_____, joint author. *See under Fortney.*
63. Black, C.A., and O. Kempthorne. Willcox's agrobiology: I. Theory of the nitrogen constant 318. *Agron. Jour.* 46:303-307. 1954.
_____, joint author. *See under Bouldin, Heslep, Legg, Mehta, White, Wm.C. Blake, J.T., joint author. See under Jacobson.*
64. Bliese, John C.W. Cover density of a city summer and fall Bronzed Grackle roost. *Proc. Iowa Acad. Sci.* 61:479-491. 1954.
65. _____. Some physical measurements of the Bronzed Grackle roost at Ames, Iowa. *Proc. Iowa Acad. Sci.* 61:468-478. 1954.
66. _____. Thunderstorms, wind direction, and Grackle flights. *Iowa Birdlife* 25: 30-31. 1955.
Blunn, C.T., joint author. *See under Dickerson.*
67. Bockhop, C.W., and Kenneth K. Barnes. Power distribution and requirements of a flail type forage harvester. *Agric. Engr.* 36:453-457. 1955.
68. Boehnke, Roger H. Winter movements and covey composition of the eastern Bob-White. *Proc. Iowa Acad. Sci.* 61:500-503. 1954.
69. Bolton, J.H. The use of good English. The Exponent. (published in a series) Oct. pp.16-23; Nov. pp.16-18; Dec. pp.16-18; 1954. Jan. pp.16-23; Mar. pp.17-24; 1955. (Reprinted as Engr. Report 24, "The Use of Good English in Technical Writing." 1955.
_____, Editor, *Iowa Engr. Expt. Sta. Bulletins and Reports.*
Bonhag, Philip F., joint author. *See under Wick.*
70. Boozer, Charles E., George S. Hammond, Chester E. Hamilton, and Jyotirindra N. Sen. I. Air oxidation of hydrocarbons. II The stoichiometry and fate of inhibitors in benzene and chlorobenzene solutions. *Jour. Amer. Chem. Soc.* 77:3233-3237. 1955.
71. _____. _____, and Conrad Peterson. Air oxidation of hydrocarbons. IV The effects of varying solvent and the mechanism of uninhibited chain termination. *Jour. Amer. Chem. Soc.* 77:3380-3382. 1955.
_____, joint author. *See under Hammond.*
72. Bouldin, D.R., and C.A. Black. Phosphorus diffusion in soils. *Proc. Soil Sci. Soc. Amer.* 18:255-259. 1954.
73. Bowen, C.C. Feulgen staining of cell suspensions. *Stain Technol.* 30:135-138. 1955.
_____, joint author. *See under Wilson, G.B.*
74. Bowlen, Bernard, and Earl O. Heady. Optimum combinations of competitive crops at particular locations. *Iowa Agr. Expt. Sta. Bull.* 426. 1955.
75. Bragonier, W.H., J.D. Dodd, and J.C. Gilman. Episodes in Plant Phylogeny. Wm. C. Brown Co., Dubuque, Iowa, 2nd ed. 1955.
_____, joint author. *See under Johnson, I.J.*
76. Brehm, R.K., and V.A. Fassel. Direct spectrochemical analyses with a rapid scanning spectrometer. *Spectrochim. Acta* 6:341-372. 1954.
77. Bridger, G.L. Fertilizers. *Analyt. Chem.* 27:632-636. 1955.
78. _____. and H.A. Burzlaff. Fertilizer drying. *Jour. Agric. and Food Chem.* 2:1170-1173. 1954.
_____, joint author. *See under Smutz.*
79. Brierly, J.M., J.L. Ellingboe, and Harvey Diehl. Catalytic behavior of vitamin B_{12a} in the oxidation of iodide by air. *Iowa State Coll. Jour. Sci.* 27:425-435. 1953.
80. _____. R.R. Sealock, and H. Diehl. The rate of ammonia production in the acid hydrolysis of vitamin B₁₂. *Iowa State Coll. Jour. Sci.* 29:141-142. 1954.
- 81. Brigham, R.D., and C.P. Wilsie. Seed setting and vegetative vigor of Ladino clover (*Trifolium repens* Leyss) clones and their diallel crosses. *Agron. Jour.* 47:125-127. 1955.
82. Brindley, T.A., H.C. Cox; W.G. Lovely, and T.A. Bancroft. Timing corn borer spray applications. *Proc. Tenth Ann. Meet. North Central Branch Ent. Soc. Amer.* 10:10-11. 1955.
83. _____. et al. Seek better borer control methods. *Your Expt. Sta. Repts.* 1951-53. A rept. of the Iowa Agric. Expt. Sta. pp.7-8. 1954.
_____, joint author. *See under Cox.*
Britton, M.H., joint author. *See under Beegle.*

- Broadbent, H.S., joint author. See under Gilman.
84. Brosier, Mark H. Let's talk fire protection. *Telephony* 149:21-39. 1955.
 85. _____. A new approach to leadership. *Internatl. Fire Fighter*. 38:9-12. 1955.
 86. _____. F.W. Nelson, Wm. R. Simpson, and Keith Roger. 333 Questions and Answers for Firemen. *Engr. Ext.* 2nd ed. 67 pp. 1955.
 - _____. Supervisor Fire Service Information.
 87. Brough, Owen L. and G.S. Shepherd. Objective grade specifications for slaughter barrow and gilt carcasses. *Agric. Res. Bull.* No. 421. 1955.
 88. Brown, F.E., and R.C. Peabody. Cuprous oxide as a catalyst. II. Adsorption by cuprous oxide. *Proc. Iowa Acad. Sci.* 61:233-249. 1954.
 89. _____. and J.D. Woods. Radiochlorine³⁶ as a tracer in the decomposition of potassium chlorate. *Proc. Iowa Acad. Sci.* 61:250-254. 1954.
 - _____. Abstractor for Chemical Abstracts.
 90. Brown, William G. and Earl O. Heady. Economic instability and choices involving income and risk in livestock and poultry production. *Iowa Agric. Expt. Sta. Res. Bull.* 431. 1955.
 - _____. joint author. See under Heady.
 91. Browning, J.A., and K.J. Frey. Stem rust resistant oat strains derived from irradiated seeds. *Phytopath.* 44:483. 1954.
 92. _____. H.C. Murphy, M.D. Simons, and S.C. Wiggins. Contributions from Iowa. *Natl. Oat Newsletter* 5:38-41. 1955.
 93. Bryan, John H.D. Differential staining of tissue components with a mixture of safranin and fast green FCF. *Stain Technol.* 30:153-157. 1955.
 94. Buchanan, R.E. Taxonomy. Chapter in: *Ann. Rev. Microbiol.* 9:1-20. 1955.
 95. _____. S.T. Cowan, and T. Wikén. The bacterial generic names Micrococcus and Staphylococcus. *Internatl. Bull. Bact. Nomen. and Tax.* 4:163-166. 1954.
 96. _____. _____. Amendments to the international code of nomenclature of the bacteria and viruses. *Internatl. Bull. Bact. Nomen. and Tax.* 4:167-188. 1954.
 97. _____. _____. Status of the generic names Escherichia, Klebsiella, Aerobacter, and Cloaca. *Internatl. Bull. Bact. Nomen. and Tax.* 4:159-162. 1954.
 98. _____. _____. The status and synonymy of the bacterial generic name Asterococcus. *Internatl. Bull. Bact. Nomen. and Tax.* 5:13-20. 1955.
 99. _____. _____. Status of the generic names Nitromonas, Nitrosomonas, Nitrosococcus, and Nitrobacter. *Internatl. Bull. Bact. Nomen. and Tax.* 5:28-32. 1955.
 100. _____. _____. The status of the generic names Arthrobacter Fischer and Arthrobacter Conn and Dimmick. *Internatl. Bull. Bact. Nomen. and Tax.* 5:83-84. 1955.
 101. _____. _____. and the Judicial Commission. Status of generic names published without names of included species. *Internatl. Bull. Bact. Nomen. and Tax.* 5:21-26. 1955.
 102. _____. _____. Status of the generic name Selenomonas von Prowazek. *Internatl. Bull. Bact. Nomen. and Tax.* 5:7-12. 1955.
 - _____. Editor, General Bacteriology Section, Biological Abstracts.
 - _____. Chairman Editorial Board, International Bulletin of Bacteriological Nomenclature and Taxonomy.
 - _____. Editor-in-Chief, Iowa State College Journal of Science.
 103. Buchele, W.F., E.V. Collins, and W.G. Lovely. Ridge farming for soil and water control. *Agric. Engr.* 36:324-329, 331. 1955.
 104. _____. _____. Ridge planting saves. . . soil, water. *Iowa Farm Sci.* 9:3-5. 1955.
 - Buchholtz, W.F., joint author. See under Fink.
 105. Buck, Griffith J. Roses. Iowa State College Pamphlet 218. 4 pp. 1955.
 106. _____. and E.C. Volz. A handbook for rose growers. Iowa State College Bull. P-117. 31 pp. 1955.
 - Bulgrin, V.C., joint author. See under Duke.
 - Bundy, Clarence E., joint author. See under Diggins.
 107. Bureau, A.J., H.A. Austrheim, and D.J. Zaffarano. An electron injector for a 70 Mev synchrotron. *Rev. Sci. Instruments* 25:1129-1130. 1954.
 - Burnett, George Jr., joint author. See under Arnold.

- Burroughs, Wise, joint author. See under Cheng, Repp.
 _____, Editorial Board, Journal of Animal Science
 Burzlaff, H.A., joint author. See under Bridger.
 Butler, J.P., joint author. See under Diehl, Goetz.
- Cady, J.G., joint author. See under Ruhe.
108. Campbell, Vincent, and James T. Freeman. Some functions of experimentally-induced language in perceptual learning. *Perceptual and Motor Skills*. 1:71-79. 1955.
109. Carlander, Harriet Bell and Kenneth D. Carlander. Commercial Fishing. Chapter VI, pp.57-70 in: *History of Fish and Fishing in the Upper Mississippi River*, by Harriet Bell Carlander. Upper Miss. River Conser. Com. 96 pp. 1954.
110. Carlander, Kenneth D. Do fish tell time? *The Fisherman* 5:20-22. 1954.
111. _____. Yellow bass population changes at Clear Lake. *Iowa Conservationist* 11:50, 56. 1954.
112. _____. Clear Lake bullheads. *Iowa Conservationist* 11:61, 64. 1954.
113. _____. Length vs weight for common fresh-water game fish. *The Fisherman* 6:96-97. 1955.
114. _____. Books on fish and fishing. *Iowa Conservationist* 14:131. 1955.
 _____, joint author. See under Carlander, H.B.
115. Carlin, Agnes Frances, John C. Ayres, and Paul G. Homeyer. Consumer evaluation of the flavor of angel cakes prepared from yeast-fermented and enzyme-treated dried albumen. *Food Technol.* 8:580-583. 1954.
 _____, joint author. See under Funk.
 Carlson, J.F., joint author. See under Altshuler.
 Carlson, O.N., joint author. See under Wilhelm.
116. Carman, Grant M. Interrelationships of milk production and breeding efficiency in dairy cows. *Jour. Animal Sci.* 14:753-759. 1955.
117. Carneiro, G.G. and J.L. Lush. Reproductive rates and growth of purebred Brown Swiss cattle in Brazil. *Jour. Dairy Sci.* 37:1145-1157. 1954.
- Carr, P.H., joint author. See under Mueller.
118. Carter, Dennis L. Yellow-Crowned Night Heron at Ames. *Iowa Bird Life* 25:40. 1955.
 Carvalho, José C.M., joint author. See under Drake.
 Casas, I., joint author. See under Duncan.
119. Catron, Damon V. Pasture vs. concrete for feeding hogs. *Ctry. Gent. Mag.* July, 1954.
120. _____. New early wean ration brings more pig profits. *Ctry. Gent. Mag.* Sept., 1954.
121. _____. Life cycle feeding program. *Ctry. Gent.* Dec., 1954.
122. _____. Future of hog raising. . . and your hog feed business. *Feed Age*. 8 pp. 1955.
123. _____. A new feeding program for hogs. *Feed Bag Red Book*. 9 pp. 1954.
124. _____. F. Diaz, V.C. Speer, G.C. Ashton, and C.H. Liu. Distillers dried solubles in starter and growing-finishing rations for swine. *Proc. 10th Distillers Feed Conf.* 10:58-63. 1955.
 _____, joint author. See under Ashton, Jensen, A.H.
125. Cavazos, L.F. and R.M. Melampy. Cytometric effects of testosterone propionate on rat seminal vesicles. *Anat. Rec.* 118:286. 1954.
126. _____. A comparative study of periodic acid-reactive carbohydrates in vertebrate testes. *Amer. Jour. Anat.* 95:467-496. 1954.
127. _____. Response of rat accessory glands to different levels of male hormone. *Anat. Rec.* 121:273-274. 1955.
 _____, joint author. See under Melampy.
128. Chadderdon, Hester and Mary S. Lyle. Reasons given by Iowa women for attending homemaking classes for adults. *Agric. Expt. Sta. Spec. Rept.* No.12. 26 pp. 1955.
 Chapman, A.B., joint author. See under Dickerson, Drake.
129. Chapman, H.W., L.S. Gleason, and W.E. Loomis. The carbon dioxide content of field air. *Plant Physiol.* 29:500-503. 1954.
- Charles, Don C., joint author. See under Plumb.

130. Cheng, Edmund W. and Wise Burroughs. Determination of small amounts of diethylstilbestrol in feeds. Jour. Assoc. Offic. Agric. Chemists. pp.146-150. 1955.
131. _____, Lester Yoder, Charles D. Story, and Wise Burroughs. Estrogenic activity of some naturally occurring isoflavones. Ann. New York Acad. Sci. 61:652-659. 1955.
_____, joint author. See under Repp.
132. Chiotti, P. Adaptation of a Geiger-counter X-ray diffractometer for high temperature investigations. Rev. Sci. Instruments. 25:683-688. 1954.
133. _____. Measurement of the electrical resistance of metals and alloys at high temperatures. Rev. Sci. Instruments 25:876-883. 1954.
134. _____. High temperature crystal structure of thorium. Jour. Elect. Soc. 101:567-570. 1954.
135. Christensen, George C. Angioarchitecture of the canine penis and the process of erection. Amer. Jour. Anat. 95:227-262. 1954.
Christensen, P.S. Editor, Iowa Custodian's Newsletter, Eagle Grove Eagle, with the cooperation of I.S.C. Engineering Extension staff.
136. Chu, T.Y. and D.T. Davidson. Deflocculating agents for mechanical analysis of soils. Natl. Acad. Sci. - Natl. Res. Coun. Publ. 343. pp.15-26. 1955.
137. _____, _____, and J.B. Sheeler. Mathematical analysis of a layer extraction method for separating clay-size material from soils. Proc. 2nd Natl. Conf. Clays and Clay Minerals. Natl. Acad. Natl. Res. Coun. Publ. 327. pp. 462-479. 1954.
138. _____, _____, and A.E. Wickstrom. Permeability test for sands. Symposium on Permeability of Soils. Amer. Soc. Testing Materials Spec. Tech. Publ. No.163, pp. 43-55. 1954.
_____, joint author. See under Handy.
139. Chung, N.Y., L. Northrop, R. Getty, and G. Everson. Effect of varying the intake of calcium pantothenate of rats during pregnancy. II. Histological and histochemical studies of the liver, adrenal, duodenum and tibia of the young at birth. Jour. Nutrition 54:97-106. 1954.
_____, joint author. See under Everson.
Clampitt, B.H., joint author. See under Hansen, R.S.
Clar, R.M., joint author. See under Gilman.
140. Claridge, C.A. and C.H. Werkman. Intermediates of the aerobic dissimilation of 2-ketogluconate by *Pseudomonas aeruginosa*. Arch. Biochem. and Biophys. 51:395-401. 1954.
141. _____, _____. Evidence for alternate pathways for the oxidation of glucose by *Pseudomonas aeruginosa*. Jour. Bact. 68:77-79. 1954.
Clark, R.G., joint author. See under Winders.
142. Clifton, E.S., R.J. Jessen, and E.M. Jacobs. Marketing hogs on the Chicago market. Jour. Farm Econ. 36:611-619. 1954.
143. _____, _____, _____. Amount and source of shrink and fill in market hogs. Jour. Farm Econ. 36:688-692. 1954.
_____, joint author. See under Wiegmann.
144. Collier, J.R. and L.E. Barnes. Identification of bacterial pathogens of bovine mastitis by the practicing veterinarian. Iowa State Coll. Vet. 17:137-142. 1955.
Collins, E.V., joint author. See under Buchele.
Cook, T.H., joint author. See under Gilman.
145. Coover, M.S. Safety in college electrical laboratories. Elect. Engr. 74:151-153. 1955.
146. Cotterill, O.J. and A.R. Winter. Egg white lysozyme. II. Effect of thermostabilization. Poultry Sci. 33:1185-1191. 1954.
147. _____, _____. Egg white lysozyme. III. The effect of pH on the lysozyme ovomucin interaction. Poultry Sci. 34:679-686. 1955.
Cowan, John C., joint author. See under Picken, Jr.
Cowan, S.T., joint author. See under Buchanan.
148. Cox, H.C. Recent developments in European corn borer control--granular insecticides. Proc. 10th Ann. Meet. North Central Branch Ent. Soc. Amer. 10:11-12. 1955.
149. _____, T.A. Brindley, W.G. Lovely, J.E. Fahey. Granular insecticides for European corn borer control. Jour. Econ. Ent. 49:113-119. 1955.

- _____, joint author. See under Brindley.
150. Crowley, M.S. and A. Philip Welch. Clay particle dispersion by ultrasons. *Jour. Amer. Ceramic Soc.* 37:433-439. 1954.
 151. Csanyi, Ladis H. Ungraded local aggregates in bituminous paving mixes. *Rural Roads.* 5:15-17. 1955.
 152. _____, and Hon-Pong Fung. A mortar theory for the use of local ungraded aggregates in bituminous mixes. *Proc. Highway Res. Bd.* 34: 1955.
 153. _____, and Robert M. Nady. The design of bituminous paving mixtures by a mortar theory using local ungraded aggregates. *Proc. Assoc. Asphalt Paving Technol.* 24: 1955.
 154. Cuber, John F. and William F. Kenkel. *Social Stratification in the United States* Appleton-Century-Crofts. 1st ed. 359 pp. 1954.
 155. Daane, A.H., R.E. Rundle, H.G. Smith, and F.H. Spedding. The crystal structure of samarium. *Acta Crystallographica* 7:532-535. 1954.
 156. Dachtler, William C., Eileen M. McDonald, Richard Phillips, and David N. Harrington. Costs of storing reserve stocks of corn in country elevators at bin sites and on farms. *Marketing Res. Rept. No. 93*, U.S. Govt. Print. Off. 60 pp. 1955.
 157. Dahm, Paul A. How insecticides kill. *Iowa State Hort. Soc.* 89:96-98. 1954.
 158. _____, and Earle S. Raun. Fly control on farms with several organic thiophosphate insecticides. *Jour. Econ. Ent.* 48:317-322. 1955.
 - _____, joint author. See under Fairchild, Robbins.
 - _____, Editorial Committee, ENTOMA, a directory of insect and plant pest control published by the Entomological Society of America.
 159. Dale, Jim, A.L. McComb, and W.E. Loomis. Chlorosis, mycorrhizae and the growth of pines on a high-lime soil. *Forest Sci.* 1:148-157. 1955.
 160. Dana, F.C., and L.R. Hillyard. *Engineering Problems Workbook.* Wm. C. Brown Co., Dubuque, Iowa. 3rd ed. 112 pp. 1955.
 - Danielson, G.C., joint author. See under Shaw, W.C.
 161. Davey, Harold W. Government intervention in labor disputes. *Labor Law Jour.* 5:739-742, 800. 1954.
 162. _____. *Contemporary Collective Bargaining.* Prentice-Hall, Inc. 1st ed. 532 pp. 1951.
 163. _____. The legislative council movement, 1933-1953. *Amer. Political Sci. Rev.* 47:785-797. 1953.
 164. _____, Edgar M. Jacobs and John Monroe. The growth of American unions: Comment. *Amer. Econ. Rev.* 45:389-390. 1955.
 165. David, H.A., H.O. Hartley, and E.S. Pearson. The distribution of the ratio, in a single normal sample of range to standard deviation. *Biometrika* 41: 482-493. 1954.
 166. Davidson, D.T. and R.L. Handy. Studies of the clay fraction of southwestern Iowa loess. "Clays and Clay Minerals" *Proc. 2nd Natl. Conf. Natl. Acad. Sci. Natl. Res. Coun. Publ.* 327. pp. 190-208. 1954.
 167. _____, and Associates. Further papers on the Wisconsin loess of southwestern Iowa 1953-1954. *Engr. Rept. No. 22*, Iowa Engr. Expt. Sta. 102 pp. 1954.
 168. _____, _____. Mechanical analysis of soils. *Engr. Rept. No. 21*, Iowa Engr. Expt. Sta; 60 pp. 1954.
 - _____, joint author. See under Chu, Handy, Lyon.
 - Davis, A.W., joint author. See under Higdon.
 - Davis, Arthur W. Editor, *Bulletin of the Mechanics Division--American Society for Engineering Education.*
 169. Davis, Norman T. Morphology of the female organs of reproduction in the *Miridae* (Hemiptera). *Ann. Ent. Soc. Amer.* 48:132-150. 1955.
 - Davis, R.J., joint author. See under Banks.
 - Dawn, E.E., joint author. See under Day.
 170. Day, A.D., E.E. Dawn, and K.J. Frey. Association between diastatic power and certain visible characteristics in barley. *Agron. Jour.* 47:163-165. 1955.
 171. Decker, Richard W., and Dean W. Stebbins. Photoelectric work functions of the borides of lanthanum, praseodymium, and neodymium. *Jour. Appl. Physics* 26:1004-1006. 1955.

172. Delouche, James C. and Louis N. Bass. Effect of light and darkness upon germination of seeds of *Agropyron smithii* L. Proc. Assoc. Offic. Seed Anal. 44:104-113. 1954.
173. Denisen, E.L. Weed control in small fruits. Iowa State Hort. Soc. Trans. 89:66-69. 1954.
_____, joint author. See under Lantz.
Dewell, E.H., joint author. See under Winders.
Diaz, F., joint author. See under Catron.
174. Dicke, F.F. The most important corn insects. Amer. Soc. Agron. 5:537-612. 1955.
175. _____ and L.H. Penny. Breeding for resistance to European corn borer. Proc. 9th Ann. Hybrid Corn Ind. Res. Conf. 9:44-53. 1955.
176. _____ and W.D. Guthrie. Advances in European corn borer resistance research. Proc. 10th Ann. Meet. North Central Branch, Ent. Soc. Amer. 10:12-13. 1955.
177. Dickerson, G.E., C.T. Blunn, A.B. Chapman, R.M. Kottman, J.L. Krider, E.J. Warwick, J.A. Whatley Jr., in collaboration with M.L. Baker, J.L. Lush, and I.M. Winters. Evaluation of selection in developing inbred lines of swine. North Central Reg. Publ. No.38. Univ. of Missouri Coll. Agric. Res. Bull. 551. 58 pp. 1954.
Dickinson, J.M., joint author. See under Wilhelm.
178. Diehl, H. and J.P. Butler. Ferricyanide titration of cobalt using ethylenediamine. Analyt. Chem. 27:777-785. 1955.
179. _____, and J.L. Ellingboe. Determination of amide nitrogen in vitamin B₁₂ and B_{12a}. Iowa State Coll. Jour. Sci. 27:421-423. 1953.
180. _____, and F.H. Lohman. 5,8-Dihydroxyquinoline and its nickel derivatives. Proc. Iowa Acad. Sci. 61:255-259. 1954.
_____, joint author. See under Brierly, Jaselskis, Murie.
_____, Advisory Board, Analytical Chemistry.
181. Diggins, Ronald V. and Clarence E. Bundy. Dairy Production. 1st ed. 342 pp. 1955.
Dinwiddie, Shirley W., joint author. See under Herrnstadt.
182. Dodd, John D. An approximation of the minimal tetrakaidocahedron. Amer. Jour. Bot. 42:566-569. 1955.
_____, joint author. See under Bragonier.
183. Dodds, J. Parry and K.R. Marvin. How do Iowa farmers obtain and use market news? Iowa Agric. Expt. Sta. Res. Bull. 417. 168 pp. 1954.
184. _____, _____. What does the Iowa farmer want from radio market news? Iowa Agric. Expt. Sta. Res. Bull. 413. 16 pp. 1954.
Dogger, J.R., joint author. See under Long, W.H.
185. Donovan, G.A., E.L. Johnson, S.L. Balloun, and R.E. Phillips. The long range effect of low level methionine supplementation in growing turkey rations. Poultry Sci. 34:251-256. 1955.
186. Dooley, M.E. and D.F. Atkins. A new electronic vacuum dilatometer. Rev. Sci. Instruments 26:568-571. 1955.
187. Douglas, Edna. The retail lumber establishment and farm dwelling construction in Iowa. Iowa Agric. Expt. Sta. Res. Bull. 415. 30 pp. 1954.
188. _____. An estimate of the volume of farm dwelling construction in Iowa. Iowa Agric. Expt. Sta. Res. Bull. 414. 25 pp. 1954.
_____, Editorial Staff, Journal of Marketing.
189. Dove, Richard C. Strain measurement errors in materials of low modulus. Proc. Amer. Soc. Civ. Engrs. Engr. Mechanics Div. 81:691-699. 1955.
190. Drake, Carl J. New and little-known Hemiptera-Heteroptera, Tingidae, from Ethiopia. Ann. and Mag. Nat. Hist. 7:657-665. 1954.
191. _____. Some tingids from the Belgian Congo (Hemiptera:Tingidae). Inst. Roy. Sci. Nat. Belgique Bull. 30:1-10. 1954.
192. _____. New genera and species of Tingidae from the old world (Hemiptera). Philippine Jour. Sci. 83:69-73. 1954.
193. _____. Tingidae: Descriptions and synonymic data (Hemiptera). Great Basin Nat. 14:1-10. 1954.
194. _____. An undescribed leptopodid from India (Hemiptera). Jour. Kansas Ent. Soc. 27:111-112. 1954.

195. _____. An undescribed *Metrobates* Uhler from Brazil (*Hemiptera*). So. Calif. Acad. Sci. Bull. 53:50-51. 1954.
196. _____. Synonymical data: Descriptions of new *Hydrometridae* (*Hemiptera*). Great Basin Nat. 14:61-66. 1954.
197. _____. Angolan *Tingidae* (*Hemiptera*). Camphanhia de Diamantes de Angola (Diamang) Servicos Culturais, Museu do Dundo. No.27, pp.85-90. 1955.
198. _____, and C.M. José. New water-striders from Brazil (*Hemiptera*). Proc. Biol. Soc. Washington 67:223-226. 1954.
199. _____, and Harold C. Chapman. New American waterstriders (*Hemiptera*). Florida Ent. 37:151-155. 1954.
200. _____, and Juan Gomez-Menor. A new genus of American *Ochteridae* (*Hemiptera*). Eos. Revista espanola de entomologia. Madrid. 30:157-159. 1954.
201. _____, _____. Some *Tingidae* from Spanish Guinea (*Hemiptera*). Eos. Revista espanola entomologia 30:89-93. 1954.
202. _____, _____. Notes on some American *Veliidae* (*Hemiptera*), with descriptions of two new *Microvelias* from Jamaica. Florida Ent. 37:133-138. 1954.
203. _____, and Tsing-chao Maa. Chinese and other oriental *Tingitoidea* (*Hemiptera*) III. Quart. Jour. Taiwan Museum 8:1-11. 1955.
204. _____, _____. The *Saldiodea* of China (*Hemiptera*). Quart. Jour. Taiwan Museum 7:263-266. 1954.
205. _____, and J. Maldonado-Capriles. Puerto Rican water-striders (*Hemiptera*). Proc. Biol. Soc. Washington 67:219-221. 1954.
206. _____, and Fritz Plaumann. Some *Microvelia* from southern Brazil (*Hemiptera: Veliidae*). Bull. So. Calif. Acad. Sci. 54:22-24. 1955.
207. _____, and Janis A. Roze. New Venezuelan *Gerridae* (*Hemiptera*). Proc. Biol. Soc. Washington 67:227-229. 1954.
208. Dudley, Pauline and Elizabeth Beveridge. Opportunities for teaching equipment in the homemaking class. Homemaking and Kelvinator Kitchen Reporter. April, 1955.
209. Duke, F.R. Qualitative theory of electron transfer mechanism. Rec. Chem. Prog. 15:55-59. 1954.
210. _____, and V.C. Bulgrin. The kinetics of the periodate oxidation of ethylene glycol and a series of methylated ethylene glycols. Jour. Amer. Chem. Soc. 76:3803-3806. 1954.
211. _____, and R.S. Laity. The measurement of transport numbers of pure fused salts. Jour. Phys. Chem. 59:549-552. 1955.
212. _____, _____. Transport numbers in pure fused salts. Jour. Amer. Chem. Soc. 76:4046. 1954.
213. _____, and F.R. Parchen. Kinetics of the reduction of ferric ion by hydroquinone in the presence of 1,10-phenanthroline. Jour. Amer. Chem. Soc. 77:3198-3199. 1955.
214. _____, and P.R. Quinney. The kinetics of the reduction of perchlorate ion by Ti(III) in dilute solution. Jour. Amer. Chem. Soc. 76:3800-3802. 1954.
215. Dumenil, Lloyd, H.R. Meldrum, and John Pesek. Best time to fertilize--fall or spring. Iowa Farm Sci. 9:3-6. 1954.
- _____, joint author. See under Hanway, Pesek.
216. Duncan, G.W., I. Casas, M.A. Emmerson, and R.M. Melampy. Estrogen in bovine blood, ovarian tissue, and follicular fluid. Fed. Proc. 14:40. 1955.
217. Dunleavy, John. Soybean diseases in Iowa in 1954. Plant Dis. Reporter 39:169-170. 1955.
218. _____. Control of damping-off of sugar beet by *Bacillus subtilis*. Phytopath. 45:252-258. 1955.
- Dunn, G.E., joint author. See under Gilman.
- DuToit, Schalk, joint author. See under Heady.
- Dwelle, Marion. Assistant Editor, Iowa Farm Science.
- Edminster, T.W., joint author. See under Frevert.
- Edwards, R.B., joint author. See under Beyer.
- Edwards, R.E., joint author. See under Banks.
- Egbert, Robert L., joint author. See under Hawkes.
219. Ehrenkranz, Florence and Mary S. Pickett. Energy consumption, temperatures, and palatability of foods stored in electric refrigerators with automatic defrosting. Jour. Home Econ. 47:185-188. 1955.

- Eileman, T.S., joint author. See under Grantham.
- Ellingboe, J.L., joint author. See under Brierly, Diehl.
- Emmerson, M.A., joint author. See under Duncan.
220. Engelhard, Arthur W. Occurrence of oak wilt fungus mats and pads on members of the red and white oak groups in Iowa. Plant Dis. Reporter 39:254-255. 1955.
221. Eppright, E.S. When does poor nutrition start? Iowa Farm Sci. 9:786-788. 1955.
222. _____. Dietary differences among school children. Natl. Livestock and Meat Board 26; 4 pp. 1955.
223. _____. Our responsibilities to the dietetic intern. Jour. Amer. Dietetic Assoc. 31:152-155. 1955.
224. _____. and C. Roderuck. Diet and nutritional status of Iowa school children. Amer. Jour. Publ. Health 45:464-471. 1955.
225. _____. _____, V.D. Sidwell, and P.P. Swanson. Relationships of estimated nutrient intakes of Iowa school children to physical and biochemical measurements. Jour. Nutrition 54:557-570. 1954.
226. _____. _____, and V.D. Sidwell. Physical measurements of Iowa school children. Jour. Nutrition 54:543-556. 1954.
227. _____. _____, and E. Jebe. Food intake and body size of Iowa children. A chapter in: Weight Control - A collection of papers presented at the Weight Control Colloquim. pp.80-96. Iowa State Coll. Press, Ames. 244 pp. 1955.
228. _____. _____, and P.P. Swanson. Nutritive value of the diets of Iowa school children. Jour. Nutrition 54:371-388. 1954.
229. _____. _____, and P.P. Swanson. Distribution of calories in diets of Iowa school children. Jour. Amer. Dietetic Assoc. 31:144-148. 1955.
230. _____. _____, Distribution of nutrients among meals and snacks of Iowa school children. Jour. Amer. Dietetic Assoc. 31:256-260. 1955.
- _____. Editorial Committee, the book "Weight Control" - A collection of papers presented at the Weight Control Colloquium.
231. Errington, Paul L. Review of "The Monkey Book" by Ernest P. Walker. Sci. Monthly 79:44. 1954.
232. _____. Review of "Principles of General Ecology", by Angus M. Woodbury. Quart. Rev. Biol. 30:57. 1955.
233. _____. Review of "Wildlife in Alaska", by A. Starker Leopold and F. Fraser Darling. Quart. Rev. Biol. 30:59. 1955.
234. _____. Review of "The Distribution and Abundance of Animals", by H.G. Andrewartha and L.C. Birch. Science 121:389-390. 1955.
235. _____. Our little wild dogs--the foxes. Audubon Mag. 57:14-17, 27. 1955.
236. _____. The special responsiveness of minks to epizootics in muskrat populations. Ecol. Monogr. 24:377-393. 1954.
- Esmay, D.L., joint author. See under Gilman.
237. Evans, D.D. and A.D. Scott. A polarographic method of measuring dissolved oxygen in saturated soil. Proc. Soil Sci. Soc. Amer. 19:12-16. 1955.
- _____. joint author. See under Schaller, Scott, A.D.
238. Everson, G., L. Northrop, N.Y. Chung, and R. Getty. Effect of ascorbic acid on rats deprived of pantothenic acid during pregnancy. Jour. Nutrition 54:305-312. 1954.
239. _____. _____, _____, and C. Pudalkewicz. Effect of varying the intake of calcium pantothenate of rats during pregnancy. I. Chemical findings in the young at birth. Jour. Nutrition 53:341-350. 1954.
- _____. joint author. See under Chung.
240. Everson, L.E. Are you sure it's good seed? Crops and Soils 7:16-17, 34. 1955.
241. _____. What kind of seed did you get? Iowa Farm Sci. 10:9-10. 1955.
242. _____. The germination of mature and immature seeds of quack grass (Agropyron repens). Proc. Assoc. Offic. Seed Analysts 44:127-128. 1954.
- Fahey, J.E., joint author. See under Cox.
243. Fahien, R.W. Mass transfer in packed beds. Amer. Inst. Chem. Engr. Jour. 1:28-37. 1955.
244. Fairchild, H.E. and Paul A. Dahm. A taxonomic study of adult chicken lice found in the United States. Jour. Kansas Ent. Soc. 27:106-111. 1954.

245. _____. Lice control on chickens with chlorinated hydrocarbon insecticides. *Jour. Econ. Ent.* 48:141-146. 1955.
246. Farnsworth, G.M. Jr., and A.W. Nordskog. Breeding for egg quality. 3. Genetic differences in shell characteristics and other egg quality factors. *Poultry Sci.* 34:16-26. 1955.
247. Fassel, V.A. and H.J. Hettel. Isotopic assay of lithium by means of hydride band emission spectra. *Spectrochimica Acta* 7:175-178. 1955.
248. _____, B. Quinney, L.C. Krotz and C.F. Lentz. Quantitative spectrographic analysis of the rare earth elements. *Analyt. Chem.* 27:1010-1014. 1955.
_____, United States Editor, *Spectrochimica Acta*.
_____, joint author. See under Brehm, Margoshes.
249. Fay, R.W. and Donald A. Lindquist. Laboratory studies on factors influencing the efficiency of insecticide impregnated cords for house fly control. *Jour. Econ. Ent.* 47:975-980. 1954.
_____, joint author. See under McCauley.
250. Fellingner, Robert C. Problems in Kinematics. 1st ed. 70 pp. Wm. C. Brown Co. 1955.
251. _____, H.A. Webber, and David Goldstein. Determination of the thermal conductivity of molten lithium. *Trans. Amer. Soc. Mech. Engr.* 77:97-102. 1955.
252. Fieller, E.C. and H.O. Hartley. Sampling with control variables. *Biometrika* 41:494-501. 1954.
253. Fink, H.C. *Prunus tomentosa* as an index plant for sour cherry viruses. *Phytopath.* 45:320-323. 1955.
254. _____ and W.F. Buchholtz. Correlation between sugar beet crop losses and greenhouse determinations of soil infestation by *Aphanomyces cochlioides*. *Proc. Amer. Soc. Sugar Beet Technol.* 8:252-259. 1954.
255. Finkner, R.E., H.C. Murphy, R.E. Atkins, II, and D.W. West. Seed fluorescence in oats. *Proc. Assoc. Offic. Seed Analysts* 44:202-205. 1954.
256. Fisher, R.W. and G.R. Winders. Lab for radiochemistry, metallurgy tests. *Nucleonics* 12:44. 1954.
257. Fitts, J.W., W.V. Bartholomew, and H. Heidel. Predicting nitrogen fertilizer needs of Iowa soils: I. Evaluation and control of factors in nitrate production and analysis. *Proc. Soil Sci. Soc. Amer.* 19:69-73. 1955.
Fitzsimmons, E.S., joint author. See under Nadler, Wilder.
Forbes, R.M., joint author. See under Albert.
258. Fortney, Cecil G. Jr., Merle P. Baker, and Emerson W. Bird. Cleaning stainless steel sanitary lines in-place. *Jour. Milk and Food Technol.* 18:150-156. 1955.
Foster, Joseph F., joint author. See under Thompson, Sarane.
259. Fowler, Eric B. and C.H. Werkman. Synthesis of amino acids by *Aerobacter aerogenes*. *Arch. Biochem. and Biophys.* 56:22-27. 1955.
260. _____, Laboratory Manual in General Bacteriology. 91 pp. 1955.
261. Fox, Sidney W. and Paul G. Homeyer. A statistical evaluation of the kinship of protein molecules. *Amer. Nat.* 89:163-168. 1955.
262. Frans, R.E. and E.F. Lind. On the kinetics of inhibition of growth by herbicides. *Proc. North Central Weed Control Conf.* 11:12-13. 1954.
_____, joint author. See under Loomis.
263. Frederickson, C.F. and J.H. Lilly. Measuring wireworm reactions to soil insecticides by tagging with radioactive cobalt. *Jour. Econ. Ent.* 48:438-442. 1955.
Freeland, M.Q., joint author. See under Fritz.
Freeman, James T., joint author. See under Campbell.
264. French, Dexter. Isolation and identification of planteose from tobacco seeds. *Jour. Amer. Chem. Soc.* 77:1024-1025. 1955.
265. _____. The raffinose family of oligosaccharides. *Advances in Carbohydrate Chem.* 9:149-184. 1954.
266. Frevert, R.K., G.O. Schwab, T.W. Edminster, and K.K. Barnes. Soil and Water Conservation Engineering. 1st ed. 479 pp. John Wiley and Sons. 1955.
_____, joint author. See under Schilfgaarde.
267. Frey, K.J. Agronomic mutations in oats induced by X-ray treatment. *Agron. Jour.* 47:207-209. 1955.

268. _____. The uses of F_2 lines in predicting the performance of F_3 selections in two barley crosses. *Agron. Jour.* 46:541-544. 1954.
269. _____, and T. Horner. Comparison of actual and predicted gains in barley selection experiments. *Agron. Jour.* 47:186-188. 1955.
_____, joint author. See under Browning, Day, Wiggans.
270. Fribourg, H.A. and I.J. Johnson. Response of soybean strains to 2,4-D and 2,4,5-T. *Agron. Jour.* 47:171-174. 1955.
271. _____, _____. Dry matter and nitrogen yields of legume tops and roots in the fall of the seeding year. *Agron. Jour.* 47:73-77. 1955.
_____, joint author. See under Johnson, I.J.
272. Fritz, J.S. Titration of bismuth with ethylenediaminetetraacetic acid. *Analyt. Chem.* 26:1978-1980. 1954.
273. _____. Acid-base titrations in nonaqueous solvents-selection of the medium for a particular titration. *Analyt. Chem.* 26:1701-1704. 1954.
274. _____. Organic quantitative analysis. *Jour. Chem. Educ.* 31:468-470. 1954.
275. _____ and M.Q. Freeland. Direct titrimetric determination of sulfate. *Analyt. Chem.* 26:1593-1595. 1954.
276. _____, and M.O. Fulda. Titrimetric determination of zirconium. *Analyt. Chem.* 26:1206-1208. 1954.
277. Fritz, Martin F. Q and L difference scores on the A.C.E. test. *Proc. Iowa Acad. Sci.* 61:356-357. 1954.
278. Frost, George. Membership in an Engineering Society. *Iowa Exponent* pp.24-25. Jan., 1955.
Fulda, M.O., joint author. See under Fritz.
Fung, Jon-Pong, joint author. See under Csanyi.
279. Funk, E.M., Agnes Frances Carlin, et al. Treating shell eggs to maintain quality. *North Central Regional Publ. No. 62. Missouri Agric. Expt. Sta. Bull.* 659. 19 pp. 1955.
280. Futrell, G.A. Ways to improve market news and information. *Natl. Proc. Agric. Communications Bull.* 24 pp. 1955.
281. Garcia, P., C. Roderuck, and P. Swanson. The relation of age to fat absorption in adult women together with observations on concentration of serum cholesterol. *Jour. Nutrition* 55:601-609. 1955.
282. Garner, William V. The future of immature insect systematics. *Proc. North Central Branch, Ent. Soc. Amer.* 10:81-82. 1955.
Garrigus, U.S., joint author. See under Albert.
283. Getty, Robert. Atlas for Applied Veterinary Anatomy. 1st ed. 227 pp. Burgess Publ. Co. 1955.
_____, joint author. See under Chung, Everson.
_____, Editor, *Anatomy and Histology.*
_____, Editor, *American Journal of Veterinary Research.*
Ghostley, F.J., joint author. See under Nordskog.
284. Giese, Henry. A rational approach to the control of farm fires. *Proc. 74th Ann. Conven. Iowa Assocn. Mutual Ins. Associations.* pp.34-44. 1954.
285. _____. Prevent Wind Damage (rev.). *Iowa Mutual Tornado Ins. Assoc.* 24 pp. 1954.
286. Gilman, Henry and Souren Avakian. Some aromatic and heterocyclic dialkyl arsines. *Jour. Amer. Chem. Soc.* 76:4031. 1954.
287. _____, _____, R.A. Benkesser, H.S. Broadbent, R.M. Clar, G. Karmas, F.J. Marshall, S.M. Massie, D.A. Shirley, and L.A. Woods. N-methyl-3-isopropyl-4-dimethyl-aminophenyl carbamate and some related derivatives. *Jour. Organ. Chem.* 19:L067-1079. 1954.
288. _____, T.H. Cook, J.A. Hogg, J. Swiss, and R.G. Johnson. Some 1,2-, 2,3-, and 3,4-disubstituted dibenzofurans. *Jour. Amer. Chem. Soc.* 76:5783-5785. 1954.
289. _____, G.E. Dunn, Howard Hartzfeld, and A.G. Smith. The piperidine-catalyzed reaction of triphenylsilane with some hydroxy compounds. *Jour. Amer. Chem. Soc.* 77:1287-1288. 1955.
290. _____, and J. Eisch. Preparation and stability of some organotin and organo-lead compounds. *Jour. Organ. Chem.* 20:763-769. 1955.
291. _____, and D.L. Esmay. Hydroxy derivatives of dibenzofuran, phenoxathiin and thianthrene. *Jour. Amer. Chem. Soc.* 76:5787. 1954.

292. _____, _____. Some derivatives of dibenzothiophene. Jour. Amer. Chem. Soc. 76:5786. 1954.
293. _____, and Richard D. Gorsich. The direct preparation of benzyllithium. Jour. Amer. Chem. Soc. 77:3134. 1955.
294. _____, and Robert K. Ingham. Some tetraorganosilanes. Jour. Amer. Chem. Soc. 77:1680. 1955.
295. _____, and Katashi Oita. Cleavage of some organosilicon compounds by formic acid. Jour. Amer. Chem. Soc. 77:3386. 1955.
296. _____, _____. Some 2-biphenyl substituted organosilicon compounds. Jour. Organ. Chem. 20:862-870. 1955.
297. _____, and Dhairyasheel R. Swayampati. Conversion of thianthrene-5-oxide to dibenzothiophane by *n*-butyllithium. Jour. Amer. Chem. Soc. 77:3387. 1955.
_____, joint author. See under Benkeser, Meen, Merten, Oita.
_____, Board of Editors, Journal of Organic Chemistry.
_____, Board of Editors, Organic Syntheses.
298. Gilman, J.C., joint author. See under Bragonier, Hanson, Ralph, Tiffany.
298. Gleason, L.S. and W.E. Loomis. Basal sprays and injection treatments on trees. Proc. North Central Weed Control Conf. 11:102-104. 1954.
_____, joint author. See under Chapman.
299. Goetz, Charles A. and John P. Butler. Chemistry + TV = Challenge. Chem. and Engr. News 32:3368-3370. 1954.
300. Goheen, Harry. The Wedderburn theorem. Canadian Jour. Math. 7:60-62. 1955.
301. _____, Review of "Principles of Numerical Analysis" by Alston S. Householder. Econometrica 22:540-541. 1954.
Goldstein, David, joint author. See under Fellinger.
Gomez-Menor, Juan, joint author. See under Drake.
302. Goodman, L. and H. Shull. Molecular calculations. III. A modification of the naive semi-empirical MO method. Jour. Chem. Physics 23:33-43. 1955.
303. Goring, C.A.I. Biological transformations of phosphorus in soil. I. Theory and methods. Plant and Soil 6:17-25. 1955.
304. _____, Biological transformations of phosphorus in soil. II. Factors affecting synthesis of organic phosphorus. Plant and Soil 6:26-37. 1955.
_____, joint author. See under Mehta.
Gorsich, Richard D., joint author. See under Gilman, Henry.
Gould, Marvin. Managing Editor, Iowa Custodians Newsletter.
305. Gowen, John W. Significance and utilization of animal individuality in disease research. Jour. Natl. Cancer Inst. 15:555-570. 1954.
306. _____, and Janice Stadler. Influence of genotype on weight control in mice. Iowa State Coll. Jour. Sci. 30:49-63. 1954.
_____, joint author. See under Thompson, Sarane.
_____, Editor, Biometric Section, Biological Abstracts.
_____, Editorial Board, Genetics.
307. Grabe, Don F. and Louis N. Bass. Comparative methods of germination of smooth bromegrass. Proc. Assoc. Offic. Seed Analysts 44:121-126. 1954.
308. Graebner, Norman A. Politics in foreign policy. Curr. Hist. 28:7-14. 1955.
309. _____, The farm issue. Curr. Hist. 27:249-255. 1954.
_____, Associate Editor, World Affairs Quarterly.
310. Graf, G.E. and S. Aronoff. Carbon dioxide fixation by roots. Science 121:211-212. 1955.
Grainger, Mary M., joint author. See under McCauley.
Gram, M.R., joint author. See under Swanson, P.P.
311. Grantham, L.F., T.S. Elleman, and D.S. Martin, Jr. Exchange of chlorine in aqueous systems containing chloride and tetrachloropentate (II). Jour. Amer. Chem. Soc. 77:2965-2971. 1955.
Graybill, Bruce M., joint author. See under Hawthorne.
312. Gregory, G. A method for rapidly transferring a substance on paper to the origin of a chromatogram. Science 121:169. 1955.
313. Griffing, Bruce and E.W. Lindstrom. A study of the combining abilities of corn inbreds having varying proportions of corn belt and non-corn belt germ plasm. Agron. Jour. 46:545-552. 1954.
314. Gross, W.A. Timely tips - dimensional algebra. Jour. Engr. Educ. 45:497. 1954.

315. Groth, A.H. Jr., and J.R. Rosdail. Cystic bile ducts and renal tubules in baby pigs. *Jour. Amer. Vet. Med. Assoc.* 126:133-134. 1955.
Grundemeier, Winston, joint author. *See under Hammond.*
316. Gunderson, Harold. Insects? Chemicals ban them. *Better Farming Methods* p.20. January, 1955.
_____, joint author. *See under Hansen, R.W., Lilly, Sylwester.*
317. Gurland, John. On regularity conditions for maximum likelihood estimators. *Skandinavisk Aktuarietidskrift. Haft 1-2*:71-76. 1954.
318. _____. Distribution of definite and of indefinite quadratic forms. *Ann. Math. Statistics* 26:122-127. 1955.
319. _____. Distribution of the maximum of the arithmetic mean of correlated random variables. *Ann. Math. Statistics* 26:294-300. 1955.
Guthrie, W.D., joint author. *See under Dicke.*
320. Haber, E.S. Hybridization in related fields. *Proc. 9th Ann. Hybrid Corn Ind. Res. Conf. Publ. No.9.* pp.80-84. December, 1954.
321. Hale, W.H. and R.P. King. Possible mechanism of urea toxicity in ruminants. *Proc. Soc. Exptl. Biol. and Med.* 89:112-114. 1955.
_____, joint author. *See under Albert, Repp.*
Hamilton, Chester E., joint author. *See under Boozer, Hammond, George S.*
322. Hamilton, Howard L. Relations between alkaline phosphatase, beryllium, and morphogenesis of the feather. *Anat. Rec.* 122:418-419. 1955.
323. _____. and Gertrude W. Hinsch. The fate of the second visceral pouch in the chick. *Anat. Rec.* 120:762. 1954.
324. _____. and Alice L. Koning. Changes in feather structure produced by a phosphatase-inhibitor. *Anat. Rec.* 120:710-711. 1954.
325. _____. Topographical relation of alkaline phosphatase, ribonucleic acid, and mucopolysaccharides to the morphogenesis of the down feather. *Anat. Rec.* 117:522. 1953.
_____, joint author. *See under Hinsch, Koning.*
326. Hammer, C.L., R.W. Pidd, K.M. Terwilliger. Betättron oscillations in the synchrotron. *Rev. Sci. Instruments.* 26:555-556. 1955.
_____, joint author. *See under Pidd.*
327. Hammond, Earl G. and W.O. Lundberg. Molar refraction, molar volume, and refractive index of fatty acid esters and related compounds in the liquid state. *Jour. Amer. Oil Chemists' Soc.* 31:427-432. 1954.
328. Hammond, George S. A correlation of reaction rates. *Jour. Amer. Chem. Soc.* 77:334-338. 1955.
329. _____. Charles E. Boozer, Chester E. Hamilton, and Jyotirindra N. Sen. Air oxidation of hydrocarbons. III. Mechanism of inhibitor action in benzene and chlorobenzene solutions. *Jour. Amer. Chem. Soc.* 77:3238-3244. 1955.
330. _____. and Winston Grundemeier. The mechanism of the benzidine rearrangement. III. The rearrangement of p,p'-didenterohydrazobenzene. *Jour. Amer. Chem. Soc.* 77:2444-2446. 1955.
331. _____. and Donald H. Hogle. The dissociation of sterically hindered acids. *Jour. Amer. Chem. Soc.* 77:338-340. 1955.
332. _____. and George B. Lucas. The reaction of chloranil with the decomposition products from azonitriles. *Jour. Amer. Chem. Soc.* 77:3249-3251. 1955.
333. _____. and Lawrence R. Parks. The displacement of halogen from 1-x-2,4-dinitrobenzenes. *Jour. Amer. Chem. Soc.* 77:340-343. 1955.
334. _____. Jyotirindra N. Sen, and Charles E. Boozer. The efficiency of radical production from azo-bis-isobutyronitrile. *Jour. Amer. Chem. Soc.* 77:3244-3248. 1955.
_____, joint author. *See under Boozer, Hawthorne, Parks.*
335. Hammond, L.C., W.H. Allaway, and W.E. Loomis. Effects of oxygen and carbondioxide levels upon absorption of potassium by plant. *Plant Physiol.* 30:155-161. 1955.
336. Handy, R.L., D.T. Davidson, and T.Y. Chu. Effect of petrographic variations of southwestern Iowa loess on stabilization with Portland cement. *Bull. 98, Highway Res. Bd. "Stabilization of Soils". Natl. Acad. Sci.-Natl. Res. Coun. Publ.* 346. pp. 1-20. 1954.
337. _____. C.A. Lyon, and D.T. Davidson. Analyses of wind-blown silt--March, 1954. *Proc. Iowa Acad. Sci.* 61:278-290. 1954.

- Handy, R.L., joint author. See under Davidson, Lyon.
- Hansen, J.H., joint author. See under Wenkert.
- Hansen, Norman J., joint author. See under McComb.
338. Hansen, R.S. and B.H. Clampitt. The apparent adsorption of some aliphatic compounds from aqueous solutions as inferred from Hydrogen overvoltage measurements. *Jour. Physical Chem.* 59:496-498. 1955.
339. _____ and R.D. Hansen. The adsorption of hydrocarbons for methanol and ethanol solutions by non-porous carbons. *Jour. Physical Chem.* 59:496-498. 1955.
340. _____, F.A. Miller, and S.D. Christian. Activity of coefficients of components in the systems water-acetic acid, water-propionic acid and water-*n*-butyric acid at 25°C. *Jour. Physical Chem.* 59:391-395. 1955.
341. Hansen, Ralph W., Harold Gunderson, J.C. Gilman, and Richard Phillips. Rat race in the grain bin. *Co-op. Grain Quart.* 12:67-72. 1954.
- _____, joint author. See under Phillips, R.
342. Hanway, John and Lloyd Dumenil. Predicting nitrogen fertilizer needs of Iowa soils: III. Use of nitrate production together with other information as a basis for making nitrogen fertilizer recommendations for corn in Iowa. *Soil Sci. Soc. Amer., Proc.* 19:77-80. 1955.
343. _____ and George Stanford. Now - A satisfactory nitrogen test. *What's New in Crops and Soils* 6:16-17. 1954.
- _____, joint author. See under Stanford.
- Harrington, David N., joint author. See under Dachtler.
- Harrington, W.F., joint author. See under Anfinssen.
344. Harris, H.M. Inspection and quarantine problems in the past years. *Trans. State Hort. Soc.* 1954. 89:188-190. 1955.
345. _____. Orchard mites. *Trans. State Hort. Soc.* - 1954. 89:75-78. 1955.
346. _____. Nursery inspection as it applies to the fruit grower. *Trans. State Hort. Soc.* - 1954. 89:78-79. 1955.
- _____, Editorial Committee, *Annual Review of Entomology.*
- Harrison, Gladys L., joint author. See under Rutledge.
347. Hartley, H.O. Some recent developments in analysis of variance. *Communications on Pure and Appl. Math.* 8:47-72. 1955.
348. _____ and Alan Ross. Unbiased ratio estimators. *Nature (London)* 174:270-271. 1954.
- _____, joint author. See under David, Fieller, Kozicky.
- _____, Editorial Panel, *Journal of the Royal Statistical Society, Series B.*
- _____, Editorial Panel, *Biometrika.*
- Hartzfeld, Howard, joint author. See under Gilman, H.
- Houghton, Kenneth E., joint author. See under Almfeldt.
- Haver, C.B., joint author. See under Heady.
349. Hawk, V.B., H.D. Hughes, and F.W. Schaller. Reed canarygrass heals gullies. *Jour. Soil and Water Conserv.* 9:203-209, 237. 1954.
350. Hawkes, Glenn R. and Robert L. Agbert. Personal values and the emphatic response: their interrelationships. *Jour. Educ. Psychol.* 45:469-476. 1954.
351. Hawthorne, M. Frederick and George S. Hammond. The oxygenation of the tri-(*p*-nitrophenyl)-methide ion. A novel oxidation-reduction reaction. *Jour. Amer. Chem. Soc.* 77:2549-2550. 1955.
352. _____, _____, and Bruce M. Graybill. The nucleophilicity of the cyanide ion. *Jour. Amer. Chem. Soc.* 77:486-487. 1955.
- _____, joint author. See under Parks, Laurence R.
353. Hazel, L.N. Hogs to fit our needs. *Iowa Farm Sci.* 9:467. 1954.
354. _____, and L.E. Johnson. Gene - the universal calf. *Aberdeen-Angus Jour.* 36:(3)-(12). (a continuing series) 1955.
- _____, joint author. See under Kline, E.A.
- _____, Editorial Board, *Journal of Animal Science.*
355. Heath, Harry E. Jr. (Editor) *Directory of Journalism Films.* 2nd ed. 95 pp. Iowa State College Press. 1954.
356. Heady, Earl O. Supply of U.S. farm products under conditions of full employment. *Amer. Econ. Rev.* 45:228-238. 1955.
357. _____. Simplified presentation and logical aspects of linear programming. *Jour. Farm Econ.* 36:1035-1048. 1954.

358. _____. Farm production economics research. In: *Contemporary Readings in Agricultural Economics*, edited by H.G. Halcrow. 1st ed. 411 pp. Prentice-Hall. 1955.
359. _____ and C.B. Baker. Resource adjustments to equate productivities in agriculture. *Southern Econ. Jour.* 21:36-52. 1954.
360. _____ and Schalk DuToit. Marginal resource productivity for agriculture in selected areas of South Africa and the United States. *Jour. Political Econ.* 62:494-505. 1954.
361. _____ and Dean McKee. What crop diversification can do. *Iowa Farm Sci.* 9:3-6. 1954.
362. _____, _____, and C.B. Haver. Farm size adjustments in Iowa and cost economies in crop production. *Iowa Agric. Expt. Sta. Bull.* 428. 24 pp. 1955.
363. _____, Russell O. Olson, and J.M. Scholl. Economic efficiency in pasture production and improvement in southern Iowa. *Iowa Agric. Expt. Sta. Res. Bull.* 419. 25 pp. 1954.
364. _____ and John Pesek. Hutton and Thorne on isoclines. *Jour. Farm Econ.* 37:363-368. 1955.
365. _____, _____. A fertilizer production surface with specification of economic optima for corn grown on calcareous Ida silt loam. *Jour. Farm Econ.* 36:466-482. 1954.
366. _____, _____, and Wm. G. Brown. Crop response surfaces and economic optima in fertilizer use. *Iowa Agric. Expt. Sta. Res. Bull.* 424. 1955.
367. _____ and James Seagraves. How do Iowa farm incomes stack up? *Iowa Farm Sci.* 9:582-585. 1954.
368. _____ and Russel Shaw. Resource returns and productivity coefficients in selected farming areas of Iowa, Alabama, and Montana. *Iowa Agric. Expt. Sta. Res. Bull.* 425. 1955.
369. _____ and E.G. Strand. Efficiency within American agriculture. *Jour. Farm Econ.* 37:524-537. 1955.
- _____, joint author. See under Babbar, Baumann, Bowlen, Brown, Jensen, A.H., Jensen, H.R., Kehrberg, McKee, Peterson, G.A.
- _____, Editorial Council, *Journal of Farm Economics*.
- Heer, John F., joint author. See under Rutledge.
- _____, Assistant Publications Editor, *Iowa Agric. Expt. Sta. Res. Publications*.
- _____, Editor, *Iowa Farm Science*.
- Heidel, H., joint author. See under Fitts.
- Hendrickson, George O., joint author. See under Kline, P.D., Kozicky, Tanner.
- _____, Abstractor, *Biological Abstracts*.
370. Herman, Robert D. The 'going steady' complex: A re-examination. *Marriage and Family Living* 17:36-40. 1955.
371. Herrstadt, Richard L., and Shirley W. Dinwiddie. Amos Bronson Alcott: A bibliography--Part I. *Bull. Biblio.* 21:64-67. 1954.
372. _____, _____. Amos Bronson Alcott: A bibliography--Part II. *Bull. Biblio.* 21:92-96. 1954.
373. Herwig, L.O. and G.H. Miller. Relative ionization yields for fission fragments in various gases. *Physical Rev.* 95:413-417. 1954.
374. Heslep, J.M., and C.A. Black. Diffusion of fertilizer phosphorus in soils. *Soil Sci.* 78:389-401. 1954.
- Hettel, H.J., joint author. See under Fassel.
375. Hibbs, E.T. The value and use of European corn borer surveys. *Proc. 8th Ann. Meet. North Central Branch Ent. Soc. Amer.* 8:25-27. 1953.
376. _____. Cereal crop insects: present and future look. *Proc. 10th Ann. Meet. North Central Branch Ent. Soc. Amer.* 10:8-9. 1955.
377. _____. Effect of late application of chemicals to field corn. *Proc. 10th Ann. Meet. North Central Branch Ent. Soc. Amer.* 10:10. 1955.
378. _____. Latest developments in sweet corn insect control. *Ohio Vegetable and Potato Growers Assoc., Ann. Proc.* 40:63-68. 1955.
379. Higdon, Archie, Wm. B. Stiles, A.W. Davis, and H.O. Ustrud. *Engineering Mechanics*. 2nd ed. 585 pp. Prentice-Hall. 1955.
- Hillyard, L.R., joint author. See under Dana.
380. Hinsch, Gertrude W. and Howard L. Hamilton. The development fate of the first somite of the chick. *Anat. Rec.* 122:448-449. 1955.
- _____, joint author. See under Hamilton, H.L.

381. Hippaka, Thomas A. What it means to be a Finnish-American. *Kalevainen* 43:6-7. 1955.
382. _____. Some administrator soul searching. *Indust. Arts and Voc. Educ.* 44:179-183. 1955.
383. _____. Above and beyond the call of earning a living. *School Shop* 15:7-44. 1955.
384. Hoffman, P.F. Physiology of *Endoconidiophora fagacearum* Bretz. I. Factors influencing growth and toxin production. *Iowa State College Jour. Sci.* 29:27-38. 1954.
385. Hofstad, M.S. The secondary immune response in chickens revaccinated with inactivated Newcastle disease virus vaccine. *Amer. Jour. Vet. Res.* 15:604-606. 1954.
- Hogg, J.A., joint author. See under Gilman, Henry
- Hogle, Donald H., joint author. See under Hammond, George S.
386. Hogrefe, Pearl. John More's translations. *Papers of the Bibliogr. Soc. Amer.* 49:188-189. 1955.
387. Hollander, W.F. Pigeons in research. *Animal Care Panel, 5th Ann. Meet.* pp.71-80. 1954.
388. _____. Reveiw of the principals of genetics (sic). *Jour. Heredity* 46:123-124.
389. Hollen, Norma and Jane Saddler. *Textiles*. 1st ed. 197 pp. Macmillan. 1955.
- _____, joint author. See under Roberson.
390. Holt, Imy Vincent. Cytohistological responses of varieties of *Avena* to 2,4-D. *Iowa State Coll. Jour. Sci.* 29:581-629. 1955.
- Homeyer, P.G., joint author. See under Allen, Carlin, Fox, Jacobson, Jensen, A.H., Kozicky.
391. Hooker, A.L., G.F. Sprague, and W.A. Russell. Resistance to *Puccinia sorghi* in corn. *Agron. Jour.* 47:388. 1955.
- Hooker, D.T., joint author. See under Klingman.
392. Horner, T.W. and O. Kempthorne. The components of variance and the correlations between relatives in symmetrical random mating populations. *Genetics* 40:310-320. 1955.
- _____, joint author. See under Frey.
393. Horrocks, D.L. and A.F. Voigt. Half-life of thallium²⁰⁴. *Physical Rev.* 95:1205. 1954.
- Howard, N., joint author. See under Beegle.
394. Hoyt, Elizabeth E. The spender's choice. *Corona (London)* 6:332-335. 1954.
- Hudson, D.E., joint author. See under Shaw, W.C.
395. Huffman, M.D. Testing for resistance to the *Septoria* disease of oats. *Plant Dis. Reporter* 39:25-28. 1955.
396. Huffman, Marion D. Disease cycle of *Septoria* disease of oats. *Phytopath.* 45:278-280. 1955.
397. Hughes, F.J. and D.S. Martin, Jr. Kinetics of the oxidation of selenious acid by hydrogen peroxide. *Jour. Phys. Chem.* 59:410-415. 1955.
- _____, joint author. See under Martin, D.S.
- Hughes, H.D., joint author. See under Hawk, Scholl.
398. Hughes, Wm.L. A technique for solving certain networks containing a nonlinear element. *Proc. Natl. Electronics Conf.* 10:337-348. 1954.
399. _____. Experimental equipment for recording and reproducing color television images on black and white film. 1955 I.R.E. Convention Record, Part 7. pp.69-80. 1955.
400. Hukill, W.V. and N.C. Ives. Radial air flow resistance of grain. *Agric. Engr.* 36:332-335. 1955.
401. _____. and C.K. Shedd. Non-linear air flow in grain drying. *Agric. Engr.* 36:462-466. 1955.
- Hull, Dale O., joint author. See under Armstrong.
402. Hunt, Donnell. *Farm Power and Machinery Management. A laboratory manual* 137 pp. 1954.
403. _____. *Farm Tractor Power. A laboratory manual.* 44 pp. 1955.
404. Huntress, Keith. Atomic Institute. (a poem) *Saturday Rev.* p.29, June 4, 1955.
405. Hurlburt, Virgil L. Farm rental practices and problems in the midwest. *North Central Reg. Publ. No. 50. Iowa Agric. Expt. Sta. Res. Bull.* 416. 1954.
- Hussey, Roland F., joint author. See under Drake.

406. Hutchcroft, Charles D. and Jos. L. Robinson. Iowa corn yield test - 1954. Iowa Agric. Expt. Sta. Bull. P118. 36 pp. 1955.
_____, joint author. See under Wengert.
Hviot, A., joint author. See under Anfinssen.

Ingham, Robert K., joint author. See under Gilman.
407. Isaacson, Stanley. Review of "Design for Decision," by Irwin D. J. Bross. *Econometrica* 23:224-225. 1955.
408. Isely, Duane. Keys to sweet clover (Melilotus). Iowa Acad. Sci. Proc. 61:119-131. 1954.
Ives, N. C., joint author. See under Hukill.

Jacobs, E. M., joint author. See under Clifton, Davey.
Jacobsen, F. M., joint author. See under Beyer.
409. Jacobson, N. L., R. S. Allen, J. T. Blake, and P. G. Homeyer. The effect of method of administration on the absorption and storage of vitamin A by dairy calves. *Jour. Nutr.* 54:143-153. 1954.
_____, joint author. See under Allen, Lambert, Picken, Voelker.
410. Jaselskis, B., and Harvey Diehl. The polarography of vitamins B_{12r} and B_{12a}. *Jour. Amer. Chem. Soc.* 76:4345-4349. 1954.
411. Jebe, Emil H., and Norman V. Strand. Livestock marketing practices in Iowa. *Agric. Econ. Res.* 6:84-92. 1954.
_____, joint author. See under Altman, Eppright.
412. Jensen, A. H., D. C. Acker, H. M. Maddock, G. C. Ashton, P. G. Homeyer, E. O. Heady, and D. V. Catron. Different protein levels with and without antibiotics for growing-finishing swine: Effect on growth rate and feed efficiency ^{1,2,3}. *Jour. Animal Sci.* 14:69-81. 1955.
_____, joint author. See under Ashton.
Jensen, E. N., joint author. See under Jones, J. T., Nichols, Pohm.
413. Jensen, Harold R., Earl O. Heady, and Ross V. Baumann. Costs, Returns, and Capital Requirements for soil conserving farming on rented farms in western Iowa. Iowa Agr. Exp. Sta. Res. Bull. 423. 1955.
414. Jessen, Raymond J. Determining the fruit count on a tree by randomized branch sampling. *Biometrics* 11:99-109. 1955.
415. _____. Agricultural sample survey of the province of Pichincha, Ecuador, 1952. *Estadistica* 12:401-421. 1954.
_____, joint author. See under Clifton.
416. Johansson, Sven A. E. The decay of UX₁, UX₂, and UZ. *Physical Rev.* 96:1075-1080. 1954.
417. _____. Experiments on the direct photonuclear effect. *Physical Rev.* 97:434-443. 1955.
Johnson, E. L., joint author. See under Donovan.
418. Johnson, I. J., Effect of 2,4-D treatment of oats on the succeeding crop. *Agron. Jour.* 46:475-476. 1954.
419. _____. W. V. Bartholomew, and H. A. Fribourg. More legumes for green manure = more corn. Iowa Farm Sci. 9:10-12. 1954.
420. _____. and W. H. Bragonier. Your recommended crop varieties for 1955. Iowa Farm Sci. 9:9-12. 1955.
_____, joint author. See under Fribourg.
_____, Associate Editor, *Agronomy Journal*.
- Johnson, L. E., joint author. See under Hazel.
Johnson, R. G., joint author. See under Gilman, Oita.
Johnston, W. B., joint author. See under Larson.
421. Jones, Jack Colvard and Oscar E. Tauber. Abnormal hemocytes in mealworms (Tenebrio molitor L.). *Ann. Ent. Soc. Amer.* 47:428-444. 1954.
422. Jones, J. T. Jr., and E. N. Jensen. Radiation from cerium¹⁴¹. *Physical Rev.* 97:1031-1033. 1955.
423. Jones, W. Paul. Editor: *Essays on Thinking and Writing*. 2nd ed. 87 pp. Wm. C. Brown Co., Dubuque, Iowa. 1955.
Judge, George, joint author. See under Nordin.
Juhl, W. G., joint author. See under Arnold,

424. Kaldor, Donald R. Dumping CCC stock abroad. *Farm Policy Forum* 7:18-23. 1955.
425. _____. Moving resources out of agriculture. *Farm Policy Forum* 7:31-36. 1954.
426. Kalton, R.R., R.C. Leffel, C.E. Wassom, and M.E. Weiss. Evaluation of combining ability in *Dactylis glomerata* L. I. Clonal and outcross progeny performance. *Iowa State Coll. Jour. Sci.* 29:631-657. 1955.
_____, joint author. See under Lebsack, Leffel.
Karmas, G., joint author. See under Gilman.
Kastelic, J., joint author. See under Ashton.
427. Kehr, August E. Turnor induction on nicotiana species by use of coconut milk and yeast extract. *Science* 121:869-870. 1955.
428. _____, Y.C. Ting, and J.R. Miller. The site of carotenoid and anthocyanin synthesis in sweet potatoes. *Amer. Soc. Hort. Sci.* 65:396-398. 1955.
429. Kehrberg, Earl W., Earl O. Heady, and Dean McKee. Is crop diversification the answer? *Iowa Farm Sci.* 9:3-5. 1954.
430. Kempthorne, O. The correlation between relatives in a random mating population. *Proc. Roy. Soc. London, Ser. B.* 143:103-113. 1954.
431. _____. The theoretical values of correlations between relatives in random mating populations. *Genetics* 40:153-167. 1955.
432. _____. The correlation between relatives in a simple autotetraploid population. *Genetics* 40:168-174. 1955.
_____, joint author. See under Anderson, Black, Horner, Zoellner.
Kenkel, William F., joint author. See under Cuber.
King, R.P., joint author. See under Hale.
433. King, W. Bernard. *Semimicro Experiments in General Chemistry*. 2nd ed. 165 pp. Prentice-Hall. 1955.
434. _____. and John A. Beel. The Grignard reagent reaches the freshman. *Jour. Chem. Educ.* 32:146-148. 1955.
435. Kirk, L.K. and J.P. Mahlstede. Packaging dormant nursery stock for shipment to retail markets. *Proc. Amer. Soc. Hort. Sci.* 65:472-482. 1955.
_____, joint author. See under Mahlstede.
436. Kirkham, Don. Measurement of the hydraulic conductivity of soil in place. *Amer. Soc. Testing Materials Spec. Tech. Publ. No. 163. "Symposium on Permeability of Soils."* pp.80-97. 1955.
437. _____. Seepage of artesian and surface water into drain tubes in stratified soil. *Trans. Amer. Geophys. Union* 35:775-790. 1954.
438. _____. and W.V. Bartholomew. Equations for following nutrient transformations in soil, utilizing tracer data: II. *Soil Sci. Soc. Amer. Proc.* 19:189-192. 1955.
_____, joint author. See under Rollins, Schilfgaarde.
_____, Consulting Editor, *Soil Science*.
439. Kline, E.A. and L.N. Hazel. Loin area at tenth and last rib as related to leanness of pork carcasses. *Jour. Animal Sci.* 14:659. 1955.
_____, joint author. See under Ashton.
440. Kline, Paul D. and George O. Hendrickson. Autumnal decimation of Mearns cottontail, Decatur County, Iowa, 1952. *Proc. Iowa Acad. Sci.* 61:524-527. 1954.
441. Kline, R.M. and D.J. Zaffarano. Decay characteristics of some short-lived nuclides of low atomic number. *Physical Rev.* 96:1620. 1954.
442. Klingman, D.W., D.T. Hooker, and C.V. Banks. Photometric titration assembly for Beckman model DU spectrophotometer. *Analyt. Chem.* 27:572. 1955.
443. Klonglan, Eugene D. Pheasant production on the Winnebago Research Area, 1954. *Iowa Conservationist* 11:1. 1954.
444. _____. Factors influencing the fall roadside pheasant census in Iowa. *Jour. Wildlife Management* 19:254-262. 1955.
445. Koning, Alice L. Histochemical localization of some of the constituents of the developing juvenile feather. *Anat. Rec.* 120:753-754. 1954.
446. _____. and Howard L. Hamilton. Changes in the localization of alkaline phosphatase ribonucleic acid, and polysaccharides during the transition from down to juvenile stages of the feather. *Anat. Rec.* 122:422-423. 1955.
_____, joint author. See under Hamilton.
- Kottman, Roy M., joint author. See under Beneke, Dickerson.

447. Kozicky, E.L., H.O. Hartley, and G.O. Hendrickson. A proposed comparison of fall roadside pheasant counts and flushing rates. *Proc. Iowa Acad. Sci.* 61:528-534. 1954.
448. _____, George O. Hendrickson, Paul G. Homeyer, and Richard Nomsen. Weather and fall pheasant populations in Iowa. *Jour. Wildlife Management* 19:136-142. 1955.
- Krider, J.L., joint author. See under Dickerson.
- Krotz, L.C., joint author. See under Fassel.
- Krusekopf, H.H., joint author. See under Muckenhirn.
- Kuetemeyer, Carol A., Assistant Editor, *Iowa Farm Science.*
- Kutish, Francis A., Editor, *Farm Policy Forum.*
- Laity, R.W., joint author. See under Duke.
449. Lambert, M.R., N.L. Jacobson, R.S. Allen, and M.R. Bell. The relation of growth, feed consumption, and certain blood constituents to changes in the dietary of young dairy calves. *Jour. Dairy Sci.* 38:6-12. 1955.
450. Lana, E.P. and Lewis E. Peterson. Irrigation in Iowa. *Market Growers Jour.* 48:13-30. 1954.
- _____, joint author. See under Peterson, L.E.
451. Lantz, H.L. and E.L. Denisen. Black Hawk, a new raspberry for the midwest. *Iowa Farm Sci.* 9:639-641. 1954.
452. Larson, W.E. Effect of method of application of double superphosphate on the yield and phosphorus uptake by sugar beets. *Amer. Soc. Sugar Beet Technol. Proc.* 8:25-31. 1954.
453. _____ and W.B. Johnston. The effect of soil moisture level on the yield, consumptive use of water, and root development by sugar beets. *Soil Sci. Amer. Proc.* 19:275-279. 1955.
- _____, joint author. See under Schaller.
454. Lawton, W.C., and F.E. Nelson. Influence of sublethal treatment with heat or chlorine on the growth of psychrophilic bacteria. *Jour. Dairy Sci.* 38:380-387. 1955.
455. _____, _____. The effect of storage temperatures on the growth of psychrophilic organisms in sterile and laboratory pasteurized skim milk. *Jour. Dairy Sci.* 37:1164-1172. 1954.
456. LeBaron, Helen R. Career day planning. *Jour. Home Econ.* 47:91-94. 1955.
457. Lebsock, K.L. and R.R. Kalton. Variation and its evaluation within and among strains of *Bromus inermis* Leyss. I. Spaced-plant studies. *Agron. Jour.* 46:463-467. 1954.
458. Leffel, R.G., R.R. Kalton, and C.E. Wassom. Fertility evaluations and their relationships to clonal performance and combining ability in orchardgrass, *Dactylis glomerata* L. *Agron. Jour.* 46:370-374. 1954.
- _____, joint author. See under Kalton.
459. Legg, J.O. and C.A. Black. Determination of organic phosphorus in soils. II. Ignition method. *Soil Sci. Soc. Amer. Proc.* 19:139-143. 1955.
- _____, joint author. See under Mehta.
- Legvold, Sam, joint author. See under Rossing.
- Lentz, C.F., joint author. See under Fassel.
- Levertson, R., joint author. See under Swanson.
- Lewis, W.E., joint author. See under Pohn.
460. Lilly, J.H. Where are we going in soil insect control? *Proc. 10th Ann. Meet. North Central Branch Ent. Soc. Amer.* 10:14-15. 1955.
461. _____. Insecticide fertilizer mixtures. *Natl. Fertilizer Rev.* 30:3-6. 1955.
462. _____. Insecticides help control soil insects. *Better Farming Methods* 26:1-2. 1954.
463. _____. Many advantages seen for fertilizer-pesticides in controlling soil insects. *Croplife* 2:7, 18. 1955.
464. _____. Recent development in the use of soil insecticides. *FAO Plant Protection Bull.* 3:81-85. 1955.
465. _____. How new chemicals are being applied to control rootworms. *Implement and Tractor* 70:22-24, 64. 1955.
466. _____ and Harold Gunderson. Soil insecticides ... crop insurance. *Iowa Farm Sci.* 9:9-11. 1955.
- _____, joint author. See under Fredericksen, Walstrom.

- Lind, E.F., joint author. See under Frans.
- Linderström-Lang, K., joint author. See under Anfinsen.
- Lindquist, Donald A., joint author. See under Fay, McCauley.
- Lindstrom, E.W., joint author. See under Griffing.
- Liston, Glenn C., joint author. See under Rutledge.
- Liu, C.H., joint author. See under Catron.
467. Lockhart, William R. A rapid micromethod for bioautographic assay. *Appl. Microbiol.* 3:152-153. 1955.
- Lohman, F.H., joint author. See under Diehl.
468. Long, W.H. and J.R. Dogger. Seasonal history studies on the southern corn rootworm and its relation to peanut culture in North Carolina. *Jour. Elisha Mitchell Sci. Soc.* 71:1-8. 1955.
469. Loomis, W.E. Resistance of plants to herbicides. Chapter in: "Origins of Resistance to Toxic Agents." pp. 99-120. 1st ed. Academic Press. 1955.
470. _____. Factôres fisiológicas que limitan as colheitas. *Rev. Agric.* 29:303-315. 1954.
471. _____. E.S. Rodgers, R.L. Warden, and R.E. Frans. Association of American Railroads Cooperative weed control project. *Bull. Amer. Rwy. Engr. Assoc.* 56:718-724. 1955.
- _____, joint author. See under Chapman, Dale, Gleason, Hammond, L.C., Mueller.
- _____, Associate Editor, *Plant Physiology*.
- _____, Editor, *Plant Physiology Section, Biological Abstracts*.
472. Lorch, Fred W. Mark Twain's lecture tour of 1868-1869: The American vandal abroad. *Amer. Literature* 26:515-527. 1955.
- Lovely, W.G., joint author. See under Brindley, Buchele, Cox.
- Lovestead, Howard S., joint author. See under McComb.
473. Lowe, Belle. *Experimental Cookery*. 4th ed. 573 pp. John Wiley and Sons, Inc. 1955.
- Lucas, George B., joint author. See under Hammond, George S.
- Ludlow, Patricia, joint author. See under Pattison.
- Lunan, Kenneth, joint author. See under Sinsheimer.
- Lundberg, W.O., joint author. See under Hammond, Earl G.
474. Lunden, Walter A. Interpretation of changes in crimes in Japan. *Jour. Criminal Law, Criminology, and Police Sci.* 46:243-246. 1955.
475. _____. *Offenders in Court and in Prison*, 1955. Iowa State College Bookstore. 125 pp. 1955.
- _____, Abstract Editor, *Excerpta Medica*.
476. Lush, Jay L. Estimates of heritability in a breeding program. Chapter in: "Breeding Beef Cattle for Unfavorable Environments", edited by Albert O. Rhoad. University of Texas Press. 1st ed. pp. 113-126. 1955.
477. _____. Breeding structure of populations. I. General Considerations. Chapter 42 in: "Statistics and Mathematics in Biology", edited by Oscar Kempthorne et al. Iowa State College Press. 1st ed. 632 pp. 1954.
478. _____. Rates of genetic changes in populations of farm animals. *Proc. 9th Internatl. Congr. Genetics (Caryologia)* Florence, Italy. 6:589-599. 1954.
479. _____. and Lon D. McGilliard. Proving dairy sires and dams. *Jour. Dairy Sci.* 38:163-180. 1955.
- _____, joint author. See under Carneiro, Dickerson.
- Lyle, Mary S., joint author. See under Chadderdon.
480. Lyon, C.A., R.L. Handy, and D.T. Davidson. Property variations in the Wisconsin loess of east-central Iowa. *Proc. Iowa Acad. Sci.* 61:291-312. 1954.
- _____, joint author. See under Handy.
481. McCauley, Robert H. Jr., Mary M. Grainger, Donald A. Lindquist, and Richard W. Fay. Laboratory comparison of some insecticides as larvicides against nonresistant house flies. *Jour. Econ. Ent.* 48:269-273. 1955.
482. McComb, A.L. and Norman J. Hansen. A naturally occurring aspen-poplar hybrid. *Jour. Forestry* 52:528-529. 1954.
483. _____. and Howard S. Lovestead. Viability of cottonwood seeds in relation to storage temperature and humidities. *Tree Planter's Notes. Forest Serv. U.S.D.A. No. 17.* pp. 9-11. 1954.
- _____, joint author. See under Dale.
- McDonald, Eileen M., joint author. See under Dachtler.

484. McHugh, Richard B. and William A. Owens, Jr. Age changes in mental organization -- A longitudinal study. *Jour. Gerontology* 9:296-302. 1954.
485. McJimsey, Harriet T. *Costume Selection*. Burgess Publishing Co., Minneapolis, Minn. 1st ed. 84 pp. 1955.
486. McKee, Dean and Earl O. Heady. How big will our farms get? *Iowa Farm Sci.* 9:3-6. 1955.
_____, joint author. See under Heady, Kehrberg.
487. McKinney, Leonard L., Eugene H. Uhing, John L. White, and Joseph C. Picken, Jr. Autotoxidation of products of trichloroethylene. *Agric. and Food Chem.* 3:413-419. 1955.
_____, joint author. See under Picken.
McMiller, P.R., joint author. See under Muckenhirn.
488. McNabb, Harold S. Jr. Tagging method for rapid recognition of culture tubes. *Plant Dis. Reporter* 39:437. 1955.
489. _____. Variations among isolates of *Fomes annosus* (Fr.) Cke. of different geographical and host origin. *Huitième Congrès International de Botanique, Paris 1954, Rapports et Communications parvenu avant le Congrès, Sect.* 20. p.133. 1954.
490. _____. The status of oak wilt in Iowa. *Proc. Iowa Acad. Sci.* 61:141-148. 1954.
491. _____. Oak wilt research--a lesson in cooperation. *Ames Forester* 42:17-19. 1955.
McWilliams, Richard, joint author. See under Scholl.
- Maa, Tsing-chao, joint author. See under Drake.
- Maddock, H.M., joint author. See under Ashton, Jensen, A.H.
492. Mahlstede, J.P. Handling and packaging perennials for shipment. *Amer. Nurseryman* 100:17, 98. 1954.
493. _____. Shipping roses in polyethylene wraps. *Amer. Nurseryman* 101:12. 1955.
494. _____. Chemical weed control in the nursery. *Amer. Nurseryman* 101:11, 106. 1955.
495. _____. A review of retail packaging: From boxes to polyethylene. *Amer. Nurseryman* 101:9, 125. 1955.
496. _____. Handling evergreens for shipment. *Amer. Nurseryman* 100:12, 92. 1954.
497. _____ and L.K. Kirk. Polyethylene: A solution to nursery shipping problems. *Amer. Nurseryman* 100:7, 64. 1954.
_____, joint author. See under Kirk.
498. Mairs, P., E. Willis, and P. Swanson. We've counted the calories in your diet. *Iowa Farm Sci.* 9:651-654. 1955.
_____, joint author. See under Swanson, P., Willis.
Malaby, K.L., joint author. See under Winders.
- Maldonado-Capriles, J., joint author. See under Drake.
499. Margoshes, M. and V.A. Fassel. The infrared spectra of aromatic compounds. I. The out-of-plane C-H bending vibrations in the region 625-900 Cm^{-1} . *Spectrochimica Acta* 7:14-24. 1955.
500. _____ and V.A. Fassel. Infrared functional group analysis of aryl silanes. *Analyt. Chem.* 27:351-355. 1955.
Marshall, F.J., joint author. See under Gilman.
501. Martin, Don S. Jr., and F.J. Hughes. Kinetics of hydrogen peroxide oxidation of selenious acid. *Jour. Phys. Chem.* 59:410-415. 1955.
_____, joint author. See under Grantham, Hughes, F.J.
502. Martin, T.G., G.E. Stoddard, and R.S. Allen. The effects of varied rates of hay feeding on body weight and production of lactating dairy cows. *Jour. Dairy Sci.* 37:1233-1240. 1954.
Marvin, K.R., joint author. See under Dodds.
- Massie, S.M., joint author. See under Gilman.
503. Mathews, John and D.A. Buthala. Electrophoretic determination of the immunological response of swine to hog cholera. *Vet. Med.* 50:213-215. 1955.
_____, joint author. See under Schwarte.
504. Meen, R.H. and H. Gilman. Preparation and cleavage of some organosilanes containing heterocyclic groups. *Jour. Organ. Chem.* 20:73-81. 1955.

505. Mehta, N.C., J.O. Legg, C.A.I. Goring, and C.A. Black. Determination of organic phosphorus in soils: I. Extraction method. *Soil Sci. Soc. Amer. Proc.* 18:443-449. 1955.
506. Melampy, R.M. and L.F. Cavazos. Comparative study of lipids in vertebrate testes. *Proc. Soc. Exptl. Biol. and Med.* 87:297-303. 1954.
507. _____. Review of: "Artificial Breeding and Livestock Improvement" by G.W. Stamm. (Windsor Press, Chicago, 1954. 304 pp.) *Jour. Heredity* 46:71. 1955.
_____, joint author. See under Cavazos, Duncan.
Meldrum, H.R., joint author. See under Dumenil, Pesek, Stanford.
508. Merchant, I.A. The challenge to Veterinary Public Health. *Proc. Amer. Vet. Med. Assoc.* 91st Ann. Meet; pp.439-441. 1954.
509. Merten, Helmut and Henry Gilman. Some long-chained organosilicon compounds. *Jour. Amer. Chem. Soc.* 76:5798. 1954.
Metcalfe, D.S., joint author. See under Thompson, H.E., Wiggans.
510. Metzler, David E. and Esmond E. Snell. Spectra and ionization constants of the vitamin B₆ group and related 3-hydroxy-pyridine derivatives. *Jour. Amer. Chem. Soc.* 77:2431-2437. 1955.
Miller, F.A., joint author. See under Hansen, R.S.
Miller, G.H., joint author. See under Herwig.
Miller, J.C., joint author. See under Kehr.
511. Millett, M.L. Rolling friction. *Iowa Engineer* 55:40-41. 1955.
Monroe, John, joint author. See under Davey.
Monson, Paul H., joint author. See under Beal.
Morgan, Agnes Fay, joint author. See under Arnrich.
512. Morgan, Paul E. and E. Robert Baumann. Recent sewage treatment plant costs in Iowa. *Jour. Sewage and Indust. Wastes* 27:257-260. 1955.
513. Morinaga, H. Effects of isotopic spin selection rules on photonuclear yields. *Phys. Rev.* 97:444-446. 1955.
514. Muckenhirn, R.J., L.T. Alexander, R.S. Smith, W.D. Shrader, F.F. Riecken, P.R. McMiller, H.H. Krusekopf. Loess-derived gray-brown podzolic soils in the upper Mississippi River valley. *University of Illinois Agric. Expt. Sta. Bull.* 587. 80 pp. 1955.
515. Mueller, L.E., P.H. Carr, and W.E. Loomis. The submicroscopic structure of plant surfaces. *Amer. Jour. Bot.* 41:593-600. 1954.
516. Murie, R., and Harvey Diehl. Effects of heating vitamin B₁₂ in a stream of dry nitrogen at various temperatures. *Iowa State Coll. Jour. Sci.* 29:143-145. 1954.
Murphy, D.R., joint author. See under Tiffany.
517. Murphy, Glenn. Objectives of advanced programs in mechanics. *Bull. Mech. Div. - Amer. Soc. Engr. Educ.* 1:12-16. 1954.
518. _____. Report on the Carnegie conference. *Jour. Engr. Educ.* 45:418-422. 1955.
519. _____. Complementary role of physics and mechanics in education. *Jour. Engr. Educ.* 45:753-757. 1955.
520. Murphy, H.C. Registration of oat varieties, XIX. *Agron. Jour.* 46:525. 1954.
_____, joint author. See under Browning, Finkner, Simons.
521. Murray, William G. State action for better assessments. *State Govt.* 28:90-94. 1955.
_____. Iowa farm values up again. *Iowa Farm Sci.* 9(8):12-13. 1955.
523. _____. Improving property assessments in the midwest. *North Central Land Tenure Com.* 1st prelim. ed. 48 pp. 1954.
_____, joint author. See under Aandahl.
524. Nadler, M.R. and E.S. Fitzsimmons. Preparation and properties of calcium zirconate. *Jour. Amer. Ceramic Soc.* 38:214-217. 1955.
Nady, Robert M., joint author. See under Csanyi.
Nelson, F.E., joint author. See under Bennett, Lawton, van der Zant, Wilkowske.
525. Nelson, Floyd Wm. How to build an efficient volunteer fire department. *Fire Engr.* 108:383-470. 1955.
_____, joint author. See under Brosier.
Newton, R.C., joint author. See under Parker.

526. Nichols, R.T., A.V. Pohm, J.H. Talboy, Jr., and E.N. Jensen. A beta-ray spectrometer for coincidence measurements. *Rev. Sci. Instruments* 26: 580-583. 1955.
Nicholson, Ray, joint author. *See under* Pesek, John.
Nomsen, Richard, joint author. *See under* Kozicky.
527. Nordin, J.A., George Judge, and Omar Wahby. Application of econometric procedures to the demands for agricultural products. *Iowa Agric. Expt. Sta. Res. Bull.* 410. 1954.
528. Nordskog, A.W. and F.J. Ghostley. Heterosis in poultry. *Poultry Sci.* 33: 704-715. 1954.
_____, joint author. *See under* Farnsworth.
529. Norris, Dale M., Jr. Natural spread of *Endoconidiophora fagacearum* Bretz to wounded red oaks in Iowa. *Plant Dis. Reporter* 39:249-253. 1955.
Northrop, L., joint author. *See under* Chung, Everson.
530. Novak, Ralph S. Up for review--The guaranteed annual wage. *Natl. Assoc. Cost Accts. Bull.* 36(11):1515-1521. 1955.
531. Oita, K., R.G. Johnson, and H. Gilman. The chlorination of dibenzofuran and some of its derivatives. *Jour. Organ. Chem.* 20:657-667. 1955.
_____, joint author. *See under* Gilman.
Olson, Russell O., joint author. *See under* Heady.
532. Orr, Robert W. A few aspects of acquiring serials. *Libr. Trends* 3:393-402. 1955.
533. Ostle, Bernard. *Statistics in Research*. 2nd ed. 496 pp. Iowa State College Press. 1954.
534. Ott, John L. and C.H. Werkman. Bacterial formation of adenosine. Properties of the cell-free enzyme system in *Escherichia coli*. In: *Biochemistry of Nitrogen*. pp. 174-180. 1955.
Ottesen, M., joint author. *See under* Anfinson.
535. Oulman, Charles S., joint author. *See under* Baumann, E.
535. Owens, William A. Jr. A reply to Drs. Peck-Stephenson. *Jour. Appl. Psychol.* 38:371-372. 1954.
536. _____ and Duane E. Thompson. Differential effects of age upon intellectual functions differing in degree of genetic conditioning. *Jour. Gerontology* 9:303-305. 1954.
_____, joint author. *See under* McHugh.
537. Paddock, Floyd B. Report of the State Apiarist for Year Ending December 31st, 1954. 125 pp. Des Moines. 1955.
Parchen, F.R., joint author. *See under* Duke.
538. Parker, J.R., R.C. Newton, and R.L. Shotwell. Observations and mass flights and other activities of the migratory grasshopper. *USDA Tech. Bull.* 1109. 46 pp. 1955.
539. Parks, Laurence R., George S. Hammond, and M. Frederick Hawthorne. Substituent effects on the reaction of benzoate ions with 2,4-dinitrochlorobenzene and methyl iodide. *Jour. Amer. Chem. Soc.* 77:2903-2904. 1955.
_____, joint author. *See under* Hammond, George S.
540. Parks, W. Robert. The meaning of democracy. *Farm Policy Forum* pp. 2-8. Winter, 1955.
541. _____ and Joseph F. Wall. *Freedom*. Iowa Farm Bur. Fed. 68 pp. 1955.
_____, Technical Editor, winter, 1955, *Farm Policy Forum*.
Parmelee, C.E., joint author. *See under* Wilkowske.
542. Pattillo, Walter H. and Elery R. Becker. Cytochemistry of *Eimeria brunetti* and *E. acervulina* of the chicken. *Jour. Morphology* 96:61-96. 1955.
543. Pattison, Mattie, Patricia Ludlow, Helen S. Paulson. Teaching foods in a short period. *Jour. Home Econ.* 46:707-712. 1955.
Paulson, Helen S., joint author. *See under* Pattison.
Peabody, R.C., joint author. *See under* Brown, F.E.
Pearson, E.S., joint author. *See under* David.
544. Peet, Louise J. and Lenore Sater Thyne. *Household Equipment*. 4th ed. 443 pp. John Wiley and Sons, Inc. 1955.
545. Pennell, Ellen. Ideas to help you with public relations. *Pract. Home Econ.* 33:9,39-44. 1955.
546. _____. Why they love to stay home. *Town Jour.* 62:56-58. 1955.

- Penny, L.H., joint author. See under Dicke.
547. Penquite, Robert and R.E. Phillips. Laboratory exercises in poultry husbandry. Iowa State College Press. 3rd ed. 1955.
- Pesek, I., joint author. See under Beegle, Swanson.
548. Pesek, John. The absorption of phosphorus from phosphate fertilizers applied to established meadows. III. Effect of phosphorus source and placement and of a meadow composition. Soil Sci. Soc. Amer. Proc. 19:63-66. 1955.
549. _____ and Lloyd Dumenil. How much fertilizer carryover? What's New in Crops and Soils 7:10-11. 1955.
550. _____, Ray Nicholson, and Clifford Spies. What about fertilizers in dry years? Iowa Farm Sci. 9(10):3-6. 1955.
551. _____, R.P. Nicholson, and H.R. Meldrum. Don't neglect lime! Iowa Farm Sci. 9(7):21-24. 1955.
552. _____ et al. Profitable use of fertilizer in the midwest. University of Wisconsin Agric. Expt. Sta. North Central Reg. Publ. No. 54. 1954.
- _____, joint author. See under Dumenil, Heady, Webb.
- Peterson, Conrad, joint author. See under Boozer.
553. Peterson, G.A. and Earl O. Heady. Application of input-output analysis to a simple model involving agriculture. Iowa Agric. Expt. Sta. Res. Bull. 427. 1955.
554. Peterson, Lewis E. and E.P. Lana. Irrigation, fertilizer and cultural studies with sweet potatoes, muskelons and watermelon on Buckner coarse sand. Trans. Iowa State Hort. Soc. 88:237-252. 1953.
- _____, joint author. See under Lana.
- Peturson, B., joint author. See under Simons.
- Phillips, Donald L., joint author. See under Spangler.
555. Phillips, Richard. Iowa economist sees need for changing buying practices also. Soybean Digest p. 19. February, 1955.
556. _____. Avoid discounts when selling grain. Hoard's Dairyman 10: 1955.
557. _____ and Ralph W. Hansen. More grain storage on Iowa farms. Iowa Farm Sci. 9(1): 1954.
- _____, joint author. See under Beal, Beneke, Dachtler, Hansen, R.W.
- Phillips, R.E., joint author. See under Donovan, Penquite.
558. Picken, Joseph C. Jr., Norman L. Jacobson, Robert S. Allen, Harry E. Biester, Paul C. Bennett, Leonard L. McKinney, and John C. Cowan. Toxicity of trichloroethylene-extracted soybean oil meal. Agric. and Food Chem. 3:420-424. 1955.
- _____, joint author. See under Bauriedel, McKinney.
- Pickett, Mary S., joint author. See under Ehrenkranz.
559. Pidd, R.W., C.L. Hammer. Electron scattering on tungsten at 31, 40 and 60 mev. Phys. Rev. 99:1396-1400. 1955.
- _____, joint author. See under Hammer.
- Plaumann, Fritz, joint author. See under Drake.
560. Plumb, Galen R. and Don C. Charles. Scoring difficulty of Wechsler comprehension responses. Jour. Educ. Psychol. 46:179-183. 1955.
561. Pohl, Richard W. A rapid softening agent for dried plant structures. Proc. Iowa Acad. Sci. 61:149-150. 1954.
562. Pohm, A.V., W.E. Lewis, J.H. Talboy, Jr., and E.N. Jensen. The beta-spectra of Pr^{142} , Tm^{170} , and Rb^{86} . Phys. Rev. 95:1523-1529. 1954.
563. _____, R.C. Waddell, J.P. Powers, and E.N. Jensen. Beta spectrum of C^{14} . Phys. Rev. 97:432-433. 1955.
- _____, joint author. See under Nichols, R.T., Willis.
564. Porter, A.R. and R.S. Allen. Dairy cows like ground corn in grass silage. Iowa Farm Sci. 9:6-7. 1954.
565. Porter, Blaine M. The relationship between marital adjustment and parental acceptance of children. Jour. Home Econ. 47:157-164. 1955.
- Powell, J.E., joint author. See under Spedding.
- Powers, J.P., joint author. See under Pohm.
- Prill, R.C., joint author. See under Ruhe.
- Pudelkewicz, C., joint author. See under Everson.
- Quaife, E.L., joint author. See under Catron.
- Quinney, B., joint author. See under Fassel.
- Quinney, P.R., joint author. See under Duke.

566. Racusen, D.W. and S. Aronoff. Metabolism of soybean leaves. VI. Exploratory studies in protein metabolism. Arch. Biochem. and Biophys. 51:68-78. 1954.
567. Ramsey, Frank K. The pathology of mucosal disease of cattle. Proc. 91st Meet. Amer. Vet. Med. Assoc. pp. 162-167. 1954.
568. Raun, Earle S. Weight gains in feeder calves treated with low pressure rotenone sprays to control cattle grubs. Jour. Econ. Ent. 48:
569. _____. Use of synergized pyrethrins to prevent oviposition by cattle grubs. Jour. Econ. Ent. 48:
570. _____. Pestproof your livestock. Successful Farming. pp. 52-98. May, 1953.
571. _____. Fly control ... is it possible in 1953? Hoard's Dairyman p.429. May 10, 1953.
- _____, joint author. See under Dahm.
- Reid, Th.L., joint author. See under Wenkert.
572. Repp, Ward W., W.H. Hale, E.W. Cheng, and Wise Burroughs. The influence of oral administration of non-protein nitrogen feeding compounds upon blood ammonia and urea levels in lambs. Jour. Animal Sci. 14:118-131. 1955.
- Riecken, F.F., joint author. See under Muckenhirn, Ruhe, Swenson.
- _____, Associate Editor, Soil Science Society of America Proceedings.
- Risch, J., joint author. See under Gilman.
573. Robbins, Russell. Flushing bar saves pheasants. Iowa Farm Sci. 9:1. 1954.
574. Robbins, William E. and Paul A. Dahm. Absorption and excretion, distribution, and metabolism of carbon-14-labeled DDT by the American cockroach. Agric. and Food Chem. 3:500-508. 1955.
575. Roberson, Opal and Normal Hollen. New Fabrics. More wear with good care. Agric. Ext. Bull. HE31. 8 pp. 1955.
- Roberts, H., joint author. See under Swanson, Willis.
- Roberts, P.H., joint author. See under Beegle.
576. Robertson, Fred. Review of trigonometry texts. Amer. Math. Month. 61:656-657. 1954
- Robinson, Joe L., joint author. See under Hutchcroft.
577. Robotka, Frank. A theory of co-operation. Zeitschrift für das Gesamte Genossenschaftswesen. Univ. of Erlangen, W.Germany. 5:155-175. 1955.
- Roderuck, C.R., joint author. See under Eppright, Garcia.
- Rodgers, E.G., joint author. See under Loomis.
578. Rollins, Ralph L., M.G. Spangler, and Don Kirkham. Movement of soil moisture under a thermal gradient. Proc. Highway Res. Bd. 33:492-508. 1954.
- Rosdail, J.R., joint author. See under Groth.
- Ross, Alan, joint author. See under Hartley.
579. Ross, E.D. The college textbook in American history before the war era. Social Studies 46:14-21. 1955.
580. _____. James F. Wilson, legalistic free-soiler. Ann. of Iowa 32:365-375. 1954.
- _____, Editorial Board, Social Science.
581. Rossing, Thomas D. and Sam Legvold. Collision excitation of molecular vibrations in halogen-substituted methanes. Jour. Chem. Phys. 23:1118-1125. 1955.
- Royer, Keith, joint author. See under Brosier.
- Roze, Janis A., joint author. See under Drake.
582. Ruhe, R.V. Relations of the properties of Wisconsin loess to topography in western Iowa. Amer. Jour. Sci. 252:663-672. 1954.
583. _____. and J.G. Cady. Latosolic soils of central African interior high plateaus. Trans. 5th Internatl. Congr. Soil Sci. 4:401-407. 1954.
584. _____. R.C. Prill, and F.F. Riecken. Profile characteristics of some loess-derived soils and soil aeration. Soil Sci. Soc. Amer. Proc. 3:345-347. 1955.
585. _____. and W.H. Scholtes. Radiocarbon dates in central Iowa. Jour. Geology 63:82-92. 1955.
586. Rundle, R.E. Review of: "Quantum Chemistry", by K.S. Pitzer. Jour. Amer. Chem. Soc. 77:253. 1955.
- _____, joint author. See under Daane.
- Russell, W.A., joint author. See under Hooker.

587. Rutledge, Glenn, Gladys L. Harrison, John F. Heer, Glenn C. Liston and David B. Tolman. Our periodicals tell the story. Amer. Assoc. Agric. Coll. Editors 37:1-3. 1955.
- Saddler, Jane, joint author. See under Hollen.
588. Schaller, F.W. Fertilizer helps control erosion. Iowa Farm Sci. 9:20-22. 1955.
589. Schaller, F.W., and D.D. Evans. Some effects of mulch tillage. Agric. Engr. 35:731-734. 1954.
590. Schaller, F.W. and W.E. Larson. Effect of wide spaced corn rows on corn yields and forage establishment. Agron. Jour. 47:271-276. 1955.
_____, joint author. See under Hawk.
- Schellman, J., joint author. See under Anfinssen.
591. Schmidt, C.H., Studies of synergists for pyrethrins and related synthetic compounds. Proc. 10th Ann. Meet. North Central Branch Ent. Soc. Amer. 10:56-60. 1955.
592. Schmidt, Louis Bernard. Andrew Jackson and the agrarian west. Current Hist. 28:331-330. 1955.
593. _____. Agriculture in transition. Current Hist. 26:74-80. 1954.
594. Scholl, J.M. An attachment for a field chopper for measuring area harvested. Agron. Jour. 47:105. 1955.
595. _____. H.D. Hughes, and Richard McWilliams. Renovation can double pasture production. Iowa Farm Sci. 9:7-8. 1955.
596. _____. and H.E. Thompson. Birdsfoot trefoil ... best legume for permanent pastures. Iowa Farm Sci. 10:3-5. 1955.
_____, joint author. See under Heady.
- Scholtes, W.H., joint author. See under Aandahl, Ruhe.
597. Schuldt, Paul H. Comparison of anthracnose fungi on oak, sycamore, and other trees. Contributions from the Boyce Thompson Inst. 18:85-107. 1955.
598. Schwab, G.O. Plastic tubing for subsurface drainage. Agric. Engr. 36:86-89, 92. 1955.
_____, joint author. See under Frevert,
599. Schwarte, L.H. Stability of hog cholera virus. Vet. Med. 49:375-376. 1955.
600. _____. and John Mathews. Aerosol properties of lyophilized hog cholera virus. Vet. Med. 49:233-234. 1954.
601. _____. Transmission of hog cholera via the respiratory tract. North Amer. Vet. 35:671-672. 1954.
_____, joint author. See under Zimmerman.
602. Scott, A.D. and D.D. Evans. Dissolved oxygen in saturated soil. Soil Sci. Soc. Amer. 19:7-12. 1955.
_____, joint author. See under Evans.
603. Scott, Jesse F. and Robert L. Sinsheimer. Technique of Study of Biological Effects of Ultraviolet Radiation. Chap. IV in "Radiation Biology", Vol. II. pp. 119-163. McGraw-Hill. 593 pp. 1955.
- Seagraves, James, joint author. See under Heady.
- Sealock, R.R., joint author. See under Brierly.
604. Semeniuk, G. Microflora. Chap. III in: "Storage of cereal grains and their products." Amer. Assoc. Cereal Chem. Monogr. Ser. 2:77-151. 1954.
- Sen, Jyotirindra N., joint author. See under Boozer, Hammond, G.S.
- Shaw, K.G., joint author. See under Smutz.
605. Shaw, R.H. A comparison of two types of dew duration recorders. Air Force Cambridge Res. Center Spec. Rept. No. 2. 40 pp. 1955.
606. _____. Climatic requirement. Chap. 7 (24 pp.) in: "Corn and Corn Improvement." Monograph of Amer. Soc. Agron. Ed. by G.F. Sprague. Academic Press. 1st ed. 1955.
607. _____. Comparison of dew duration on Duvdevan's dew gauges and several crop cover. Air Force Cambridge Res. Center Spec. Rept. No. 1. 18 pp. 1954.
608. _____. H.C.S. Thom, and G.L. Barger. The climate of Iowa I. The occurrence of freezing temperatures in spring and fall. Iowa State Coll. Spec. Rept. No. 8. 88 pp. 1954.
_____, joint author. See under Wiggans, Wilsie.
- Shaw, Russell, joint author. See under Heady.
609. Shaw, W.C. Microforceps for handling very small specimens. Rev. Sci. Instruments 26:238. 1955.

610. _____, D.E. Hudson, and G.C. Danielson. Preparation of microscopic crystals for electrical measurements. *Rev. Sci. Instruments* 26:237-238. 1955.
Shedd, C.K., joint author. See under Hukill, Thompson, H.J.
Sheeler, J.B., joint author. See under Chu.
611. Shepherd, G.S. What can a research man do in agricultural price policy? *Jour. Farm Econ.* 37:305-314. 1955.
612. _____. *Marketing Farm Products--Economic Analysis*. Iowa State Coll. Press. 3rd ed. 488 pp. 1955.
613. _____. When does it pay to store grain? *Iowa Farm Sci.* 9:23-26. 1954.
614. _____. Changes in structure. in: *The Yearbook of Agriculture, 1954*. pp.52-59. USDA. 1955.
615. _____. There's a bigger job for parity. *Co-op Grain Quart.* 12:14-21. 1954-55.
616. _____. Surplus disposal and domestic market expansion. in: *United States Agriculture: Perspectives and Prospects*. American Assembly-Graduate School of Business, Columbia University. 1955.
_____, joint author. See under Brough, Wiegmann.
617. Sherf, A.F. and M.C. Shurtleff. Rusts in grain crops. *Iowa Agric. Ext. Serv. Pamphlet* 185 (rev.) 1955.
Shirley, D.A., joint author. See under Gilman.
Shotwell, R.L., joint author. See under Parker, J.R.
Shrader, W.D., joint author. See under Muckenhirn.
Shull, H., joint author. See under Goodman.
618. Shurtleff, Malcolm C. Jr. Control of turf brown patch. *Rhode Island Expt. Sta. Bull.* No.328. 1955.
_____, joint author. See under Sherf, Sylwester.
Sidwell, V.D., joint author. See under Eppright.
619. Simons, M.D. Adult plant resistance to crown rust of certain oat selections. *Phytopath.* 45:275-278. 1955.
620. _____. Physiologic races of crown rust of oats identified in 1953. *Plant Dis. Reporter* 38:649-652. 1954.
621. _____, and H.C. Murphy. Inheritance of resistance to two races of *Puccinia coronata* Corda var. *avenae* Fraser and Led. *Proc. Iowa Acad. Sci.* 61:170-176. 1954.
622. _____. B. Peturson, and H.C. Murphy. Purification of differential oat varieties for identification of races of crown rust. *Plant Dis. Repr.* 39:23-24. 1955.
_____, joint author. See under Browning.
Simpson, Wm. R., joint author. See under Brosier.
623. Sinsheimer, Robert L. Nucleotides from T_{2V} bacteriophage. *Science* 120:551-553. 1954.
624. _____. The photochemistry of uridylic acid. *Radiation Res.* 1:505-513. 1954.
625. _____. Ultraviolet absorption spectra. Chap. V in: *Radiation Biology, Vol. II*. pp.165-201. McGraw-Hill. 1955.
626. _____ and Kenneth Lunan. Nucleic acid from T₇ bacteriophage. *Fed. Proc.* 14:282. 1955.
627. Skodje, M.T. and E. Robert Baumann. A college town finances sewage treatment. *Water and Sewage Works* 102:228-230. 1955.
_____, joint author. See under Baumann.
628. Slater, James A. A revision of the subfamily Pachygronthinae of the world (*Hemiptera:Lygaeidae*). *Philippine Jour. Sci.* 84:1-160. 1955.
629. Sloss, Margaret W. The microscopic anatomy of the digestive tract of *Sus scrofa domestica*. *Amer. Jour. Vet. Res.* 15:578-593. 1955.
Smith, A.G., joint author. See under Gilman.
Smith, Frederick G., joint author. See under Throneberry.
Smith, H.G., joint author. See under Daane.
Smith, J., joint author. See under Beegle.
630. Smith, J.F. Downie. Improving the teaching of engineering. *Jour. Engr. Educ.* 45:73-79. 1954.
631. Smith, J.F. Electrostatic energy calculations for sodium tungsten bronze (Na_xWO₃). *Phys. Rev.* 95:1369-1372. 1954.
Smith, R.S., joint author. See under Muckenhirn.
632. Smutz, M., G.L. Bridger, K.G. Shaw, and M.E. Whatley. The Ames process for separation of monazite. *Chem. Engr. Progr. Symp. Ser.* 13:167-170. 1954.

633. Snedecor, George W. Review of: "A primer of Statistics for Political Scientists, by V.O. Key, Jr. Jour. Amer. Statist. Assoc. 50:608-609. 1955.
_____, Editorial Board, Biometrics.
_____, Editor, Queries Section, Biometrics.
Snell, Esmond E., joint author. See under Metzler.
634. Spangler, M.G. Secondary stresses in buried high pressure pipe lines. The Petrol. Engr. Nov. 1954. Engr. Expt. Sta. Rept. No.23. 10 pp. 1954-55
635. _____ and Donald L. Phillips. Deflections of timber-strutted corrugated-metal-pipe culverts under earth fills. Highway Res. Bd. Bull. 102. pp.12-18. 1954.
_____, joint author. See under Rollins.
636. Spedding, F.H. and J.E. Powell. A practical separation of yttrium group rare earths from gadolinite by ion-exchange. Chem. Engr. Progr. Symposium Ser. 50:7-15. 1954.
637. _____, J.E. Powell, and H.J. Svec. A laboratory method for separating nitrogen isotopes by ion-exchange. Jour. Amer. Chem. Soc. 77:1393. 1955.
_____, joint author. See under Daane.
Speer, Vaughn, joint author. See under Catron.
Spies, Clifford, joint author. See under Pesek, John.
Spink, D.R., joint author. See under Beyer.
638. Sprague, G.F. Corn breeding. Chapt. 5 in: Corn and Corn Improvement, 71 pp. 1955.
639. _____, Industrial utilization. Chapt. 14 in: Corn and Corn Improvement. 24 pp. 1955.
640. _____. The world production of corn. Chapt. 16 in: Corn and Corn Improvement. 9 pp. 1955.
641. _____. Breeding for resistance to stalk rot. Proc. Hybrid Corn Ind.--Res. Conf. 9:38-43. 1955.
_____, joint author. See under Hooker.
_____, Associate Statistical Editor, Journal of Agronomy
_____, Editor, Corn and Corn Improvement. 699 pp. Academic Press. 1955.
642. Sprugel, George Jr. The growth of bluegills (*Lepomis macrochirus*) in a new lake with particular reference to false annuli. Trans. Amer. Fish. Soc. 83:58-75. 1954.
643. _____. The growth of green sunfish (*Lepomis cyanellus*) in Little Wall Lake, Iowa. Iowa State Coll. Jour. Sci. 29:707-719. 1955.
644. Stacy, W.H. Guide for building your tomorrow's community. Agric. Ext. Serv. 6 pp. 1955.
645. _____. Farmers Organizations. In: 1955 ed. of Encyclopaedia Britannica.
Stadler, Janice, joint author. See under Gowen.
646. Stanford, George and John Hanway. Predicting nitrogen fertilizer needs of Iowa soils: II. A simplified technique for determining relative nitrate production in soils. Soil Sci. Soc. Amer. Proc. 19:74-77. 1955.
647. _____, John Hanway, and H.R. Meldrum. Effectiveness and recovery of initial and subsequent fertilizer applications on oats and the succeeding meadows. Agron. Jour. 47:25-31. 1955.
_____, joint author. See under Hanway.
Stebbins, Dean W., joint author. See under Decker.
648. Stewart, Lowell O. Surveying is at the crossroads--answer of the Educator. Proc. 3rd Natl. Surveying Teach. Conf. 1952. 1955.
Stiles, Wm.B., joint author. See under Higdon.
Stoddard, G.E., joint author. See under Martin, T.G.
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Svec, H.J., joint author. See under Spedding.
649. Swanson, P.P., R. Leverton, M.R. Gram, H. Roberts, and I. Pesek. Blood values of women: Cholesterol. Jour. Gerontology 10:41-47. 1955.
650. _____, H. Roberts, E. Willis, I. Pesek, and P. Mairs. Food intake and body weight of older women. A chapter in "Weight Control". pp. 80-96. Iowa State Coll. Press. 1955.
651. _____, and E. Willis. You don't outgrow your need for calcium. Iowa Farm Sci. 9:720-722. 1955.

652. _____, _____, and P. Mairs. Do Iowa women eat enough protein? Iowa Farm Sci. 9:698-700. 1955.
653. _____, _____, _____, Overweight--A problem among Iowa women. Iowa Farm Sci. 9:766-768. 1955.
_____, joint author. See under Epwright, Garcia, Mairs, Willis.
_____, Editorial Committee, Weight Control. A collection of papers presented at the Weight Control Colloquium.
Swayampati, Dhairyasheel R., joint author. See under Gilman.
654. Swenson, R.M. and F.F. Riecken. Movement of iron in the development of loess-derived Brunizem soils. Soil Science 79:177-186. 1955.
Swiss, J., joint author. See under Gilman.
655. Switzer, C.M. Effects of herbicides on mitochlondrial enzyme systems. Proc. North Central Weed Control Conf. 11:10-12. 1954.
656. Switzer, William P. Studies on atrophic rhinitis. Proc. Amer. Vet. Med. Assoc. 91st Ann. Meeting. pp.102-106. 1954.
657. _____. Observations on infectious atrophic rhinitis. Proc. U.S. Livestock Sanitary Assoc. 58th Ann. Meet. pp.363-364. 1954.
658. Sylwester, E.P., H. Gunderson, and M.C. Shurtleff. Spraying calendar. Hoard's Dairyman 100:454. 1955.
- Talboy, J.H. Jr., joint author. See under Nichols, Pohn.
659. Tamsma, A.F. A study of the volatile fraction isolated from oxidized milk fat. I. Isolation, preliminary characterization, and chromatographic separation of the volatile fraction. Jour. Dairy Sci. 38:284-291. 1955.
660. _____. A study of the volatile fraction isolated from oxidized milk fat. II. Further characterization of compounds responsible for the oxidized flavor. Jour. Dairy Sci. 38:487-498. 1955.
661. Tanner, Ward D. and George O. Hendrickson. Ecology of the Virginia Rail in Clay County, Iowa. Iowa Bird Life 24:65-70. 1955.
Tauber, Oscar E., joint author. See under Jones, J.C., Thompson, S.
662. Taylor, L.H. and R.E. Atkins. Effects of natural selection in segregating generations upon bulk populations of barley. Iowa State Coll. Jour. Sci. 29:147-162. 1954.
Terwilliger, K.M., joint author. See under Hammer.
Thom, H.C.S., joint author. See under Shaw, R.H.
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663. Thompson, H.E. and D.S. Metcalfe. Producing birdsfoot trefoil seed. Iowa Farm Sci. 9(12):6-8. 1955.
_____, joint author. See under Scholl, Wiggans.
664. Thompson, H.J. and C.K. Shedd. Equilibrium moisture and heat of vaporization of shelled corn and wheat. Agric. Engr. 35:786-788. 1954.
665. Thompson, Louis M. Opportunities in college teaching in the field of agronomy. Agron. Jour. 47:87-88. 1955.
666. Thompson, Sam H. Problems in choosing a hog marketing place. Iowa Farm Sci. 9:19-22. 1954.
667. Thompson, Sarane, Joseph F. Foster, John W. Gowen, and Oscar E. Tauber. Hereditary differences in serum proteins of normal mice. Proc. Soc. Exptl. Biol. and Med. 87:315-317. 1954.
668. Throneberry, Glyn O. and Frederick G. Smith. Seed viability in relation to respiration and enzymatic activity. Assoc. Offic. Seed Analysts, Proc. 44:91-95. 1954.
Thye, joint author. See under Peet.
669. Tiffany, Lois H., J.C. Gilman, and D.R. Murphy. Fungi from birds associated with wilted oaks in Iowa. Iowa State Coll. Jour. Sci. 29:659-706. 1955.
Ting, Y.C., joint author. See under Kehr.
670. Tintner, Gerhard. The distribution of the variances of variate differences in the circular case. Metron 17:1-12. 1955.
671. _____. The teaching of econometrics. In: The University Teaching of the Social Sciences--Economics. UNESCO on behalf of the Internatl. Econ. Assoc. pp. 282-295. Columbia University Press. 1st ed. 1954.
672. _____. Ein Beitrag zur nicht-statischen Werttheorie. Zeitschrift für National-ökonomie. 14(2-4):358-365. 1954.

673. _____. Review of: *Econométrie*. Colloques Internationaux du Centre National de la Recherche Scientifique, Paris 12-17 mai 1952. (Paris: Centre National de la Recherche Scientifique. 1953 pp.334, v.) *Amer. Econ. Rev.* 44:664-666. 1954.
674. _____. Review of: *Grundfragen der Ökonometrie*, by W. Winkler. *Zeitschrift für National Ökonomie*. 14:539-540. 1954.
675. _____. The use of mathematical methods in econometrics and economic statistics. *Internatl. Soc. Sci. Bull.* 6:640-651. 1954.
_____, joint author. See under Babbar.
_____, Associate Editor, *Metroeconomica*.
_____, Administrative Board of Editors, *Metroeconomica*.
_____, Book Review Editor, *Econometrica*.
_____, Associate Editor, *Econometrica*.
Tolman, David B., joint author. See under Rutledge.
- Uhing, Eugene H., joint author. See under McKinney.
676. Ulmer, Martin J. Experimental definitive hosts of *Fibricola cratera* (Barker and Noll, 1915) Dubois 1932 (*Trematoda Diplostomatidae*) *Abst. Jour. Parasitology* 40:33. 1954.
677. _____. Precociously developed brachylamid *Metacercariae* within sporocysts. *Proc. Iowa Acad. Sci.* 61:631-635. 1954.
Underkofler, Leland A., joint author. See under Bauriedel.
Ustrud, H.O., joint author. See under Higdon.
- VanderHaar, R.W., joint author. See under Banks.
VanderWal, R.P., joint author. See under Banks.
678. van der Zant, W.C., and F.E. Nelson. Characteristics of some endocellular peptidases of *Streptococcus lactis*. *Jour. Dairy Sci.* 37:795-804. 1954.
679. _____. Amino acids and peptides in the protein-free fraction of milk before and after incubation with *Streptococcus lactis*. *Jour. Dairy Sci.* 37:790-794. 1954.
680. van Schilfgaarde, Jan, R.K. Frevert, and Don Kirkham. A tile drainage field laboratory. *Agric. Engr.* 35:474-478. 1954.
681. Voelker, H.H., N.L. Jacobson, and R.S. Allen. Relationship of antibiotic feeding and the rate of growth to blood reducing sugar levels and glucose absorption in dairy calves. *Antibiotic Chemotherapy* 5:224-231. 1955.
Voigt, A.F., joint author. See under Horrocks.
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- Waddell, R.C., joint author. See under Pohm.
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682. Walstrom, R.J. and J.H. Lilly. Protect forage crops from insects. *Iowa Farm Sci.* 9(11):7-8. 1955.
Warden, R.L., joint author. See under Loomis.
Warwick, E.J., joint author. See under Dickerson.
Wassom, C.E., joint author. See under Kalton, Leffel.
683. Webb, J.R. and John Pesek. Evaluation of residual soil phosphorus in permanent fertility plots. *Soil Sci. Soc. Amer. Proc.* 18:449-453. 1954.
Webber, H.A., joint author. See under Fellingner.
684. Weber, C.R. Effects of defoliation and topping simulating hail injury to soybeans. *Agron. Jour.* 47:262-266. 1955.
Weiss, M.R., joint author. See under Kalton.
Welch, A. Philip, joint author. See under Crowley.
685. Wellman, F.E. Some suggestions for the interpretation of test results. *Guid-O-Gram* (State of Iowa, Dept. of Public Instruction publ.) pp.4, 5. Oct. 1954.
686. Wengert, Gerald and Charles Hutchcroft. Farmers plant many corn varieties. *Iowa Farm Sci.* 9(9):19. 1955.
687. Wenkert, E. The role of oxidation in the biogenesis of alkaloids. *Experientia* 10:346-350. 1954.
688. _____. Structural and biogenetic relationships in the diterpene series. *Chem. and Ind.* pp.282-284. 1955.

689. _____. The structure of tazettine. *Experientia* 10:476-480. 1954.
690. _____. The constitution of lycorine. *Chem. and Ind.* pp.1175-1177. 1954.
691. _____ and J.H. Hansen. The amaryllidaceae alkaloids. *Chem. and Ind.* pp.1262-1263. 1954.
692. _____ and Th.L. Reid. Intramolecular reactions in the oxindole series. *Experientia* 10:417. 1954.
- Werkman, C.H., joint author. See under Claridge, Fowler, Ott.
- _____, Assistant Editor, *Enzymologia*.
- _____, Editor, *Archives of Biochemistry and Biophysics*.
- West, D.W., joint author. See under Finkner.
- West, J.B., joint author. See under Beyer.
693. Weston, Henry G. The winter-spring movements of the ring-necked pheasant in northern Iowa. *Iowa State Coll. Jour. Sci.* 29:39-60. 1954.
- Whatley, J.A.Jr., joint author. See under Dickerson.
- Whatley, M.E., joint author. See under Smutz.
- White, John L., joint author. See under McKinney.
694. White, Wm.C. and C.A. Black. Wilcox's agrobiology: III The inverse yield-nitrogen law. *Agron. Jour.* 46:310-315. 1954.
695. Wick, James R. and Philip F. Bonhag. Postembryonic development of the ovaries of *Oncopeltus fasciatus* (Dallas). *Jour. Morphology* 96:31-60. 1955.
- Wickstrom, A.E., joint author. See under Chu.
696. Wiegmann, Fred H., E.S. Clifton, and G.S. Shepherd. Comparison of costs of service and self-service methods in retail meat departments. *Iowa Agric. Expt. Sta. Res. Bull.* 422. 10 pp. 1955.
697. Wiggans, S.C. Review of: Auxins and Plant Growth, by A. Carl Leopold. *Agron. Jour.* 47:238. 1955.
698. _____. The effect of planting date on maturity and yield of oats. *Amer. Soc. Agron. Abstr.* 1954 Natl. Meet. p.83. 1954.
699. _____. Germination of immature oat seeds. *Plant Physiol.* "Supple." 30:10. 1955.
700. _____ and K.J. Frey. The effect of photoperiodism on nitrogen nutrition and tillering capacity of oats. *Amer. Soc. Agron. Abst. of 1954 Natl. Meet.* p.77. 1954.
701. _____. The effect of nitrogen on growth and development of oats. *Plant Physiol. Abstr. of 1954 Meet.* p.77. 1954.
702. _____. The effect of increased daylengths on the production of greenhouse grown oats. *Agron. Jour.* 47:387. 1955.
703. _____, D.S. Metcalfe, and H.E. Thompson. The effect of chemicals on harvesting of brome grass (*Bromus inermis*) and birdsfoot trefoil (*Lotus corniculatus*) in Iowa. *Amer. Soc. Agron. Absts. of 1954 Natl. Meet.* p. 83. 1954.
704. _____ and R.H. Shaw. The effect of fluorescent and incandescent light on temperatures in photoperiodic chambers. *Iowa State Coll. Jour. Sci.* 30:139-144. 1955.
- _____, joint author. See under Browning.
- Wikén, T., joint author. See under Buchanan.
705. Wilder, D.R. High temperature laboratory furnaces. *Ames Lab. ISC-551.* 23 pp. 1954.
706. _____ and E.S. Fitzsimmons. Further study of sintering phenomena. *Jour. Amer. Ceramic Soc.* 38:66-71. 1955.
707. _____, _____. Evaluation of a short column water elutriator. *Jour. Amer. Ceramic Soc.* 34:109-114. 1955.
708. _____, _____. Improved calcium fluoride shapes. *Ames. Lab. ISC-514.* 12 pp. 1954.
709. Wilhelm, H.A., O.N. Carlson, and J.M. Dickinson. Columbium-vanadium alloy system. *Trans. Amer. Inst. Met. Engrs.* 200:915-918. 1954.
- _____, joint author. See under Beyer.
710. Wilk, M.B. The randomization analysis of a generalized randomized block design. *Biometrika* 42:70-79. 1955.
711. Wilkowskie, H.H., F.E. Nelson, and C.E. Parmelee. Serological classification of bacteriophage active against lactic streptococci. *Appl. Microbiol.* 2:243-249. 1954.

712. _____, _____, _____. Heat inactivation of bacteriophage strains active against lactic streptococci. *Appl. Microbiol.* 2:250-253. 1954.
713. Williams, J.T. The vanadium-sirconium alloy system. *Jour. Metals* 7:345-350. 1955.
714. Willis, E. and P. Swanson. Fruits and vegetables. Keys to vitamins A and C. *Iowa Farm Sci.* 9:742-744. 1955.
715. _____, _____, R. Angus, and A. Pohm. Iowa women: The kinds of food they eat. *Iowa Farm Sci.* 9:631-633. 1954.
716. _____, _____, P. Mairs, and H. Roberts. Iowa women contributors to science. *Iowa Farm Sci.* 9:600-602. 1954.
_____, joint author. See under Mairs, Swanson.
717. Wilsie, C.P. Relative efficiency of lattice and randomized block designs for forage crop trials. *Agron. Jour.* 46:355-357. 1954.
718. _____. Legumes for tomorrow. *Rept. State Apiarist for 1954.* pp.21-25. 1955.
719. _____ and R.H. Shaw. Crop adaptation and climate. pp.199-252 in *Advances in Agronomy.* Academic Press. 1954.
_____, joint author. See under Brigham.
720. Wilson, G.B. and C.C. Bowen. The study of mitotic poisons. *Huitieme Congres Internatl. de Botanique, Rapports et Communications, Paris, 1954.* Sect. 9 et 10. pp.31-34. 1954.
721. Winders, G.R., E.H. Dewell, R.G. Clark, M.D. Voss, and K.L. Malaby. Equipment for processing reactor fuels. *Nucleonics* 12:50-51. 1954.
_____, joint author. See under Fisher.
Winter, A.R., joint author. See under Cotterill, Funk.
Winters, L.M., joint author. See under Dickerson.
Woods, J.D., joint author. See under Brown, F.E.
Woods, L.A., joint author. See under Gilman.
722. Yoder, Lester, Edmund Cheng, and Wise Burroughs. Synthesis of estrogenic isoflavone derivatives. *Proc. Iowa Acad. Sci. for 1954* 61:271-277. 1954.
_____, joint author. See under Cheng.
723. York, George T. Present status of parasites of the European corn borer. *Proc. 10th Ann. Meet. North Central Branch, Ent. Soc. Amer.* 10:65. 1955.
Ystgaard, O.M., joint author. See under Bird.

Zaffarano, D.J., joint author. See under Bureau, Kline, R.M.
724. Zimmermann, W.J., L.H. Schwarte, and H.E. Biester. Incidence of trichiniasis in pork products. *North Amer. Vet.* 36:367-370. 1955.
_____, joint author. See under Becker.
725. Zoellner, J.A. and Oscar Kempthorne. Incomplete block designs with blocks of two plots. *Iowa Agric. Expt. Sta. Res. Bull.* 418. 12 pp. 1954.
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